Original Research

A Novel Role for Type 1 Angiotensin Receptors on T Lymphocytes to Limit Target Organ Damage in Hypertension

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Rationale: Human clinical trials using type 1 angiotensin (AT1) receptor antagonists indicate that angiotensin II is a critical mediator of cardiovascular and renal disease. However, recent studies have suggested that individual tissue pools of AT1 receptors may have divergent effects on target organ damage in hypertension.

Objective: We examined the role of AT1 receptors on T lymphocytes in the pathogenesis of hypertension and its complications.

Methods and Results: Deficiency of AT1 receptors on T cells potentiated kidney injury during hypertension with exaggerated renal expression of chemokines and enhanced accumulation of T cells in the kidney. Kidneys and purified CD4+ T cells from “T cell knockout” mice lacking AT1 receptors on T lymphocytes had augmented expression of Th1-associated cytokines including interferon-γ and tumor necrosis factor-α. Within T lymphocytes, the transcription factors T-bet and GATA-3 promote differentiation toward the Th1 and Th2 lineages, respectively, and AT1 receptor-deficient CD4+ T cells had enhanced T-bet/GATA-3 expression ratios favoring induction of the Th1 response. Inversely, mice that were unable to mount a Th1 response due to T-bet deficiency were protected from kidney injury in our hypertension model.

Conclusions: The current studies identify an unexpected role for AT1 receptors on T lymphocytes to protect the kidney in the setting of hypertension by favorably modulating CD4+ T helper cell differentiation. (Circ Res. 2012;110:00-00.)

Key Words: hypertension ■ kidney disease ■ T lymphocytes ■ inflammation

Chronic persistent hypertension is widely prevalent and leads to several catastrophic cardiovascular complications including stroke, congestive heart failure, and chronic kidney disease. The remarkable efficacy of type 1 angiotensin (AT1) receptor blockers in lowering blood pressure suggests that in hypertensive patients the renin-angiotensin system is broadly activated via AT1 receptor stimulation. Clinical trials have shown that pressure reduction during treatment with angiotensin receptor blockers slows but does not halt or reverse the progression of hypertensive kidney injury. A more precise understanding of AT1 receptor functions in hypertension will therefore be critical to the optimization of treatment strategies for chronic kidney disease.

Several studies have suggested that the regulation of immune responses by angiotensin (Ang) II during hypertension may alter the pattern and severity of associated kidney damage. For example, broad immunosuppression mitigates renal injury during Ang II-dependent hypertension, and lymphocyte deficiency during Ang II infusion blunts blood pressure elevation and target organ damage, suggesting that Ang II can potentiate kidney injury by stimulating proinflammatory signals. However, these earlier studies did not determine whether Ang II shapes inflammatory responses through direct activation of AT1 receptors on immune cells or other intermediary cell lineages.

More recent studies have explored the role of AT1 receptors on immune cells in modulating target organ damage by using AT1 receptor-deficient bone marrow chimeras. These experiments have yielded conflicting results. However, several confounding elements of the bone marrow transfer model may complicate the interpretation of these results. For example, the lethal irradiation required for bone marrow transplantation may itself alter gene expression programs.
in the recipient’s heart and kidney thereby influencing the injury response,\(^1\) and repopulation of the recipient’s bone marrow with transplanted donor cells may not reliably recapitulate normal development of the endogenous immune system.\(^1\) These limitations highlight the need for a novel genetic approach to examine functions of AT\(_1\) receptors on immune cell lineages.

K T lymphocytes are known to express AT\(_1\) receptors\(^\text{13}\) and infiltrate the kidney in the setting of hypertension.\(^4\) Therefore, to precisely determine the role of AT\(_1\) receptors on T lymphocytes in regulating blood pressure and target organ damage in hypertension we used a Cre-lox gene targeting strategy to selectively remove AT\(_{1\alpha}\) receptors from T lymphocytes. We find that AT\(_1\) receptors on T cells protect the kidney in hypertension and that activation of AT\(_1\) receptors on T cells in this setting suppresses their differentiation toward the proinflammatory Th1 phenotype. We further confirm that this proinflammatory Th1 immune response mediates hypertensive kidney damage by showing that T-bet-deficient mice lacking Th1 responses are protected from kidney injury in our hypertension model.

**Methods**

An expanded Methods section is provided in the online-only Data Supplement.

**Mice**

C57BL/6 \(\text{Agtr1}^{\text{lox/lox}}\) mice were previously generated\(^\text{14}\) and were crossed in 2 steps of breeding with mice harboring Cre recombinase under the control of the CD4 promoter\(^\text{15}\) to generate CD4 \(\text{Cre}^+\) \(\text{Agtr1}^{\text{lox/lox}}\) (T cell knockout [KO]) mice. To increase the susceptibility to kidney injury, T cell KO mice from the C57BL/6 strain were crossed with \(\text{Agtr1}^{\text{lox/lox}}\) mice from the 129/SvEv strain to yield the [C57BL/6×129/SvEv]\(_\text{F1}\) \(\text{T cell KO and T cell wild type (WT)}\) (CD4 \(\text{Cre}^+\) \(\text{Agtr1}^{\text{lox/lox}}\)) littermates for our experiments. To confirm selective CD4 \(\text{Cre}\) expression in lymphoid tissues, \(\text{mTmG}\) mice from Jackson Laboratory were crossed with the CD4 \(\text{Cre}\) recombinase transgenic lines. \(\text{mTmG}\) mice normally express red fluorescence protein in all tissues. When \(\text{Cre}\) is present, the \(\text{mT}\) cassette is deleted, triggering expression of the membrane-targeted \(\text{EGFP}\).\(^\text{16}\) T-bet\(^-/-\) mice on the C57BL/6 background were purchased from Jackson Laboratory and backcrossed to the 129/SvEv strain for 6 generations to increase susceptibility to kidney damage. Then, T-bet heterozygotes were intercrossed to yield the T-bet KO and WT littermates for our experiments. Further details are included in the online-only Data Supplement.

**Model of Angiotensin II-Dependent Hypertension**

Experimental animals underwent left nephrectomy followed 1 week later by implantation of a pressure-sensing catheter via the left common carotid artery as previously described.\(^\text{17}\) After allowing 7 days for reestablishment of diurnal blood pressure variation, blood pressure measurements were recorded at baseline and during 4 weeks of chronic Ang II or saline infusion as detailed in the Online Supplement.

**Histological Analysis**

At the end of the 28-day experimental period, the heart and kidney were harvested and weighed. Portions of kidney were fixed and analyzed for renal damage and inflammation was performed as outlined in the online-only Data Supplement. To stain glomerular podocytes on kidney sections, we used a WT1 antibody (Santa Cruz) at a 1:400 dilution per a previously established method.\(^\text{18}\) WT1-positive podocytes in each glomerular cross-section were then enumerated with a blinded approach, scoring 15 to 20 glomeruli per mouse.

**Evaluation of Lymphocyte Populations**

At the conclusion of the Ang II infusion protocol, the spleen, thymus, and kidney were collected for analysis. Single-cell suspensions of splenocytes, thymocytes, and kidney cells were prepared, and flow cytometry was performed as described previously.\(^\text{19,20}\) The antibodies used and cell sorting procedures are detailed in the online-only Data Supplement.

**Isolation of RNA and Realtime PCR**

Total RNA was isolated from individual cells or tissues by using the RNeasy Mini Kit according to manufacturer’s instructions. RNA expression levels were then determined for \(\text{Agtr1}a\), interferon-\(\gamma\) (IFN-\(\gamma\)), tumor necrosis factor-alpha (TNF-\(\alpha\)), transforming growth factor-beta (TGF-\(\beta\)), interleukin-1beta (IL-1\(\beta\)), interleukin-4 (IL-4), monocyte chemoattractant protein 1 (MCP-1), CCL5 (regulated on activation, normal T expressed and secreted), neutrophil gelatinase-associated lipocalin (NGAL), CD4, CD8, T-bet, and GATA-3 by real-time PCR as detailed in the online-only Data Supplement.

**Cytokine ELISA**

Spleenic lymphocytes were obtained from both T cell KO and WT mice at the end of the Ang II infusion protocol as above. Cytokines were quantified in the media after the stated time of coculture of the lymphocytes with anti-CD3e (NA/LE, BD) or isotype antibody. ELISA assays targeting IFN-\(\gamma\), TNF-\(\alpha\), and IL-4 were performed with specific kits from Invitrogen according to manufacturer’s instructions.

**Statistics**

The values of each parameter within a group are expressed as the mean±SEM. Please see online-only Data Supplement for further details of statistical analysis.

