Loss of Pentameric Symmetry in C-Reactive Protein Induces Interleukin-8 Secretion Through Peroxynitrite Signaling in Human Neutrophils

Tarek Khreiss,* Levente József,* Lawrence A. Potempa, János G. Filep

Abstract—Plasma levels of C-reactive protein (CRP), nitrotyrosine, and interleukin-8 (IL-8) are known predictors of acute cardiovascular events. Peroxynitrite (ONOO−) may function as an intracellular signal for the production of IL-8; however, it is not known whether CRP regulates these events. Emerging evidence suggests that some bioactivities of CRP are expressed only when the pentameric structure of CRP is lost, resulting in formation of monomeric or modified CRP (mCRP). We studied the impact of human native CRP and bioengineered mCRP that cannot rearrange into the pentameric structure on ONOO− formation and ONOO−-mediated IL-8 gene expression in human leukocytes. Incubation of human whole blood or isolated neutrophils with mCRP (0.1 to 100 μg/mL) for 4 hours increased IL-8 gene expression and secretion that was blocked ~70% by the NO synthase inhibitor L-NAME. In neutrophils, mCRP simultaneously increased superoxide production and endothelial nitric oxide synthase-mediated NO formation, leading to enhanced ONOO− formation, and consequently activation of nuclear factor-κB and activator protein-1. Native CRP had no detectable effect at 4 hours, whereas it enhanced IL-8 release after a 24-hour incubation that was blocked by L-NAME. An anti-CD16 antibody, but not an anti-CD32 antibody, produced 60% to 70% reductions in mCRP-stimulated NO formation and IL-8 release (both P<0.05). These results suggest that loss of the pentameric symmetry in CRP, resulting in formation of mCRP, leads to IL-8 release from human neutrophils via peroxynitrite-mediated activation of nuclear factor-κB and activator protein-1. (Circ Res. 2005;97:0-0.)

Key Words: C-reactive protein ■ leukocytes ■ interleukins ■ signal transduction ■ inflammation

Epidemiological and clinical studies have shown strong and consistent relationships between circulating markers of inflammation and risk prediction of future coronary artery disease (CAD). Among these markers, elevated plasma levels of C-reactive protein (CRP) are predictive for subsequent acute coronary events among apparently healthy men and women and patients with stable or unstable angina.1 The C-X-C chemokine interleukin-8 (IL-8) is a sensitive marker of unstable CAD.2 Increases in IL-8 levels may coincide with peroxynitrite formation and ONOO−-mediated IL-8 gene expression in human leukocytes. Peroxynitrite (ONOO−) may function as an intracellular signal for the production of IL-8; however, it is not known whether CRP regulates these events. Emerging evidence suggests that some bioactivities of CRP are expressed only when the pentameric structure of CRP is lost, resulting in formation of monomeric or modified CRP (mCRP). We studied the impact of human native CRP and bioengineered mCRP that cannot rearrange into the pentameric structure on ONOO− formation and ONOO−-mediated IL-8 gene expression in human leukocytes. Incubation of human whole blood or isolated neutrophils with mCRP (0.1 to 100 μg/mL) for 4 hours increased IL-8 gene expression and secretion that was blocked ~70% by the NO synthase inhibitor L-NAME. In neutrophils, mCRP simultaneously increased superoxide production and endothelial nitric oxide synthase-mediated NO formation, leading to enhanced ONOO− formation, and consequently activation of nuclear factor-κB and activator protein-1. Native CRP had no detectable effect at 4 hours, whereas it enhanced IL-8 release after a 24-hour incubation that was blocked by L-NAME. An anti-CD16 antibody, but not an anti-CD32 antibody, produced 60% to 70% reductions in mCRP-stimulated NO formation and IL-8 release (both P<0.05). These results suggest that loss of the pentameric symmetry in CRP, resulting in formation of mCRP, leads to IL-8 release from human neutrophils via peroxynitrite-mediated activation of nuclear factor-κB and activator protein-1. (Circ Res. 2005;97:0-0.)

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neutrophil activation, adherence, and trafficking.\textsuperscript{5,23–25} whereas mCRP promotes neutrophil adhesion to endothelial cells\textsuperscript{19} and suppresses neutrophil apoptosis.\textsuperscript{26}

In the present study, we investigated the impact of native pentameric CRP and mCRP on IL-8 production in human whole blood and isolated neutrophils. To gain insight into the underlying molecular mechanisms in neutrophils, we characterized the immunoglobulin (Ig) G receptor subtype (ie, the CRP receptor) involved and examined whether induction of IL-8 gene and protein expression is mediated through stimulation of ONOO\textsuperscript{-}-dependent activation of NF-κB and AP-1.

