Oxygen Regulation of Tumor Perfusion by S-Nitrosohemoglobin Reveals a Pressor Activity of Nitric Oxide


Abstract—In erythrocytes, S-nitrosohemoglobin (SNO-Hb) arises from S-nitrosylation of oxygenated hemoglobin (Hb). It has been shown that SNO-Hb behaves as a nitric oxide (NO) donor at low oxygen tensions. This property, in combination with oxygen transport capacity, suggests that SNO-Hb may have unique potential to reoxygenate hypoxic tissues. The present study was designed to test the idea that the allosteric properties of SNO-Hb could be manipulated to enhance oxygen delivery in a hypoxic tumor. Using Laser Doppler flowmetry, we showed that SNO-Hb infusion to animals breathing 21% O₂ reduced tumor perfusion without affecting blood pressure and heart rate. Raising the pO₂ (100% O₂) slowed the release of NO bioactivity from SNO-Hb (ie, prolonged the plasma half-life of the SNO in Hb), preserved tumor perfusion, and raised the blood pressure. In contrast, native Hb reduced both tumor perfusion and heart rate independently of the oxygen concentration of the inhaled gas, and did not elicit hypertensive effects. Window chamber (to image tumor arteriolar reactivity in vivo) and hemodynamic measurements indicated that the preservation of tissue perfusion by micromolar concentrations of SNO-Hb is a composite effect created by reduced peripheral vascular resistance and direct inhibition of the baroreceptor reflex, leading to increased blood pressure. Overall, these results indicate that the properties of SNO-Hb are attributable to allosteric control of NO release by oxygen in central as well as peripheral tissues. (Circ Res. 2005;96:000-000.)

Key Words: nitric oxide • hemoglobin • oxygen • hemodynamics • blood flow

---

Hemoglobin (Hb) of red blood cells (RBC) is a tetrameric protein composed of 2 α and 2 β chains, each containing a heme prosthetic group. One α and 1 β chain is combined in stable αβ dimers, and 2 dimers are more loosely associated to form tetramers. The physiological role of Hb depends on its ability to reversibly bind O₂ at its heme iron centers. This transport capacity is governed by a cycle of allosteric transitions in which Hb assumes the R (relaxed, high O₂ affinity) conformation to bind O₂ in the lungs and, on partial deoxygenation, the T (tense, low O₂ affinity) conformation to bind O₂ in the lungs and, on partial deoxygenation, the T (tense, low O₂ affinity) conformation to efficiently deliver O₂ to peripheral tissues. This transition also controls the reactivity of 2 conserved cysteines on the β chains (Cys893). Thiols of the cysteines can react with the potent vasodilator nitric oxide (NO) to form S-nitrosohemoglobin (SNO-Hb) in the R conformation, and preferentially unload SNO in the T conformation.1–3 It has therefore been proposed that Hb would carry NO equivalents from the lungs to the periphery, thereby bringing tissue blood flow in line with oxygen demand. Although mechanisms are debated,4–7 some consensus has been reached, namely that Hb can act as an oxygen sensor and oxygen-dependent transducer of NO bioactivity,8–9 and the direct coupling between SNO and O₂ content of Hb has been recently affirmed in intact human erythrocytes (Doctor et al, unpublished observations).

Outside the RBC, Hb has toxic effects that prevent its use as a blood substitute in humans.10 In plasma, Hb rapidly dissociates to αβ dimers, leading to hemoglobinuria (renal injury) and extravasation (osmotic disorders).11 Several types of conjugated and intramolecularly cross-linked Hbs have been developed that reduce Hb dissociation, thereby lowering the risks for such toxicity.12 These modified Hbs also show prolonged vascular retention times, making them promising candidates for oxygen delivery.
blood substitute candidates. However, clinical safety is still not achieved, mainly because modified Hbs retain the vasoconstrictive activity of cell-free Hb.5,3 Ferrous Hb and oxy-Hb can react with NO to produce iron-nitrosyl Hb, and oxidized Hb and nitrate ions (met-Hb reaction), respectively.14,15 The Hb concentrations required to exert such activity have not been carefully examined, and the extent to which vasodilation is involved (under more physiological circumstances, Hb can also react with NO or S-nitrosothiols to yield SNO-Hb)16 remains a matter of debate. Lower doses of Hb-based blood substitutes do not typically raise blood pressure.