**Results**

**Generation of Mice With Specific Deletion of the AT\(_{1\alpha}\) Receptor on T Cells**

Consistent with previous reports,\(^\text{11}\) we detected robust mRNA expression for the \(\text{Agtr1}a\) gene encoding the AT\(_{1\alpha}\) receptor in T lymphocytes, albeit at lower levels than in kidney and heart (Online Figure I). Therefore, to examine the functions of AT\(_1\) receptors on T cells in hypertension, we removed AT\(_{1\alpha}\) receptor-mediated responses from T lymphocytes by using a Cre-lox gene targeting approach. During the double positive phase of maturation in the thymus, all thymocytes destined to become single positive CD4\(^+\) or CD8\(^+\) circulating T lymphocytes express the CD4 promoter. Accordingly, by breed-
ing a CD4-Cre mouse line15 with a double fluorescence reporter mouse (mT/mG) we confirmed robust Cre expression marked by GFP within the thymus and spleen of CD4-Cre+ mT/mG mice but the absence of Cre expression marked by red fluorescent protein in nonlymphoid tissues such as the kidney (Figure 1A). We therefore bred the CD4-Cre mouse line with an Agtr1a flox line harboringloxP sites on either side of the coding region for the AT1A receptor. For our experiments, we used CD4-Cre+ Agtr1alox/flox mice (T cell KO) and CD4 Cre- Agtr1alox/flox (T cell WT) littermates. To confirm T cell-specific deletion of AT1A receptors in our T cell KO animals, we labeled splenocytes for T and B lymphocyte markers and isolated CD4+ T cells (CD4+CD8−CD19−), CD8+ T cells (CD8+CD4+CD19−), and B lymphocytes (CD19+Thy1−) via fluorescent cell sorting (Online Figure II). Using RNA from these immune cells and from kidney and heart, we then performed real-time PCR for the Agtr1a gene to confirm the degree and precision of T cell-specific deletion. Compared with their T cell WT littermates, T cell KO mice exhibited 90% deletion of the AT1A receptor from both CD4+ and CD8+ T cells but preserved AT1A receptor expression in all other tissues examined (Figure 1B). Within the T cell WT group, we detected similar levels of AT1A receptor expression on most circulating T cell subsets including CD4+, CD8+, T regulatory, and NK T cells. However, levels of Agtr1a mRNA were roughly 50% lower in the double negative CD3−CD4−CD8− T cells (Figure 1C).

**AT1A Receptors on T Cells Do Not Regulate Blood Pressure**

Following verification of AT1A receptor deletion in the T cell KO animals, mice underwent unilateral nephrectomy to render the remaining kidney more susceptible to hypertensive damage. After uninephrectomy, baseline blood pressures were measured by radiotelemetry in the experimental groups (Figure 2A). Baseline blood pressures in the T cell WT and T cell KO mice were virtually identical. Following measurement of baseline blood pressures, an osmotic minipump was implanted subcutaneously to chronically infuse Ang II as a robust model of hypertension with accompanying inflammatory kidney damage.4,21 After the initiation of Ang II infusion, blood pressures in the T cell WT group rose 30 mm Hg and remained elevated throughout the infusion period (Figure 2A). During Ang II infusion, blood pressures measured in the T cell KO group remained similar to those of their WT littermates (Figure 2A), indicating that AT1 receptors on T lymphocytes do not regulate the chronic hypertensive response to Ang II.

Cardiac hypertrophy is a frequent complication of persistent hypertension and typically correlates closely with the degree of blood pressure elevation.22 We therefore examined cardiac hypertrophy in the T cell WT and KO groups by measuring heart weight to body weight ratios after 4 weeks of saline or Ang II infusion (Online Figure III). Compared to saline, Ang II caused 35% increase in heart weights in the T cell WT (P=0.0002) and T cell KO mice (P<0.0001). However, there was no difference between these groups in cardiac enlargement during Ang II-induced hypertension. The degree of pathological injury in the hearts of our experimental
animals was quite mild with T cell infiltration that was marginally increased following Ang II compared to saline (Online Figure IV). The hearts from the Ang II-infused T cell KO mice contained numerically but not significantly greater T cell numbers than their T cell WT counterparts, matched by a trend toward enhanced expression of the T cell chemokine CCL5 in the T cell KO group (Online Figure V). Macrophage infiltration into the heart was sparse and similar with saline and Ang II infusion (0.110212 macrophages per heart cross-section in all groups).

AT1A Receptor-Deficiency on T Cells Exacerbates Kidney Injury in Ang II-Induced Hypertension Interfering with T lymphocyte functions in the setting of hypertension reduces associated kidney damage.4,23 Therefore, following 4 weeks of saline or Ang II infusion, we quantitated urinary albumin excretion in the experimental groups as a marker for kidney injury (Figure 2B). Levels of albuminuria in the saline-infused T cell WT and KO mice were mild and identical. Ang II infusion was associated with a dramatic increase in urinary albumin excretion in the T cell WT and T cell KO groups. However, despite the similar blood pressures in these 2 groups, the Ang II-infused T cell KO mice had nearly 40% more urinary albumin excretion than the Ang II-infused T cell WTs (P<0.0009). Within the kidney glomerulus, the podocyte is a key component of the filtration barrier such that podocyte loss leads to exaggerated urinary albumin excretion. We therefore stained the T cell WT and KO kidneys for the podocyte marker WT1 and counted the number of podocytes per glomerulus in each group (Figure 2C–2E). Following Ang II, staining for podocytes was significantly reduced in the T cell KO kidneys compared to T cell WT controls (P<0.02). By contrast, semiquantitative scoring of renal tubular damage revealed no epithelial reaction in the saline- or Ang II-infused T cell WT or T cell KO mice (0.1100060.18 versus 0.47±0.17 arbitrary units; P=NS). Moreover, renal function as measured by serum creatinines deteriorated only mildly with Ang II infusion and to a similar extent in the T cell WT and KO animals (Online Figure VI).

NGAL is another sensitive marker of progressive kidney injury.24 We therefore measured NGAL mRNA expression in the kidneys from our T cell WT and KO groups following saline or Ang II infusion (Figure 2F). Renal expression of NGAL was similar in the saline-infused T cell WT and KO mice. Induction of hypertension with Ang II resulted in a nearly 2-fold increment in NGAL expression in T cell WT

Figure 2. Activation of type 1a angiotensin (AT1a) receptors on T lymphocytes protects from hypertensive kidney injury. A, Baseline blood pressures were measured by radiotelemetry for 3 days. Then experimental mice were infused for 28 days with angiotensin II (1000 ng/kg/min) via subcutaneously implanted osmotic minipump. Mean arterial pressures are depicted for T cell wild type (WT) (n=11) and T cell knockout (KO) (n=12) littermates. B, Urine samples were collected by placing experimental mice into metabolic cages after 25 days of saline or angiotensin (Ang) II infusion. Urinary albumin excretion was quantitated per methods. #P<0.001 vs saline T cell KO; †P< 0.009 vs Ang II T cell WT. C–E, Staining of glomerular podocytes with WT1 antibody following 4 wk of Ang II. Podocytes stain bright green. Representative images of (C) T cell WT and (D) T cell KO glomeruli. (Magnification 40×), and E, number of podocytes per glomerulus in T cell WT and KO kidneys (6 mice per group). *P=0.02 vs T cell WT. F, Renal neutrophil gelatinase-associated lipocalin (NGAL) mRNA expression in saline- or Ang II-infused T cell WT or T cell KO mice. #P<0.02 vs saline T cell WT; *P=0.005 vs saline T cell KO; †P=0.001 vs Ang II T cell WT.
Figure 3. Role of the type 1a angiotensin (AT1A) receptor on T cells in regulating infiltration of mononuclear cells into the kidney during angiotensin (Ang) II-dependent hypertension. Embedded kidney sections were obtained from Ang II-infused T cell WT and KO mice and stained with anti-CD3 or anti-F4/80 antibody. A, B, representative images of T lymphocyte staining in kidneys from Ang II-infused (A) T cell WT mice and (B) T cell KO mice. C, proportion of renal vessels from Ang II-infused T cell WT and KO groups surrounded by 0 to 10, 11 to 20, or >20 CD3+ T cells, respectively. *<P<0.0001 vs T cell WT (Fisher’s exact test across all 3 tertiles). D, E, representative images of macrophage staining in kidneys from Ang II-infused (D) T cell WT and (E) T cell KO mice.

kidneys. However, Ang II achieved an even more profound upregulation in NGAL expression in the T cell KO kidneys (P=0.001 versus Ang II T cell WT), suggesting that AT1 receptors on T lymphocytes paradoxically constrain the kidney injury response in the setting of Ang II-dependent hypertension.