Materials and Methods
CRP Isoforms
High purity (>99%) human native CRP (Calbiochem) was stored in Na\textsubscript{2}CO\textsubscript{3},free 20 mmol/L Tris, 150 mmol/L NaCl buffer (pH 7.5) containing 2 mmol/L CaCl\textsubscript{2} to prevent spontaneous formation of mCRP from the native pentamer. A recombinant form of mCRP (rCRP, purity >97%) that cannot rearrange into a pentameric structure was engineered, characterized, and compared with mCRP prepared from native CRP by urea elution as described.\textsuperscript{26} Native CRP was distinguished from mCRP by binding and antigenicity differences\textsuperscript{27} and by their secondary structure.\textsuperscript{26} The endotoxin of all protein solutions was below the detection limit (0.125 endotoxin units/mL), corresponding to ~0.01 ng/mL LPS of the Limulus assay (Sigma).

Cell Stimulation
Venous blood (anticoagulated with sodium heparin, 50 U/mL) was obtained from 24 healthy volunteers who had denied taking any medication for at least 2 weeks. The Clinical Research Committee approved the experimental protocols. Neutrophil granulocytes (purity >95%, viability >97%) were isolated as described.\textsuperscript{23} Whole blood aliquots or isolated neutrophils (5 \times 10\textsuperscript{6} cells/mL) in microcentrifuge tubes were placed on a rotator and challenged with native CRP or mCRP with or without the NO synthase inhibitor L-NAME (1 mmol/L), and fluorescence was analyzed by a flow cytometry (FACScan, Becton Dickinson).\textsuperscript{15} The NO synthase blocker-inhibitable proportion of DHR whereas NO does not.\textsuperscript{29} DHR 123 oxidation by mCRP was also determined in neutrophils pretreated with the calmodulin inhibitor W7 (20 \mu mol/L), the broad PKC inhibitor GF109203X (200 nM), the phosphatidylinositol 3 (PI3)-kinase inhibitor wortmannin (100 nM), or the NADPH oxidase inhibitor diphenyleneiodonium chloride (DPI, 5 \mu mol/L). In additional experiments, neutrophil extracts (prepared as in reference15) were analyzed for the presence of nitrotyrosine, a “molecular fingerprint” of NOx by an enzyme immunoassay (detection limit: 2 ng/mL) (Cayman Chemicals) using nitrotyrosine as standard.\textsuperscript{15} Intra-assay and interassay coefficients of variation were typically <6%.

NF-κB and AP-1
Intracellular, DNA bound, NF-κB/p65 and AP-1/c-Fos were measured with a flow cytometry assay\textsuperscript{13,15} and were used as estimates of NF-κB and AP-1 activation, respectively. In brief, leukocyte nuclei prepared with the Cycletest Plus DNA reagent kit (Becton Dickinson) were stained with rabbit polyclonal anti-human NF-κB/p65 or c-Fos antibodies or with normal rabbit IgG (to assess nonspecific binding of IgG to nuclei) and then with FITC-conjugated anti-rabbit IgG antibody (all from Santa Cruz Biotechnology) and propidium iodide. Single neutrophil nuclei were gated using the doublet-discrimination module and fluorescence intensity was analyzed with a FACScan flow cytometer using the Cell Quest Pro software.

Calcium Mobilization Assay
Intracellular Ca\textsuperscript{2+} concentration was monitored in Fura-2/AM (1 \mu mol/L)-loaded neutrophils in a Perkin-Elmer spectrofluorometer (excitation: 340 nm, emission: 510 nm) as previously described.\textsuperscript{30}

Western Blot Analysis
Protein extracts were prepared by lysing 2 \times 10\textsuperscript{6} neutrophils in 100 \mu L of lysis buffer, and immunoblot analysis of phosphorylated and total Akt was performed using the Phospho Plus Akt antibody kit (New England Biolabs) as described.\textsuperscript{26}

Neutrophil Viability
Neutrophil viability was assessed by flow cytometry immediately after staining with propidium iodide (0.5 \mu mol/L).