To date, SNO-Hb, which combines the capacity to carry oxygen and the potential to act as a hypoxia-activated NO donor, has not yet been fully tested as a RBC substitute.17 In the present work, we hypothesized that S-nitrosylation of cell-free Hb would reduce the vasoconstrictive effects of tetrameric Hb. We also hypothesized that blood oxygenation, through allosterically regulated control of NO release, would be a key determinant of the biological responses to SNO-Hb. These hypotheses were investigated in vivo using a physiological model of vascular hypoxia (perfused tumor vessels), in which pO2 was modulated in situ by varying inspired O2. These hypotheses were investigated in vivo using a physiological model of vascular hypoxia (perfused tumor vessels), in which pO2 was modulated in situ by varying inspired O2. This prospective study offers a new perspective on the development of transfusion substitutes, and establishes that delivery of NO bioactivity by SNO-Hb is regulated by O2 saturation.

Methods

Rats and Tumors

Seventy-four female Fischer 344 rats (Charles River, Raleigh, NC), bearing or not bearing the syngeneic mammary adenocarcinoma R3320AC,18 were used in all experiments. Tumor pieces from donor rats were implanted in anesthetized animals (100 mg/kg ketamine and 10 mg/kg xylazine i.p.). Tumors were grown in the subcutis of the left lateral quadriceps muscle or, for window chamber experiments, between 2 fascial layers on the back of rats. Animals were then randomly assigned to a treatment group. Further interventions/observations were performed on anesthetized animals (50 mg/kg pentobarbital i.p.) maintained at 37°C on a temperature-controlled thermal blanket. Where indicated, some rats were fitted with a facemask for breathing 100% O2. All experiments were approved by Duke IACUC.

Drugs

SNO-Hb was synthesized through reaction of purified human hemoglobin A0 (Apex Bioscience, Durham, NC) with SNO-cysteine in nonacidic conditions, as previously reported19 (expanded method at http://www.circresaha.org). SNO-albumin was synthesized by the reaction of an equal volume of bovine serum albumin (0.2 mmol/L NaNO2 for 30 minutes. All solutions (diluted at 200 µmol/L in PBS pH 7.4, 0.5 mmol/L EDTA) were kept on ice or frozen and protected from light until administration. Less than 2-weeks old solutions were infused at a dose of 200 nmol/kg in 0.25 mL, immediately followed by a 0.25 mL delivery of saline. Assuming a plasma volume of ~3 mL per 100 g in rats and no significant partition in blood cells, this dose would achieve a maximum plasma concentration of ~6.5 µmol/L for all drugs. To avoid volume-induced alterations of hemodynamics, solutions were infused at 0.5 mL/min from time 0.

Surgery

For i.v. infusions, the femoral artery and vein were cannulated and drugs were infused through the carotid cannula. For i.a. infusions, the femoral and left carotid arteries were cannulated and drugs were infused through the carotid cannula. For window chamber experiments, the 2 window frames were surgically positioned in the dorsal skin flap, followed by tumor cell transplantation, as previously described.20 Following several days of tumor growth, direct visualization of tumor-feeding arterioles was performed using intravital microscopy.

Blood Gas and Hemoglobin Saturation Measurements in Tumor Microvessels

Blood gases and Hb oxygen saturation were measured using a 1640 oximeter coupled to a 482 cooximeter (Instrumentation Laboratories, Lexington, Mass) from 0.5 mL samples of femoral arterial and venous blood. For tumor microvessels in window chambers, Hb saturation was calculated from transmission optical measurements of vascular absorbance using a hyperspectral imaging technique. A liquid crystal tunable filter (CRI, Inc., Woburn, Mass) placed in front of a CCD camera (DVC Company, Austin, Tex) was used for band-limited imaging.