Regulation of Mononuclear Cell Accumulation in the Kidney by T Cell AT1A Receptors

T cells and macrophages accumulate in the kidney in several forms of hypertension,^4,23,25 and cytokines produced by these cells have the capacity to direct local inflammatory damage.26,27 Therefore, following Ang II infusion, the T cell WT and KO kidneys were stained for the immune cell markers CD3 for T lymphocytes and F4/80 for macrophages. Because nearly all of the T cells were clustered around vessels in the kidney, we used a published system^3 to quantitate the density of perivascular T cell infiltrates in the kidneys from the 2 groups (Figure 3A–3C). The proportion of dense perivascular clusters containing >20 CD3+ T cells was dramatically enhanced in the kidneys from the T cell KO group (37.5%) compared with the T cell WT controls (4.3%; P=0.004). To estimate the proportions of CD4+ and CD8+ T cells infiltrating the kidney during Ang II-dependent hypertension, we computed the ratios of CD4+ and CD8+ mRNA expression in the T cell WT and KO kidneys. Compared to the T cell WTs, the T cell KO kidneys had a markedly enhanced CD4/CD8 ratio (1.00±0.16 versus 2.89±0.50 arbitrary units (au); P=0.02). To more precisely enumerate the CD4+ and CD8+ T lymphocytes infiltrating the kidney, we isolated T lymphocytes directly from the kidneys of our Ang II-infused animals using fluorescent cell sorting (Online Figure VIII). Compared to T cell WT controls, the T cell KO kidneys contained increased numbers of CD4+ T cells (P<0.04), indicating that deficiency of AT1 receptors on CD4+ T cells favors their infiltration into the inflamed hypertensive kidney. By contrast, renal infiltration of CD8+ T cells,28 recently shown to be the prohypertensive T cell subset in the setting of Ang II infusion, was similar in the 2 groups. Macrophage staining of the kidneys following Ang II-induced hypertension revealed a similar trend toward enhanced macrophage accumulation in the T cell KO kidneys compared to their T cell WT counterparts (P=0.09; Figure 3D and 3E).

Chemokines play a critical role in recruiting inflammatory cells into the injured kidney, and their actions can have profound effects on progressive renal damage in hypertension.^29 Therefore, to explore possible mechanisms through which AT1A receptor deficiency on T cells during Ang II infusion leads to accumulation of mononuclear cells in the kidney, we examined renal expression of the prototypical T cell and macrophage chemokines CCL5 and MCP-1, respectively. Compared to saline, Ang II caused significant upregulation in CCL5 mRNA expression in the kidneys from T cell WT mice. However, this exaggeration in renal CCL5 expression with Ang II was even more profound in the T cell KO group (P=0.03 versus Ang II T cell WT; Figure 4A). With saline control, expression of MCP-1 was nearly identical in the T cell WT and KO kidneys. Similar to our CCL5 findings, Ang II infusion resulted in a >6-fold increase in MCP-1 mRNA expression in T cell WT kidneys, but the induction of MCP-1 by Ang II in the kidneys of the T cell KO cohort was 4 times greater than in the T cell WTs (P<0.02; Figure 4B). These data indicate that AT1A receptors on T lymphocytes constrain renal expression of CCL5 and MCP-1 and limit the accumulation of mononuclear cells in the kidney during Ang II-dependent hypertension.

AT1A Receptors on T Lymphocytes Suppress the Renal Inflammatory Response in Ang II-Dependent Hypertension

To determine whether the enhanced renal accumulation of T lymphocytes in the T cell KOs influenced the local generation of inflammatory mediators, we used RT-PCR to profile expression of several of these mediators in the kidney following Ang II infusion (Figure 4C). Renal expressions of IFN-γ and TNF-α were significantly upregulated in the T cell KO group compared to the T cell WTs (P≤0.04 for each). By contrast, expression of IL-6 was similar in the T cell WT and KO kidneys whereas IL-4 was not detectable in the kidneys from either group. The enhanced IFN-γ and TNF-α expressions in the T cell KO kidneys suggested the possible involvement of
proinflammatory T lymphocytes in the accelerated hypertensive kidney injury in this group. Thus, we next examined how AT1A receptors on T lymphocytes regulate their differentiation and function in the setting of hypertension.

Maturation of T Lymphocytes in Thymus Is Intact in T cell KO Mice

As the CD4 promoter is active relatively late during thymocyte differentiation in the so-called “double positive” phase, we anticipated that our model would not disrupt normal maturation of T cells in the thymus. Nevertheless, to characterize the maturation of T cells in the thymus of our experimental mice, we quantitated the proportions of CD3+/CD4+/CD8− double negative, CD3−CD4+/CD8+ double positive, and CD4− or CD8− single positive thymocytes using flow cytometry (Figure 5A and 5B). This analysis revealed similar proportions of each of these thymocyte populations in the T cell WT and KO groups, suggesting that thymic maturation was indeed intact in the T cell KO animals.

Ang II-Infused T Cell KO Animals Have Altered Proportions of T Lymphocytes in Spleen

Next, we examined the proportions of T lymphocytes in the spleen, a major secondary peripheral lymph and –6B). After saline infusion, proportions of CD4 and CD8 cells were similar in the T cell WT and KO groups (top panels of Figure 6A and 6B), indicating that homing of T lymphocytes to the spleen was intact in the T cell KO group. Following Ang II

Figure 4. Deficiency of type 1a angiotensin (AT1A) receptors on T lymphocytes permits enhanced renal expression of chemokines and inflammatory cytokines in angiotensin (Ang) II-dependent hypertension. A, CCL5 (RANTES) mRNA expression. †P<0.02 vs saline T cell WT; *P<0.05 vs saline T cell KO; #P=0.03 vs Ang II T cell WT. B, MCP-1 mRNA expression. †P=0.009 vs saline T cell WT; *P<0.009 vs saline T cell KO; #P<0.02 vs Ang II T cell WT. C, Expressions of interferon (IFN)-γ, tumor necrosis factor (TNF)-α, and interleukin (IL)-6 mRNA in kidneys from Ang II-infused T cell WT and KO mice. *P=0.04 vs T cell WT; †P<0.04 versus T cell WT.

Figure 5. T cell knockout (KO) mice have normal maturation of T lymphocytes in the thymus. Thymocytes were isolated from T cell wild type (WT) and KO animals and labeled for CD4 and CD8. A, Representative dot-plots of single positive CD4+, CD8+, double positive, and double negative T cell proportions in thymus from T cell WT and KO animals (n=4). B, Summary data for thymocyte proportions showing similar percentages of CD4+, CD8+, double positive (CD4+CD8+) and double negative (CD4−CD8−) thymocytes in T cell WT and KO animals.
infusion, the proportions of CD4\(^+\) T cells in the T cell KO spleens were slightly but significantly diminished compared to their T cell WT counterparts (\(P<0.006\)) (bottom panels of Figure 6A and 6B), consistent with enhanced retention of CD4\(^+\) T lymphocytes in the T cell KO kidney during Ang II-dependent hypertension.

AT\(_1\) receptors on CD4\(^+\) T cells could limit their accumulation in the damaged kidney by influencing their recruitment from lymphoid reservoirs such as the spleen and/or by altering the proliferative response of T cells in the setting of hypertensive kidney injury. In this regard, CD4\(^+\) T cells isolated from the T cell KO group had exaggerated expression of the T cell chemo-kine CCL5 compared to controls (1.50±0.18 versus 1.00±0.13 au; \(P=0.04\)), suggesting that enhanced chemokine production by infiltrating AT\(_1\) receptor-deficient T cells potentiated their accumulation in the T cell KO kidney. Consistent with this notion, we found considerably higher levels of intrinsic CCL5 expression in lymphocytes compared to whole kidney or freshly isolated renal tubular epithelial cells (Online Figure IX). To examine T cell proliferative responses, lymphocytes were isolated from experimental animals and labeled with CFSE. After restimulation with anti-CD3\(_e\) or vehicle control, CD4\(^+\) T cells were labeled, allowing the calculation of their proliferation indices (Online Figure XA–XC).\(^{30}\) No significant differences in proliferative capacity were evident between the CD4\(^+\) T lymphocytes from T cell WT and KO groups in these studies.

Enhanced Differentiation Toward the Pro-Inflammatory Th1 Phenotype in AT\(_{1\alpha}\) Receptor-Deficient T Cells In Vivo

On activation, CD4\(^+\) T lymphocytes have the capacity to differentiate into several distinct subsets including the Th1, Th2, Treg, and Th17 phenotypes. As the pattern of cytokine expression in the hypertensive kidneys from our T cell KO group suggested the possible involvement of proinflammatory Th1 cells, we next examined the impact of the AT\(_1\) receptor on the differentiation of CD4\(^+\) T lymphocytes. To this end, we used fluorescent cell-sorting to isolate CD4\(^+\) T

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Figure 6. T cell knockout (KO) mice have diminished proportions of CD4\(^+\) T lymphocytes residing in spleen during Ang II-dependent hypertension. T cell proportions in spleen from saline- and Ang II-infused T cell wild type (WT) and KO mice (n=6). A, Representative dot plots of CD4\(^+\) and CD8\(^+\) T cell subsets in spleen following saline or angiotensin (Ang) II infusion in T cell WT and KO groups. B, Summary data for CD3\(^+\) splenocyte distribution showing diminished proportions of CD4\(^+\) T cells with a reciprocal increase in CD8\(^+\) T cell proportions in the spleens of T cell KO animals following Ang II infusion. *\(P<0.006\) vs Ang II T cell WT; †\(P<0.03\) vs Ang II T cell WT.
cells from the spleen and kidney following 4 weeks of Ang II infusion and measured mRNA expression of the cytokines that mark these T helper subsets. As shown in Figure 7A, we were able to detect in splenic CD4+ T cells expression of IFN-\(\gamma\) and TNF-\(\alpha\) as markers for the Th1 subset, IL-4 as a marker for the Th2 subset, and TGF-\(\beta1\) as a marker for Treg cells. Similar to our findings in whole kidney, mRNA expressions of IFN-\(\gamma\) and TNF-\(\alpha\) were increased in the CD4+ T cells isolated from the T cell KO spleens (Figure 7A) and kidney (Online Figure XI) compared to T cell WT controls, confirming that the enhanced expression of these Th1-associated cytokines in the T cell KO kidneys was driven in part by increased production of these cytokines from the infiltrating T lymphocytes in this group. By contrast, CD4+ T cell expressions of IL-4, produced by Th2 cells, and TGF-\(\beta1\), produced by T regulatory cells, respectively, were similar in the T cell KO and WT groups following Ang II infusion (Figure 7A).