Statistical Analysis
Results are expressed as mean±SEM. Statistical comparisons were made by ANOVA using ranks (Kruskal-Wallis test) followed by Dunn’s multiple contrast hypothesis tests to identify differences between various treatments. Repeated measures were analyzed by the Friedman test followed by the Wilcoxon-Wilcon test. Values of P<0.05 were considered significant.
Results

Effects of CRP and mCRP on IL-8 Production and IL-8 mRNA Expression

Incubation of human whole blood with mCRP for 4 hours resulted in concentration-dependent IL-8 release, whereas native CRP was without effect (Figure 1). Significant induction was detected with 5\( \mu \)g/mL, which peaked at 100\( \mu \)g/mL mCRP (Figure 1B). Native CRP started to increase IL-8 release only after 8 hours of incubation; however, it was a considerably less potent inducer of IL-8 production than mCRP at any time points studied (Figure 1A). Both mCRP- and CRP-induced IL-8 release were markedly, though never completely, inhibited by L-NAME (Figure 1A). Likewise, mCRP, but not CRP, induced concentration-dependent IL-8 release from isolated neutrophils after 4 hours of incubation (Figure 1C). On a molar basis, bioengineered mCRP and mCRP prepared from native CRP evoked similar IL-8 production (Figure 1D). Recombinant mCRP-induced (50\( \mu \)g/mL) IL-8 release (5.6\pm0.7 ng/mL) was unaffected by the formyl-peptide receptor antagonist N-t-Boc-Phe-Leu-Phe-Leu-Phe (50\( \mu \)mol/L) (5.4\pm0.5 ng/mL, \( n = 6, \ P < 0.1 \)). Therefore, because of the enhanced solubility, bioengineered mCRP was used in subsequent experiments. Heat inactivation of native CRP (60 minutes at 100°C) resulted in a complete loss of its activity (data not shown). Also, LPS (Escherichia coli serotype O111:B4) at 0.02 ng/mL, a concentration 2-fold higher than the maximum level of LPS contamination in our protein solutions, did not induce detectable IL-8 release (0.24\pm0.03 ng/mL versus 0.23\pm0.03 ng/mL in unstimulated neutrophils, \( n = 4, \ P > 0.1 \)).

We performed RNase protection assays on RNA extracted from leukocytes after 4 hours of incubation with mCRP. Consistent with the observations at the protein level, mCRP evoked concentration-dependent increases in IL-8 mRNA expression, which were suppressed by L-NAME (Figure 2). Native CRP at 100 \( \mu \)g/mL did not produce detectable changes (Figure 2).

mCRP Induces NO, Superoxide, and Peroxynitrite Formation in Neutrophils

Incubation of neutrophils for 4 hours with mCRP led to simultaneous increases in O\( _2^- \) and NO production. Significant increases were detected with 5\( \mu \)g/mL and peaked at 100\( \mu \)g/mL mCRP (Figure 3). The increases in O\( _2^- \) and NO production coincided with increases in NO-dependent DHR123 oxidation, indicating enhanced ONOO\(^- \) formation.

Figure 1. CRP isoforms induce IL-8 release from neutrophils and whole blood. Blood samples (A and B) or isolated neutrophils (PMN, 5\( \times \)10^6 cells/mL) (C) were incubated (A) with CRP or mCRP (100 \( \mu \)g/mL) in the absence and presence of L-NAME (1 mmol/L) or (B and C) with various concentrations of CRP and mCRP or (D) with bioengineered mCRP or mCRP prepared from native CRP at 37°C for the indicated times. Secreted IL-8 was determined by ELISA. Results are mean\( \pm \)SEM for 4 to 6 experiments with different donor cell preparations. Blood/neutrophils obtained from the same persons were used to study CRP and mCRP plus L-NAME. *\( P < 0.05; ** P < 0.01; *** P < 0.001 \) vs unstimulated (control). L-NAME inhibited mCRP or CRP (100 \( \mu \)g/mL)-stimulated IL-8 release by \( \approx 70\% \) (\( P < 0.01 \) at all time points studied; for the sake of clarity symbols are not shown).

Figure 2. Effects of mCRP and CRP on IL-8 mRNA expression. Neutrophils were incubated for 20 minutes with L-NAME (1 mmol/L) and challenged with mCRP or CRP for 4 hours at 37°C. A. Representative RNase protection assay using probes for IL-8 and GAPDH. B. Densitometry analysis of autoradiographs of the samples probed for IL-8 and GAPDH. The IL-8 results are expressed as percentage of control (unstimulated) after normalization with the GAPDH values. The results represent mean\( \pm \)SEM from 4 assays with different blood donors. *\( P < 0.05; ** P < 0.01; *** P < 0.001 \) vs unstimulated. ††\( P < 0.01 \).
cytochrome c in the culture medium. ONOŌ measured as superoxide dismutase-inhibitable reduction of ferricytochrome c in the culture medium. Superoxide formation was monitored using diaminoflorescein, and is expressed as relative nitrotyrosine correlated with those in DHR 123 oxidation (rhodamine fluorescence in relative fluorescence units, RFU). Values are mean ± SEM (n = 4 to 7). *P < 0.05, **P < 0.01; ***P < 0.001 vs unstimulated (control).

Figure 3. mCRP induces formation of superoxide, NO, and ONOŌ. Neutrophils (5 × 10⁶ cells/mL) were challenged with mCRP at 37°C for 4 hours. Intracellular formation of NO was monitored using diaminoflorescein, and is expressed as relative fluorescence units (RFU). Diaminoflorescein fluorescence was reduced 98% by L-NAME (1 mmol/L). Superoxide formation was measured as superoxide dismutase-inhibitable reduction of ferricytochrome c in the culture medium. ONOŌ formation was assessed as L-NAME-inhibitable oxidation of DHR 123 to rhodamine and is expressed as RFU. Values are mean ± SEM (n = 4 to 7). *P < 0.05, **P < 0.01; ***P < 0.001 vs unstimulated (control).

To identify the NOS isofrom(s) responsible for enhanced NO production, we performed reverse transcription polymerase chain reaction on cDNA prepared from neutrophils. These assays resulted in the amplification of eNOS mRNA, but not neuronal NOS and iNOS, in freshly isolated cells and in response to mCRP at 2 and 4 hours after addition of the protein (Figure 4). mCRP did not affect eNOS mRNA expression (Figure 4).

To characterize the proximal signaling events associated with mCRP-induced ONOŌ formation, we monitored calcium mobilization. mCRP evoked a rapid increase in intracellular Ca²⁺ similar to that observed with fMLP (Figure 5A), and transiently enhanced (peak at around 2 minutes) phosphorylation of Akt (Figure 5B), indicating phosphatidylinositol 3-kinase activation. Consistently, calmodulin blockade with W7 or inhibition of PI3-kinase with wortmannin significantly reduced mCRP-induced DHR 123 oxidation, and these actions were not additive with L-NAME (Figure 5C).

Figure 4. Effect of mCRP on expression of NOS isoforms. Neutrophils incubated without or with mCRP (50 μg/mL) for 2 and 4 hours were harvested for RNA preparation. Reverse-transcriptase polymerase chain reaction was performed with specific primers for human nNOS, iNOS, eNOS, and β-actin. The polymerase chain reaction products were resolved on a 1.2% agarose gel by electrophoresis and stained with ethidium bromide. RNA isolated and amplified from human chondrocytes stimulated with IL-1β for 4 hours served as a positive control for iNOS. The results are representative for 3 neutrophil preparations.

mCRP Stimulates Nuclear Accumulation of AP-1 and NF-κB in Neutrophils

Because transcription of IL-8 gene requires activation of NF-κB and AP-1, we examined whether mCRP can activate these pathways in neutrophils. We prepared nuclei from unstimulated and mCRP-stimulated neutrophils and analyzed nuclear accumulation of NF-κB and AP-1 using flow cytometry. Figure 6A shows representative results illustrating mobilization of NF-κB/p65 and AP-1/c-Fos to the nucleus (ie, the increased fluorescence represents increased amounts of NF-κB or AP-1 bound to DNA) in response to mCRP. Nuclear immunostaining was completely blocked by preincubation of the antibodies with the appropriate blocking peptides. The actions of mCRP were concentration-dependent and were markedly attenuated by L-NAME (Figure 6A and 6B), consistent with inhibition of IL-8 mRNA expression and production.

Preincubation of neutrophils with PDTC markedly attenuated, though never completely inhibited, mCRP-induced IL-8 production (Figure 7), confirming that increased nuclear accumulation of NF-κB correlated with induction of IL-8 production.

Involvement of CD16 in mCRP Signaling

Previous studies identified CD16 (FcγRIII) as a neutrophil receptor for mCRP. We used function-blocking mAbs to CD16 and CD32 (FcγRIIa) as competitors to confirm whether the mCRP actions described above were also mediated via CD16. The anti-CD16 mAb partially prevented
mCRP-induced NO formation assessed at 4 hours, and IL-8 release assessed at 4 and 24 hours after addition of mCRP, whereas the anti-CD32 or the irrelevant MOPC-21 antibody had no detectable effects (Figure 8). Increasing the concentration of the anti-CD16 mAb did not result in further inhibition (data not shown). At 24-hours' culture, native CRP-induced IL-8 release was also attenuated by the anti-CD16 mAb, but not by the anti-CD32 mAb or MOPC-21 (Figure 8B).