The enhanced expression of IFN-\(\gamma\) and TNF-\(\alpha\) in the CD4+ T cells from the T cell KO group indicated a propensity of these T cells toward a Th1 phenotype. Therefore, to confirm the differences seen in IFN-\(\gamma\) and TNF-\(\alpha\) production
at the protein level, we pursued an ELISA approach. Accordingly, we harvested splenic lymphocytes from our Ang II-infused T cell WT and KO groups, activated the T cells in the culture with anti-CD3ε, and assayed the culture supernatants for IFN-γ, TNF-α, and IL-4 (Figure 7B). Lymphocytes from the T cell KO animals had exaggerated production of IFN-γ (P<0.05) and TNF-α (P=0.005) compared to T cell WT controls. By contrast, IL-4 levels were similar in the culture supernatants from the T cell KO and WT groups. These data indicate that AT1 receptors on T lymphocytes suppress their production of Th1 cytokines, an effect that could be mediated through alterations in T cell differentiation and/or exhaustion.

Differentiation of CD4+ T lymphocytes into Th1 versus Th2 cells is reciprocally regulated by the T-bet and GATA-3 transcription factors, respectively. Therefore, the ratio of these factors within the T cell determines its propensity for commitment to the Th1 or Th2 lineages. Accordingly, we quantitated the ratio of T-bet/GATA-3 mRNA expression in the splenic CD4+ T cells from our T cell WT and KO groups. Following saline infusion, the T-bet/GATA-3 ratios were similar between the T cell WT and KO cohorts (Figure 7C). By contrast, following 4 weeks of in vivo Ang II infusion, purified CD4+ T lymphocytes from the T cell KO animals had augmented T-bet/GATA-3 ratios compared to T cell WT controls (P<0.05, Figure 7C). These changes in T cell differentiation were then confirmed in CD4+ T lymphocytes isolated directly from the kidneys of our Ang II-infused animals (Online Figure XI) with marked upregulation of T-bet (P=0.0003) and downregulation of GATA-3 (P<0.02) in the T cell KO group. Thus, in the setting of hypertension, AT1 receptor-deficiency in T lymphocytes triggers an imbalance in transcription factors within the T cell favoring Th1 differentiation and the accumulation of these proinflammatory Th1 cells within the kidney.

Recent evidence has emerged illustrating that, in the setting of chronic stimulation as might occur in our hypertension model, T cells may become exhausted and thereby secrete reduced amounts of cytokine. Exhausted T lymphocytes characteristically express higher surface levels of programmed death 1 (PD-1) coordinately with T cell immunoglobulin and mucin-domain-containing molecule 3 (Tim-3). Therefore, to evaluate whether a difference between our experimental groups in T cell exhaustion contributed to the disparate levels of IFN-γ and TNF-α produced by T lymphocytes in our model, we isolated CD4+ T cells from our Ang II-infused mice and labeled them for PD-1 and Tim-3 (Figure 7D). We found that concomitant cell surface expression of PD-1 and Tim-3 was similar on the CD4+ T lymphocytes from the T cell WT and KO groups (3.91±0.88 versus 2.91±0.82%; P=NS, Figure 7D). Thus, AT1 receptors on T lymphocytes do not appear to regulate their exhaustion via a PD-1 dependent pathway.

To gain further insights into how chronic stimulation of the AT1 receptor suppresses Th1 proinflammatory responses in T lymphocytes in our hypertension model, we harvested CD4+ T lymphocytes directly from the kidneys our Ang II-infused experimental animals and surveyed their expression of immunity-related cell signaling molecules via microarray analysis. The genes that were differentially expressed by >1.5-fold in the infiltrating T cells from the T cell WT and KO groups are listed in Online Table I. Markers that were significantly upregulated in the AT1A receptor-deficient T cells included STAT4, a transcription factor that along with T-bet directs CD4+ T cells toward the Th1 lineage, CD3 and CD28, involved in Signals 1 and 2 of T cell activation, respectively, and CXCR3, a chemokine receptor preferentially expressed on Th1 cells. By contrast, CD25, a marker of activated T cells but also T regulatory cells was numerically but not significantly downregulated on the AT1A receptor-deficient T lymphocytes.

Chronic Stimulation of AT1 Receptors on T Cells Suppresses Their Th1 Differentiation In Vitro

Previous reports indicated that AT1A receptor-deficient lymphocytes have impaired rather than exaggerated responses to acute stimuli. Therefore, to better understand how AT1 receptors on T cells shape the inflammatory response, we examined the functions of naïve lymphocytes during in vitro stimulation. Consistent with prior studies, mixed lymphocyte reactions to allogeneic stimuli demonstrated a blunted proliferative response in AT1A receptor-deficient lymphocytes (Online Figure XII). By contrast, after 24 hours of T cell receptor stimulation in an ELISPOT assay, the frequency of IFN-γ-producing responders was enhanced among AT1A receptor-deficient lymphocytes in a dose-dependent pattern (Online Figure XIII), consistent with our in vivo findings. Thus, given that lymphocytes harvested from our experimental T cell KO animals displayed normal proliferative responses (Online Figure XA–XC) and exaggerated Th1 cytokine production (Figure 7A and 7B), we hypothesized that the enhanced Th1 responses of T lymphocytes from our T cell KO animals arose from the chronic stimulation occurring in our hypertension model. We therefore subjected naïve T cell WT and KO lymphocytes to persistent T cell receptor stimulation for 7 days in vitro culture in the absence or presence of Ang II and measured their expression of IFN-γ and TNF-α (Figure 8). Similar to our in vivo findings, the persistently activated T cell KO lymphocytes had exaggerated expression of IFN-γ and TNF-α compared to activated T cell WT controls. In these cultures, concomitant exposure to Ang II suppressed IFN-γ and TNF-α expression in the T cell WT but not the T cell KO lymphocytes, indicating that chronic exposure to Ang II as occurs in our hypertension model suppresses Th1 cytokine production in T cells via an AT1 receptor-dependent pathway.

Pro-Inflammatory Th1 Immune Responses Mediate Hypertensive Kidney Damage

As our T cell KO animals had evidence of increased hypertensive kidney damage and enhanced Th1 immune responses, we hypothesized that the AT1 receptor on T lymphocytes constrains renal injury by blunting the Th1 differentiation of CD4+ T lymphocytes. Accordingly, to test the contribution of Th1 immune responses to the pathogenesis of hypertensive kidney damage, we subjected T-bet–deficient (T-bet KO) mice that are unable to mount a Th1 response and their littermate WT controls to our Ang II-dependent hypertension...
Activation of the renin angiotensin system through stimulation of type 1 angiotensin (AT1) receptors is a key regulator in blood pressure elevation and hypertensive end-organ damage. However, accumulating data have suggested that the actions of AT1 receptors in the setting of hypertension are tissue- and cell-specific.22 Blockade of inflammatory signaling cascades in the setting of Ang II-induced hypertension certainly protects target organs from injury,4,36 but whether Ang II activates the immune system during hypertension through stimulation of AT1 receptors directly on hematopoietic cells or through other intermediary cell lineages has not been clear from earlier studies. Although rodents have 2 AT1 receptor isoforms, AT1, is the dominant AT1 isoform in most murine tissues, and in our preliminary studies we were able to detect only this isoform on purified T lymphocytes. Moreover, in the current studies, we detected robust and similar AT1, expression on all T cell subpopulations with the exception of the double negative T cell subset that is prominent only during early thymocyte development. Therefore, to precisely examine the actions of AT1 receptors on mature T lymphocytes in the pathogenesis of hypertension, we ablated the AT1, receptor exclusively from T cells by a Cre-loxP gene targeting strategy and then subjected these mice and their control littermates to chronic Ang II infusion. Surprisingly, these studies reveal that activation of AT1 receptors directly on T lymphocytes limits kidney damage in hypertension and suppresses production of inflammatory mediators.