**Effect of mCRP on Neutrophil Viability**

Because increased NO and ONOO⁻ formation could affect cell viability,⁷,³² we assessed neutrophil survival. Consistent with previous studies,²⁶ after 4 hours culture in vitro, the percentage of viable neutrophils was slightly higher in the presence of mCRP (95²±⁵%, n=5, P<0.05) than in untreated (control) samples (89²±³%, n=5, P<0.05 versus mCRP).

**Discussion**

The present results provide evidence for a novel mechanism by which CRP may affect the inflammatory process by stimulating ONOO⁻ formation and signaling in human neutrophils. These actions are expressed when the pentameric structure undergoes a conformational rearrangement, however, leading to formation of mCRP. Our results also suggest a potential link between CRP, IL-8, nitrotyrosine, and neutrophil activation, as it may occur in the blood of patients with acute CAD.

Our study using human whole blood and isolated neutrophils demonstrates that mCRP stimulates a rapid (within 4 hours) and potent synthesis of IL-8. This is a primary...
response to mCRP that requires de novo protein synthesis and transcription of the IL-8 gene. Comparison of plasma and culture medium IL-8 levels suggests that ~70% of IL-8 release was of neutrophil origin in blood. Native CRP did not evoke detectable changes at 4 hours. This was unexpected, for native CRP induced rapid (within minutes) shedding of L-selectin from the surface of neutrophils, and cannot be explained by the slight differences in the viability of mCRP or CRP-treated neutrophils. Human blood contains all the yet unidentified serum cofactors that were required for CRP activation of endothelial cells. The CRP stimulation became detectable only after 8 hours of incubation, coinciding with in vitro kinetics of dissociation into subunits. Although CRP clearly enhanced IL-8 production at 8 to 24 hours of incubation, it was a considerably less potent inducer of IL-8 release than mCRP. These observations suggest that conformational rearrangement of CRP is required to induce IL-8 production in neutrophils, and that the amounts of mCRP generated from CRP within 4 hours are not sufficient to evoke detectable increases in IL-8. The time course of neutrophil activation by native CRP appears to be similar to that of CRP activation of endothelial cells. Neither LPS (at a concentration 2-fold higher than might be present in our protein preparations) nor heat-inactivated CRP evoked detectable IL-8 release, indicating that CRP and mCRP signaling was responsible for the observed effects.

Blockade of NO synthesis with L-NAME inhibited to a similar degree mCRP-induced IL-8 release and nuclear accumulation of NF-κB and AP-1 in neutrophils, coinciding with suppression of IL-8 mRNA expression. Interestingly, L-NAME also attenuated native CRP-induced IL-8 release, and the degree of inhibition was comparable to that detected with mCRP. These results pointed to the involvement of NO in mediating these responses. Previous studies have shown that ONOO⁻ rather than NO by itself mediates IL-8 release from human neutrophils in response to LPS or cytokines. Accordingly, we found that mCRP simultaneously enhanced superoxide and NO formation, coinciding with increases in NO-dependent oxidation of DHR123 and nitration of protein tyrosine residues. A significant portion of rhodamine fluorescence in mCRP-stimulated neutrophils can be attributed to ONOO⁻, because it depends on NO-related species (it can be inhibited by L-NAME), whereas NO per se does not oxidize DHR 123. Further, the NO donor spermine NONOate restored DHR 123 oxidation in L-NAME-treated neutrophils, indicating that L-NAME does not inhibit NADPH oxidase activation. Although nitrotyrosine is often considered as a distinct “molecular fingerprint” of ONOO⁻ formation, peroxidase-dependent tyrosine nitration has also been described. Interestingly, in human neutrophils,
ONOO− appears to be the predominant mechanism for tyrosine nitration.38

Reverse-transcriptase polymerase chain reaction amplified eNOS, but not iNOS and neuronal NOS-specific products, in unchallenged neutrophils and in neutrophils treated with mCRP for up to 4 hours. These results point toward increased eNOS activity as the source for enhanced NO production in response to mCRP. Previous studies on constitutive NOS expression in human neutrophils yielded contradictory results, as both the absence and presence of eNOS and neuronal NOS have been reported.28,39–41 Contaminating cells or differences in the NOS assays used might account for this apparent discrepancy. Unstimulated human neutrophils do not express iNOS, whereas iNOS-positive neutrophils have been detected in tissue exudates and after more than 16 hours incubation with cytokines.41 Whether mCRP could induce iNOS expression after prolonged incubation periods remains to be investigated. There is compelling evidence that eNOS-derived NO contributes to ONOO− formation in amounts sufficient to activate signaling mechanisms and even to induce cell damage.32