In our experiments, intraarterial blood pressures were measured in large numbers of conscious unrestrained animals using state-of-the-art radiotelemetry technology, and yet baseline blood pressures were similar between the T cell KO mice and their wild-type littermates. These data indicate that AT1 receptors on T cells do not regulate normal blood pressure homeostasis. These findings are consistent with our previous experiments using AT1 receptor deficient bone marrow chimeras.37 Moreover, in the current studies blood pressures remained similar in the T cell KO and WT groups in baseline blood pressure (Online Figure XIVA). Although both groups had a robust chronic hypertensive response to Ang II, the WT and Tbet KO animals maintained similar blood pressures throughout the Ang II infusion period (177±2 versus 174±2 mm Hg; P=NS; Online Figure XIVA), indicating that Th1 immune responses do not contribute to the regulation of blood pressure by Ang II. Despite these similar blood pressures, renal expression of the kidney injury marker NGAL was diminished by 55% in the Ang II-infused T-bet KO group compared to controls (0.45±0.041 versus 1.00±0.11 au; P<0.001). As evidence of reduced glomerular injury, following 4 weeks of Ang II infusion the T-bet KO mice had 40% less albuminuria than controls (15.3±2.7 versus 26.6±4.7 mg/mg Cr; P=0.05). Moreover, T-bet KO animals compared to WT controls had preserved podocyte numbers marked by WT1 staining in the kidney glomerulus following Ang II infusion (11.7±1.2 versus 9.1±1.3 podocytes/glomerulus; P<0.003; Online Figure XIVB–XIVD).

Within the renal interstitium, T-bet–deficiency led to diminished accumulation of F4/80+ macrophages (2.1±0.1 versus 3.1±0.2% of high-powered field; P=0.001; Online Figure XIVE and XIVF) and perivascular CD3+ T lymphocytes (58.6 versus 85.2% of vessels with >30 T cells; P<0.006 by χ²). Consistent with a muted Th1 response in the T-bet KO mice, renal expressions of IFN-γ and TNF-α were reduced by half in this group compared to WT controls following Ang II (Online Figure XIVG), whereas IL-4 was not detectable in the kidneys from either group. Collectively, these data indicate that the Th1 immune response mediates kidney damage in Ang II-dependent hypertension.

**Discussion**

Activation of the renin angiotensin system through stimulation of type 1 angiotensin (AT1) receptors is a key regulator in blood pressure elevation and hypertensive end-organ damage. However, accumulating data have suggested that the actions of AT1 receptors in the setting of hypertension are tissue- and cell-specific.22 Blockade of inflammatory signaling cascades in the setting of Ang II-induced hypertension certainly protects target organs from injury,4,36 but whether Ang II activates the immune system during hypertension through stimulation of AT1 receptors directly on hematopoietic cells or through other intermediary cell lineages has not been clear from earlier studies. Although rodents have 2 AT1 receptor isoforms, AT1, is the dominant AT1 isoform in most murine tissues, and in our preliminary studies we were able to detect only this isoform on purified T lymphocytes. Moreover, in the current studies, we detected robust and similar AT1, expression on all T cell subpopulations with the exception of the double negative T cell subset that is prominent only during early thymocyte development. Therefore, to precisely examine the actions of AT1 receptors on mature T lymphocytes in the pathogenesis of hypertension, we ablated the AT1, receptor exclusively from T cells by a Cre-loxP gene targeting strategy and then subjected these mice and their control littermates to chronic Ang II infusion. Surprisingly, these studies reveal that activation of AT1 receptors directly on T lymphocytes limits kidney damage in hypertension and suppresses production of inflammatory mediators.

In our experiments, intraarterial blood pressures were measured in large numbers of conscious unrestrained animals using state-of-the-art radiotelemetry technology, and yet baseline blood pressures were similar between the T cell KO mice and their wild-type littermates. These data indicate that AT1 receptors on T cells do not regulate normal blood pressure homeostasis. These findings are consistent with our previous experiments using AT1 receptor deficient bone marrow chimeras.37 Moreover, in the current studies blood pressures remained similar in the T cell KO mice and their littermate controls throughout several weeks of chronic Ang II infusion, suggesting that AT1 receptors on T cells do not regulate the chronic hypertensive response to Ang II. This finding contrasts with the results of our previously published experiments in which AT1 receptor-deficient bone marrow chimeras had an accentuated hypertensive response to Ang II.37 In that experiment, mice underwent lethal irradiation prior to bone marrow transfer, which may have altered injury responses in the kidney and/or compromised renal epithelial functions. Thus, we believe that the approach in our current study is more physiological than our bone marrow transfer strategy. Alternatively, the current results allow for the intriguing possibility that RAS activation on immune cell lineages other than T cells may account for the protective actions of AT1 receptors on bone marrow derived cells with regard to blood pressure elevation.38
The failure of AT1 receptors on T lymphocytes to influence the chronic hypertensive response to Ang II in the current studies also contrasts with the results from the Rag1-deficient model in which adoptive transfer of AT1 receptor-deficient T lymphocytes prior to Ang II infusion yielded a blunted chronic hypertensive response compared with transfer of WT lymphocytes. However, there are several important differences in the design of these 2 models. First, the Rag1-deficient animal develops without functional lymphocytes, whereas our experimental animals are immune-competent during development. Second, in the Rag1 model, lymphocytes were transferred 3 weeks before initiation of Ang II infusion, whereas in our model immune cell populations infiltrating target organs are endogenous. Finally, the mouse strains used in the 2 experiments were different as was the dose of Ang II used to induce hypertension. These factors may each have contributed to different patterns of lymphocyte trafficking and function in the Rag1 study versus the current model.

Infiltration of T lymphocytes into the kidney is associated with more severe kidney damage in several models of hypertension and blockade of lymphocyte functions protect the kidney from hypertensive damage. Urinary albumin excretion is the most widely used marker of kidney injury as albuminuria correlates closely with progression of kidney disease and even survival. Thus, to determine whether AT1 receptors on T cells influence the degree of renal injury in our hypertension model, we measured urinary albumin excretion following 4 weeks of Ang II infusion. We found that T cell KO animals had dramatically higher levels of albuminuria than controls in the setting of hypertension. Moreover, the T cell KO kidneys showed significantly reduced WT1 staining for podocytes, the cells within the kidney glomerulus that prevent loss of albumin in the urine, indicating that AT1 receptor activation on T lymphocytes limits glomerular damage in hypertension and thereby mitigates albuminuria. Multiple lines of evidence have suggested that NGAL can serve as a sensitive marker of both acute and chronic kidney injury. We therefore measured kidney NGAL expression as another important marker for renal injury in our experiment. Consistent with the albuminuria findings, T cell KO animals had a far more profound elevation of renal NGAL expression than controls following Ang II infusion. Thus, the podocyte, exagerated albuminuria, and augmented NGAL expression indicate that deficiency of AT1 receptors specifically on T lymphocytes permits exaggerated kidney disease during hypertension.

Interestingly, AT1 receptor deficiency on T cells worsened the level of renal injury in our model without causing an exaggerated increase in blood pressure during Ang II infusion. Although an increase in kidney damage could potentially aggravate hypertension by interfering with renal sodium handling, there are several reports in which modulating immune responses during hypertension influenced the extent of kidney injury without impacting blood pressure elevation. In the current experiments, the level of CD8 T cell infiltration into the kidneys of our Ang II-infused T cell WT and KO animals was similar (discussed below), and recent data have emerged indicating that CD8 rather than CD4 T lymphocytes are the prohypertensive T cell subset influencing blood pressure during Ang II infusion. Moreover, the level of kidney damage in our model was quite mild relative to that seen in the pure 129/SvEv mouse and spared the renal tubular epithelium responsible for sodium trafficking in the kidney. Accordingly, the increases in serum creatinine were modest with Ang II and were similar in our T cell WT and KO mice such that the level of hypertension was not altered despite their differences in glomerular injury.

As mononuclear cell accumulation is a predictive marker for the progression of chronic kidney disease including renal damage induced by Ang II-dependent hypertension, we examined accumulation of T lymphocytes and macrophages in the kidney in our hypertension model. We found that activation of AT1 receptors on T cells limits the accumulation of T cells around blood vessels in the kidney following chronic Ang II infusion. To assess the proportions of CD4 T and CD8 T cells infiltrating the kidney in our experimental groups, we measured expression of CD4 and CD8 from kidney samples and directly enumerated fluorescently sorted T cells in digested kidney specimens. These approaches revealed an exaggerated aggregation of CD4 T lymphocytes in T cell KO kidneys compared to controls. We detected a similar pattern in the heart. However, the level of T cell infiltration into the heart was minimal such that no statistical differences between the T cell WT and KO hearts emerged. Thus, activation of AT1 receptors directly on T lymphocytes suppresses the accumulation of CD4 T cells in the inflamed kidney during hypertension, leading us to explore how AT1 receptors specifically on T lymphocytes influence T cell differentiation and infiltration into the target organ.

There are several potential mechanisms that could underlie the enhanced accumulation of CD4 T cells in the injured hypertensive kidney. One possibility is that deletion of AT1 receptors on T cells impacts their maturation in the thymus leading to increased production of single positive CD4 thymocytes at the expense of the CD8, double positive, and/or double negative thymocyte lineages. Our analysis of isolated thymocytes indicates that thymic maturation is intact in T cells lacking AT1 receptors (Figure 5). A second possibility is that AT1 receptor-deficiency on T cells alters the distribution of circulating T cells in the setting of Ang II-dependent hypertension. By labeling splenic lymphocytes, we found a slight but significant decrease in the proportion of CD4 T cells in the spleens of the Ang II-infused T cell KO animals (Figure 6A and 6B, bottom panels), suggesting that more CD4 T cells remain in the kidney in this group. We speculate that this difference in proportion of CD4 T cells was quite small because an antigen-driven response would recruit only a select population of T cells based on specificity of the T cell receptor. Regarding mechanisms of T cell recruitment, we detected enhanced chemokine levels within the kidney and in circulating T lymphocytes from the T cell KO animals. Thus, augmented CCL5 production by the CD4 T cells in the T cell KO group likely contributes to their enhanced renal accumulation through a paracrine mechanism. Finally, we considered the possibility that activated CD4 T cells lacking the AT1 receptor could have an exaggerated proliferative response in our model, leading to the higher proportions of these cells within the kidney. However, using
the CFSE assay, we demonstrated a similar proliferative response to T cell receptor activation regardless of the absence or presence of the AT₁ receptor on the T lymphocyte. Taken together, our data indicate that AT₁ receptors on T cells regulate their trafficking into the injured kidney in the setting of hypertension by influencing their generation of chemokines.

CD4⁺ T lymphocytes can be categorized into several distinct functional subsets including the Th1, Th2, Treg, and Th17 subtypes, and these T helper subsets appear to play diverse roles in immunologically mediated kidney diseases. By contrast, the roles of these T cell lineages in mediating kidney damage during hypertension are less well understood. On naïve T cells, AT₁ receptor-deficiency impairs proliferative responses to allogeneic stimuli. Moreover, we have previously found that Ang II infusion promotes infiltration of CD4⁺ T cells into the kidney coupled with renal expression of Th1 cytokines. Thus, we were surprised to find in the current study that ablation of AT₁ receptors selectively from T cells enhanced their expression of Th1 cell-associated molecules such as IFN-γ and TNF-α in the setting of persistent hypertension. Therefore, to confirm our in vivo findings in vitro, we subjected T cell WT and KO lymphocytes to chronic T cell receptor stimulation in culture and found augmented Th1 cytokine expression in T cell KO lymphocytes consistent with our in vivo findings. Furthermore, Ang II suppressed these cytokines only in the T cell WT lymphocyte culture. Thus, although AT₁ receptors on T cells potentiate the acute allogeneic immune response thereby promoting acute allograft rejection, we now find that persistent activation of AT₁ receptors directly on the T lymphocyte serves a protective role by limiting Th1 cell-mediated chronic inflammation. We therefore explored potential mechanisms through which AT₁ receptors on T cells suppress the Th1 lineage during hypertension.

T-bet is a key transcription factor that is required for the differentiation of T lymphocytes into the proinflammatory Th1 cell lineage, and the ratio of T-bet to GATA-3 within the T helper cell governs its development toward the Th1 versus Th2 lineage, respectively. In the current model, we find elevated ratios of T-bet versus GATA-3 expression in the CD4⁺ T lymphocytes from our Ang II-infused T cell KO mice but similar T-bet/GATA-3 ratios in T cell WT and KO CD4 cells following saline infusion, supporting the notion that chronic activation of AT₁ receptors on T lymphocytes constrains their differentiation toward the Th1 subset in our model. As the regulation of T-bet within the T cell is poorly understood, this finding uncovers a novel mechanism underlying the selection of T helper cell phenotypes during chronic kidney damage.

Secondly, we considered the possibility that the initial blunted response of naïve AT₁ receptor-deficient T cells to acute activation insulates them from the exhaustion that arises in T cells subjected to persistent activation, allowing the T cell KO lymphocytes to generate high levels of cytokine even after chronic stimulation in vivo. However, we were not able to confirm that the AT₁ receptor on T cells influences their susceptibility to exhaustion in our chronic disease model, at least via a PD-1-dependent pathway. Thirdly, we have previously demonstrated that AT₁ receptors on T cells are required to drive cytoskeletal rearrangements necessary to recruit T cell signaling molecules into the immunologic synapse, so we postulated that expression of signaling molecules might be dysregulated in the T lymphocytes from our Ang II-infused T cell KO animals. Indeed, microarray analysis revealed abnormal expression of key signaling molecules in T cell KO CD4⁺ cells including another Th1 transcription factor STAT4 but also enhanced expression of CXCR3, a receptor on Th1 cells that guides their infiltration into damaged tissues. Although these data will require confirmation and further exploration in future studies, they suggest that, in conjunction with the increased CCL5 expression mentioned above, enhanced CXCR3 expression on T cells lacking AT₁ receptors facilitates accumulation of proinflammatory Th1 cells in the kidney during hypertension.

As AT₁ receptor-deficiency on T cells leads to an enhanced Th1 immune response and exaggerated hypertensive kidney damage, we sought to determine directly whether the Th1 response contributes to renal injury in the setting of blood pressure elevation. We therefore subjected T-bet-deficient mice that cannot mount a Th1 response to our hypertension model and measured parameters of renal damage. We find that the Th1 response does not influence the chronic hypertensive response to Ang II. As global T cell deficiency causes a muted hypertensive response in other studies, our data suggest that actions of T lymphocyte populations other than CD4⁺ Th1 cells, such as CD8⁺ T cells, influenced blood pressure levels in those experiments. Nevertheless, in the current studies T-bet deficiency affords significant protection to the kidney, particularly within the glomerular compartment. Collectively, the Th1 response is known to shape the injury response in immune-mediated crescentic glomerulonephritis. To our knowledge, however, the current experiments are the first to indicate that the Th1 immune response, potentiated by AT₁ receptor-deficiency on T cells, plays a major role in driving renal injury in hypertension.

One direct consequence of inflammatory cell infiltration into the kidney is the local generation of cytokines, which may in turn influence the course and pattern of kidney damage. Accordingly, we found that IFN-γ and TNF-α were upregulated in the kidneys from the T cell KO group manifesting an exaggerated Th1 response and conversely downregulated in the kidneys from T-bet-deficient mice lacking a Th1 response. IFN-γ and TNF-α are known to directly mediate kidney injury, particularly to the kidney glomerulus, in several models. We therefore submit that the mononuclear cells infiltrating the kidney in our model similarly contribute to renal injury via the direct actions of inflammatory cytokines on kidney cells independently of any effects these cytokines may have on blood pressure in other models, particularly as the exaggeration of glomerular damage in the T cell KO kidneys and the protection from hypertensive renal injury in our T-bet KO mice was associated with a preserved hypertensive response to Ang II.

In summary, the current experiments demonstrate a paradoxical role for AT₁ receptors on T lymphocytes to protect the kidney in the setting of chronic hypertension by constraining their propensity to propagate the proinflammatory Th1
response. We further show that this Th1 response directly mediates hypertensive renal damage. Thus, the actions of AT1 receptors on T cells to limit injury during hypertension may represent a negative feedback mechanism counteracting the broad prohypertensive and proinflammatory effects of global RAS activation. The wide use of AT1 receptor blockers in the setting of cardiovascular and renal disease makes our findings widely relevant to clinical medicine. Blocking AT1 receptor signals directly within the kidney clearly ameliorates hypertension and the resulting injury to the heart and kidney.22,49

However, the current studies suggest that preserving or even stimulating AT1 receptor signaling pathways in T lymphocytes could provide important additional protection for the kidney during hypertension, and thus uncovers the potential for a novel therapeutic maneuver to suppress the renin angiotensin system in renal and cardiovascular tissues while concomitantly preserving or even enhancing AT1 receptor-mediated signals within the immune system. Such an approach may yield even greater benefits in hypertension than the current strategy of globalized angiotensin receptor blockade.

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Disclosures
None.

References
Novelty and Significance

Despite evidence from large-scale clinical trials showing that global blockade of AT₁ receptors ameliorates target organ damage in hypertension, the precise contribution of AT₁ receptors on T lymphocytes to the pathogenesis of hypertension remains unclear. In the current study, we found that deleting AT₁ receptors selectively from T cells in mice (T cell knockout [KO]) does not alter the chronic hypertensive response to angiotensin II infusion. However, T cell KO mice display exaggerated kidney injury in angiotensin-induced hypertension, with enhanced infiltration of T lymphocytes into the kidney. Moreover, AT₁ receptor-deficient T cells infiltrating the kidney manifest exaggerated proinflammatory Th1 immune responses, which directly mediate damage to the kidney glomerulus. These findings reveal a paradoxical role for AT₁ receptors of T lymphocytes in limiting kidney damage during chronic hypertension by constraining the propensity of T cells to propagate Th1 immune responses. Thus, preserving or even stimulating AT₁ receptor-mediated responses of T lymphocytes might protect against renal injury during chronic hypertension.
A Novel Role for Type 1 Angiotensin Receptors on T Lymphocytes to Limit Target Organ Damage in Hypertension