Ca2+ mobilization and activation of calmodulin and PI3-kinase appears to be required for mCRP-induced ONOO− formation. Ca2+ transients control calmodulin-mediated eNOS activation and the activity of PLCγ1 and PLCγ2, which through formation of diacylglycerol and activation of PI3-kinase lead to activation of NADPH oxidase.42 The inhibitory actions of W7, wortmannin, and DPI on DHR 123 oxidation were not additive with those of L-NAME, suggesting that these compounds inhibited the same reaction, ie, the ONOO−-dependent oxidation through suppression of either NO or O2− formation. PKC appears to play a minor role in mCRP signaling, for GF109203X inhibited only a small portion of DHR 123 oxidation by mCRP.

The IL-8 gene contains cis-regulatory elements for NF-κB, AP-1, and NF-IL-6.31 Of these transcription factors, NF-κB plays a key role in the induced expression of IL-8. Accordingly, we found that mCRP stimulates nuclear accumulation of NF-κB and AP-1, and inhibition of NF-κB activation with PDTC decreased IL-8 production by 66%. The mechanism of action of PDTC has not been fully defined, but likely involves inhibition of formation of oxidants that would result in activation of IκB-α kinase and/or enhancing phosphorylation of I-κB.43,44 Thus, NF-κB activation could be attributed to decomposition products of ONOO− rather than the parent molecule itself.

Our results indicate that mCRP-induced IL-8 gene expression and release are predominantly mediated through the low affinity immune complex binding IgG receptor CD16, for the function-blocking anti-CD16 mAb 3G8, but not anti-CD32 mAb, markedly, though never completely, inhibited NO formation and IL-8 production in neutrophils. Furthermore, only the anti-CD16 mAb attenuated native CRP-induced IL-8 release at 24 hours incubation. These observations imply that during 24-hours’ culture, conformational rearrangement might have occurred in native CRP, yielding mCRP, because native CRP does not bind to CD16.20–22 We cannot exclude the possibility that mCRP may bind different sites from IgG on CD16, for blockade of the epitope defined by the mAb 3G8 reduced ~80% of mCRP binding at 0°C.22 Alternatively, mCRP might interact with other as yet unidentified cell surface molecules. These may include direct binding to positively charged residues on proteins and direct interaction with the lipid membrane (Potempa et al, unpublished observations). The slight inhibition of mCRP activation of endothelial cells by anti-CD16 mAb34 lends support to the existence of mCRP receptor(s) other than CD16.

The mechanisms that induce conformational rearrangement in native CRP in vivo are still unknown. Native, pentameric CRP dissociates into free subunits after binding to plasma membranes or in denaturing or oxidative environment,27 yielding mCRP. The percentage of CRP that might have dissociated into subunits during a 24-hour incubation period, however, remains to be determined. Unlike CRP, mCRP appears to be predominantly membrane-bound,45,46 therefore making its detection rather difficult. Because the commercial CRP antibody (clone 8) mainly recognizes mCRP,47 tissue immunostaining attributed to native CRP in previous studies might have been due to mCRP. Neutrophils may come into contact with mCRP expressed in the arterial wall at sites of endothelial injury or perhaps on other leukocytes, thereby aggravating the inflammatory response and contributing to plaque destabilization.11 Stimulated neutrophils release ONOO−, though the ratio of intracellularly retained and released ONOO− cannot be deduced.12 Increased ONOO− release is consistent with enhanced nitration of plasma proteins and may contribute to tissue damage. Finally, our results do not exclude contribution of other cells to elevated plasma levels of nitrotyrosine and IL-8. Indeed, mononuclear leukocytes and endothelial cells also release IL-8 in response to native CRP or mCRP, thereby increasing plasma cytokine levels in patients with CAD.

In summary, the present results indicate that loss of pentameric symmetry in CRP, resulting in formation of mCRP, stimulates IL-8 production by human neutrophils through ONOO−-mediated activation of NF-κB and AP-1. These observations point toward neutrophils as a major source of nitrosative stress and IL-8 and may provide a potential link between CRP, neutrophil activation, plasma nitrotyrosine, and IL-8, all implicated in predicting future acute CAD.

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References


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