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Supplemental Material

Detailed Methods

Mice
C57BL/6 Agtr1a<sup>flox/flox</sup> mice were previously generated<sup>1</sup> and were crossed in 2 steps of breeding with mice harboring Cre recombinase under the control of the CD4 promoter<sup>2</sup> to generate CD4 Cre<sup>+</sup> Agtr1a<sup>flox/flox</sup> (T cell KO) mice. To increase the susceptibility to kidney injury, T cell KO mice from the C57Bl/6 strain were crossed with Agtr1a<sup>flox/flox</sup> mice from the 129/SvEv strain to yield the [C57BL/6 x 129/SvEv]<sub>F<sub>1</sub></sub> T cell KO and T cell WT (CD4 Cre<sup>+</sup> Agtr1a<sup>flox/flox</sup>) littermates for our experiments. To confirm selective CD4 Cre expression in lymphoid tissues, membrane-targeted tdTomato (mT)/membrane-targeted EGFP (mG) mice with loxP sites flanking the membrane-targeted dtTomato cassette followed by an N-terminal membrane-tagged version of EGFP from the Jackson Laboratory were crossed with the CD4 Cre recombinase transgenic lines. mT/mG mice normally express red fluorescence protein in all tissues. When Cre is present, the mT cassette is deleted, triggering expression of the membrane-targeted EGFP.<sup>3</sup> T-bet<sup>−/−</sup> mice on the C57/BL6 background were purchased from Jackson Laboratory and backcrossed to the 129/SvEv strain for 6 generations to increase susceptibility to kidney damage. Then, T-bet heterozygotes were intercrossed to yield the T-bet KO and WT littermates for our experiments. Mice were bred and maintained in the Association for Assessment and Accreditation of Laboratory Animal Care–accredited animal facilities at the Durham VA Medical Center according to National Institutes of Health guidelines. All of the animal studies were approved by the Durham Veterans’ Affairs Medical Center Institutional Animal Care and Use Committee and conducted in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals. Animals had free access to standard rodent chow and water. Eight- to 12-week-old male mice and littermate controls were used for experiments.

Model of angiotensin II-dependent hypertension
Experimental animals underwent left nephrectomy followed 1 week later by implantation of a pressure-sensing catheter (TA11PA-C10, Transoma Medical) via the left common carotid artery as described.<sup>4</sup> After allowing 7 days for reestablishment of diurnal blood pressure variation, baseline blood pressure measurements were recorded for 3 days by radiotelemetry (Transoma) in conscious unrestrained animals. Then an osmotic mini-pump (ALZET model, 2004) was implanted subcutaneously to infuse Ang II (1000ng/kg/min; Sigma; n≥11 mice per group) or saline (n=4 or 5 mice per group) continuously for 28 days. Blood pressure measurements continued for 3 weeks of saline or Ang II infusion. On day 25, the mice were placed into metabolic cages, and urines were collected for 24 hours. Urinary concentration of albumin were measured in individual samples using specific ELISA kits for mouse albumin (Exocell) as previously described.<sup>5</sup> Urine creatinine concentrations were measured with a picric acid-based method using a kit (Exocell).

Histological Analysis
At the end of the 28-day experimental period, blood samples were collected, and the heart and kidney were harvested and weighed. Weighing the hearts provides separate confirmation that chronic Ang II-infusion has increased the blood pressure in these animals. Serum creatinines were quantitated by the Animal Clinical Chemistry and Gene Expression Laboratory at University of North Carolina, Chapel Hill, using a colorimetric assay kit. Portions of kidney were fixed and pathological analysis was performed as we have described previously.<sup>6</sup> Briefly, to assess macrophage and T-lymphocyte infiltration in the kidneys, sections were stained with anti-
Harvest of mouse kidney tubular cells
In order to measure gene expression for CCL5 in kidney tubular cells, primary mouse renal tubular cells were obtained and cultured by following a previously described method. Briefly, mice were anesthetized and flushed with 5 mL ice-cold HBSS. Renal cortices were dissected visually and sliced into small pieces. The fragments were transferred through two layers of nylon sieves (pore size 125 µm and 106 µm). After sieving, mouse tubular fragments were selected and seeded in collagen pre-coated flasks (Corning) with Dulbecco’s modified Eagle’s/Ham’s F12 (DMEM-F12) containing 10% heat inactivated FBS, 1% L-glutamine, and 1% penicillin/streptomycin. The plate was incubated in a standard humidified incubator equipped with 5% CO₂. The medium was changed two days later, and maintained every other day until the cell monolayer reached a confluency of 90%. Immunostaining against megalin (SC-16478, Santa Cruz) confirmed the presence of tubular cells.

Evaluation of lymphocyte populations
At the conclusion of the Ang II-infusion protocol, the thymus, spleen, and kidney were collected for analysis. Single cell suspensions of splenocytes and thymocytes were prepared as described previously, counted, and resuspended in FACS buffer for flow cytometry analysis or in R10 complete medium for in vitro culture experiments. To generate single cell suspensions from kidney, kidneys were minced with a scissors, digested with type 1 collagenase for 40 minutes at 37 degrees Celsius, and then passed through a 40µM strainer. Flow cytometry was then performed as previously described using the following antibodies: PE-labeled anti-CD3; PE-Cy5-labeled anti-CD4; FITC-labeled anti-CD8; APC-labeled anti-CD19; PE-labeled anti-Thy1; PerCP-Cy5.5-labeled anti-CD45; PE-labeled anti-CD4; APC-labeled anti-CD3; PerCP-Cy5.5-labeled CD160 (NK T cell); APC-labeled anti-CD25 (all BD Bioscience); FITC-labeled anti-PD-1; PE-labeled anti-TIM-3 (both eBioscience); FITC-labeled anti-F4/80 (Serotec); and corresponding isotype antibodies. After final washing, splenocytes and thymocytes labeled for phenotyping analysis were fixed in PBS containing 4% paraformaldehyde and analyzed within 72 hours. Cells labeled for gene expression analysis were suspended in R10 (splenocytes / thymocytes) or DMEM with 10% FBS (kidney cell suspensions) without fixation, sorted immediately for enumeration and RNA extraction. In the kidney cell suspensions, anti-CD45 expression was used to parse mononuclear cells from renal parenchymal cells. Analyses were performed on a FACSCalibur (BD Biosciences) at the Flow Cytometry Facility at Duke University.
Isolation of RNA and Realtime PCR
Total RNA was isolated from individual cells or tissues by using the RNasea Mini Kit according to manufacturer’s instructions. RNA expression levels were then determined for interferon-γ (IFN-γ), tumor necrosis factor-α (TNF-α), transforming growth factor-β (TGF-β), interleukin-1β (IL-1β), interleukin-4 (IL-4), monocyte chemoattractant protein 1 (MCP-1), CCL5 (regulated on activation, normal T expressed and secreted), neutrophil gelatinase-associated lipocalin (NGAL), CD4, CD8, T-bet, and GATA-3 using TaqMan primers (Applied Biosystems) in real-time PCR, as described previously. RNA expression levels were determined for the AT1a receptor (forward 5’-GCTTGGTGTTGATCGTCACC-3’ and reverse 5’-GGGCGAGATTAGAGAAGGC-3’ and GAPDH (forward 5’-TCACCACCATGGAGAAGGC-3’ and reverse 5’-GCTAAGCAGTTGGTGTCGA-3’) using SYBR Green (Bio-Rad) in real-time PCR.

Microarray Analysis of T cells Infiltrating the Kidney
CD4+ T cells were isolated from the kidneys of Ang II-infused T cell WT and KO experimental animals via fluorescent cell sorting as described above. RNA was then harvested as above and reverse-transcribed to cDNA, which was subjected to quantitative RT-PCR in a 92-gene Taqman Mouse Immune Response Array (Applied Biosystems; n=3 per group). The TaqMan Mouse Immune Array is a 384-well microfluidic card based system. The primers and probe for each assay are preloaded and dried onto the designated duplicate wells. Taqman cards were run on the Applied Biosystems 7900HT real-time PCR. The master mix for each array card was prepared as following: 60 µL of cDNA (20ng total input RNA/ µL) and an equal amount of TaqMan Universal Master mix (Applied Biosystems) were added into a master tube. Each reaction was mixed by pipetting and was dispensed into loading wells on the Taqman low density array (TLDA) card. The final volume in each TLDA well is approximately 1 µL. The cards were centrifuged twice at 300 × g for 1 min and placed into the PCR system. The following cycling conditions were used for the TLDA reactions: 50°C for 2 min, 95°C for 10 min, and 40 cycles of 95°C for 15 s followed by 60°C for 1 min.

Cell proliferation assays
For CFSE experiments, splenic lymphocytes were obtained as described above, labeled by CFSE (Renovar) at a concentration of 2µg/ml, and then seeded into 96-well plates at a density of 5×10^3/well, according to a previously published protocol.11 Wells were pre-coated with 0.1 µg/ml anti-CD3ε antibody (No Azide/Low Endotoxin, BD Bioscience) or vehicle. After culture at 37°C for 72 hours, cells from individual wells were collected, stained with PE-labeled anti-CD4 (Bioscience), and processed for flow cytometric analysis. Proliferation was measured as the geometric Mean Fluorescence Index (MFI), or 100 X the ratio of the geometric mean channel fluorescence in stimulated cells over that in unstimulated cells, as previously described.12

Mixed lymphocyte assays were conducted as previously described 13 using irradiated BALB/c (H-2b) stimulators to activate (C57BL/6 x 129/SvEv)F1 (H-2b) lymphocytes in culture for 72hrs followed by pulsing with 0.5 µCi of [3H]thymidine per well for the final 20 hours in culture. Each sample was conducted in quadruplicate and is representative of 3 separate experiments. Values are expressed as specific counts per minute with counts from responder wells only subtracted from counts with both stimulators and responders.

For chronic T cell receptor stimulation assays, splenic lymphocytes were harvested as above and cultured for 7 days on plates pre-coated with vehicle or anti-CD3s: 0.5µg/ml (BD Bioscience). Ang II 1µM (Sigma) or vehicle was added to the culture daily. Media was changed on day 4. Following 7 days, RNA was harvested from the cultures and analyzed for gene expression of IFN-γ and TNF-α as above. Data were collected from duplicate wells for each condition with 3 animals in each experimental group.
ELISPOT Assay for Interferon-\(\gamma\)
ELISPOT plates were coated with anti-IFN-\(\gamma\) capture Ab (4 \(\mu\)g/ml; BD) overnight. The plates were then blocked for 90 minutes and washed with sterile PBS. Splenic lymphocytes were placed in each well (5x10^5 cells/well) with vehicle or anti-CD3 Ab for 24 h at 37 deg C. After washing, biotinylated rat anti-mouse IFN-\(\gamma\) detection Ab (4 \(\mu\)g/ml) was added overnight. After washing with PBS/Tween, streptavidin-horseradish peroxidase was added for 90 minutes. The plates were developed using 3-amino-9-ethylcarbazole (AEC) mixed in 0.1 M sodium acetate plus 12ml \(\text{H}_2\text{O}_2\). The resulting spots were counted on a computer-assisted ELISPOT image analyzer.

Cytokine ELISA
Splenic lymphocytes were obtained from both \textit{T cell KO} and \textit{WT} mice at the end of the Ang II infusion protocol as above. Cytokines were quantified in the media after the stated time of coculture of the lymphocytes with anti-CD3\(\varepsilon\) (NA/LE, BD) or vehicle. ELISA assays targeting IFN-\(\gamma\), TNF-\(\alpha\), and IL-4 were performed with specific kits from Invitrogen according to manufacturer's instructions.

Statistics
The values of each parameter within a group are expressed as the mean ± the standard error of the mean (SEM). For comparisons between groups with normally distributed data, statistical significance was assessed using ANOVA followed by unpaired student’s t test. For comparisons between groups with non-normally distributed variables, the Mann Whitney U test was employed. For comparison within groups, normally distributed variables were analyzed by a paired t test, whereas non-normally distributed variables were analyzed by the Wilcoxon signed rank test. For the comparison of proportions of renal vessels in the experimental groups surrounded by T lymphocyte infiltrates, a chi square test or Fisher’s Exact test was used, depending on the category frequencies.
Online Figure I. Detection of AT$_1$A receptor on T lymphocytes. Agtr1a mRNA expression measured by Real-time PCR in heart (H), Kidney (K), and T lymphocytes (T) with representative bands on top and summary data on bottom.
Online Figure II

Online Figure II. Isolation of CD4+ and CD8+ T cells for assessment of cell-specific AT1A receptor deletion in T cell KO mice. Diagram of workflow for fluorescent cell sorting of splenic T lymphocytes.
Online Figure III. Ang II induces significant cardiac hypertrophy in T cell WT and KO animals. Cardiac hypertrophy indices were obtained by calculating the ratio of heart weight (mg) to body weight (g) at day 28 of saline (n=4) or angiotensin II (n=11-12) infusion. *P = 0.0002 vs. Saline T cell WT; #P < 0.0001 vs. Saline T cell KO.
Online Figure IV. T cell infiltration into heart is increased following Ang II. CD3+ T cells were manually counted with a blinded method on stained sections of heart after 28 days of saline (n=4) or angiotensin II (n≥11) infusion. T cell counts were similar in Ang II-infused T cell WT and KO hearts. Representative heart sections are shown on top with T lymphocytes staining brown. *P = 0.099 vs. Saline T cell WT; #P = 0.002 vs. Saline T cell KO.
Online Figure V. Ang II upregulates cardiac expression of T cell chemokine CCL5. CCL5 mRNA expression was measured in T cell WT and KO hearts following saline or Ang II infusion. *P = 0.002 vs. Saline T cell WT; #P = 0.0002 vs. Saline T cell KO, P < 0.06 vs. Ang II T cell WT.
Online Figure VI. Ang II causes rise in serum creatinine in experimental groups. Serum creatinines were measured at day 28 of saline (n=3-4) or angiotensin II (n=14-15) infusion. *P = 0.07 vs. Saline T cell WT.
Online Figure VII. Ang II induces increase in urine volumes in T cell WT and KO mice. Urine volumes were measured at day 25 of saline (n=4) or angiotensin II (n≥13) infusion. *P = 0.003 vs. Saline T cell WT; #P = 0.01 vs. Saline T cell KO.
Online Figure VIII. *T cell KO* mice have increased numbers of CD4⁺ T lymphocytes in the kidney following 4 weeks of *Ang II* infusion. Kidneys from Ang II-infused animals were digested with collagenase to single cell suspensions. T lymphocytes were then isolated through fluorescent cell sorting as illustrated in Online Figure I, and the numbers of CD4⁺ and CD8⁺ T cells were counted. *P < 0.04 vs. T cell WT.*
Online Figure IX. CCL5 is expressed in kidney tubular epithelium but at lower levels than in lymphocytes. mRNA expression of CCL5 in freshly isolated wild-type kidney tubular epithelial cells (TEC), whole kidney (KID), and lymphocytes (LYMPH) with representative gel on top and summary data on bottom.
Online Figure XA, Contour plots of proliferative response measured by CFSE method in splenic CD4+ T cells from T cell WT and KO mice in absence ("Resting") and presence ("Stimulated") of T cell receptor activation (n=3).
Online Figure XB

**Online Figure XB**, Same representative CFSE proliferation data from Figure XA displayed as histograms to reveal cellular division peaks. The CD4⁺ T cells from both groups manifested 1 full division and initiated a second division during 72hr of T cell receptor stimulation with anti-CD3ε.
Online Figure XC, Summary data of geometric mean fluorescence index for CD4+ T lymphocytes from T cell WT and KO animals [100 x (mean fluorescence of stimulated cells / mean fluorescence of resting cells)] calculated from 3 independent experiments.
Online Figure XI, Expressions of IFN-γ, TNF-α, T-bet, and GATA-3 mRNA in CD4+ T cells isolated from the kidneys of Ang II infused-T cell WT and KO mice. *P = 0.03 vs. T cell WT; †P < 0.0002 vs. T cell WT; #P = 0.0003 vs. T cell WT; ‡P < 0.02 vs. T cell WT.
Online Figure XII. Mixed Lymphocyte Assay. Proliferative response as measured by thymidine incorporation in naïve (C57BL/6 x 129/SvEv)F$_1$ WT or Agtr1a$^{-/-}$ KO lymphocytes stimulated by irradiated splenocytes (Stimulators) from BALB/c (H-2$^d$) mice. “CPM’s”= counts per minute. *$P = 0.0005$ vs. WT.
Online Figure XIII. IFN-γ ELISPOT Assay. Frequency of IFN-γ-producing lymphocytes identified by ELISPOT following T cell receptor stimulation with incremental doses of anti-CD3ε. * P < 0.006 vs. WT.
Online Figure XIV. T-bet deficiency ameliorates hypertensive kidney damage. A, Blood pressures measured by radiotelemetry in uni-nephrectomized WT and T-bet KO mice at baseline and during chronic Ang II infusion.
Online Figure XIVB-C, Staining of glomerular podocytes with WT1 following 4 weeks of Ang II. Podocytes stain bright green. Representative images of (B) wild-type and (C) T-bet KO glomeruli. (Magnification 40X)
Online Figure XIVD, Number of podocytes per glomerulus in WT and T-bet KO (“KO”) kidneys scored in blinded fashion following 4 weeks of Ang II (8 mice per group) or saline (4-5 mice per group). *P<0.003 vs. Ang II-KO, P = 0.0009 vs. Saline-WT.
Online Figure XIVE-F, T-bet-deficiency blunts Ang II-induced macrophage accumulation in kidney. F4/80+ macrophage stains in representative kidneys from Ang II-infused (E) WT and (F) T-bet KO mice.
Online Figure XIVG, mRNA expressions of IFN-γ and TNF-α in WT and T-bet KO kidneys following Ang II. * $P < 0.0002$ vs. WT; † $P < 0.005$ vs. WT.
**Online Table I.** Genes with expression altered by >1.5 fold in CD4⁺ T cells isolated from kidneys of Ang II-infused *T cell WT* and *KO* animals. *P*-values are given for differences achieving at least borderline significance (*P* ≤ 0.06). “Increased” indicates increased in *T cell KO* group.

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REFERENCES