Blood Flow

Blood flow was measured using Laser Doppler flowmetry. For flank tumor experiments, 300-µm diameter Laser Doppler probes (OxiFlo, Oxford Optronix, Oxford, UK) were simultaneously inserted into the tumor and quadriceps muscle of each animal. For window chamber experiments, Laser Doppler probes (LASERFLO, TSI, St. Paul, Minn) were positioned beneath the tumor window as previously described.22

SNO-Hb Assay

The determination of SNO-Hb concentrations is based on a reaction where cleavage of S-nitrosothiols yields a nitrosant that activates 4,5-diaminofluorescein (DAF-2, Sigma, St. Louis, Mo). Data acquired before drug infusion allowed us to determine basal SNO levels. The plasma half-life of SNO-Hb was calculated from exponential decay curves fitting each experimental curve. Hb concentration in plasma samples was determined by visible spectrophotometry. Expanded method is available online.

Vascular Reactivity In Vivo

Tumor-feeding arterioles in window chambers were visualized using transillumination. Arteriolar diameters were measured from videotaped images using an image-shearing monitor (IPM Inc., San Diego, Calif).

Mean Arterial Pressure and Heart Rate

Mean arterial pressure (MAP) and heart rate (HR) were measured with a blood pressure analyzer (Digi-Med, Micro-Med, Louisville, Ky) connected to the femoral artery cannula, as previously described.20

Statistical Analyses

All values are shown as means±S.E. Time curves were normalized to the baseline before infusion. ‘N’ refers to the number of animals per group and ‘n’ to individual measurements. Repeated-measure 2-way ANOVA or Student’s t tests were used as indicated.

Results

SNO-Hb and Oxy-Hb i.v. Reduce Muscle and Tumor Perfusion in Normoxic Rats

We first examined whether i.v. infusion of human cell-free oxy-Hb and SNO-oxy-Hb (SNO-Hb) influenced muscle perfusion in the quadriceps of rats breathing room air (normoxia). SNO-Hb decreased muscle perfusion more than Hb when compared with albumin (Figure 1A, P=0.055 and 0.3 versus albumin, respectively, 2-way ANOVA). We then determined the pharmacological effects of these molecules in flank tumors of normoxic rats. In contrast to albumin, Hb
induced a rapid and sustained decrease in tumor blood flow (Figure 1B, \(P < 0.01\) versus albumin, 2-way ANOVA). Unexpectedly, i.v. infusion of SNO-Hb also reduced perfusion (Figure 1C, \(P < 0.05\) versus SNO-albumin, 2-way ANOVA). Reductions in flow caused by oxy-Hb and SNO-Hb were not different (\(P > 0.05\), 2-way ANOVA). However, the effects of the 2 Hbs were quite different from the small increase in tumor perfusion that we observed following albumin or SNO-albumin infusion i.v. (Figure 1B and 1C). The effects of SNO-albumin and albumin were not different (\(P > 0.05\), 2-way ANOVA).

Hyperoxia Prolongs the Half-life and Modulates the Pressor Activity of Cell-Free SNO-Hb

Based on previous observations,\(^2\) we reasoned that an increase in plasma oxygenation would stabilize SNO-Hb, increase peripheral delivery and thereby increase the bioactivity in tumors. We aimed to determine whether the plasma half-life of SNO-Hb i.v. was prolonged in rats breathing 100% O\(_2\) (hyperoxia) versus room air. Blood gas measurements showed that breathing 100% O\(_2\) induced significant increases in venous and arteriolar blood pO\(_2\), pCO\(_2\), and endogenous Hb oxygen saturation. Changes in Hb saturation were identical in femoral vein and tumor venules. Changes were less pronounced in tumor-feeding arterioles versus the femoral artery, as expected from blood deoxygenation along the arterial tree and at the tumor margin (see Table in the online supplement).

Because of low concentration (\(\approx 6.5\) \(\mu\)mol/L) and alteration in the tetramer/dimer ratio as a function of concentration and O\(_2\) saturation (dimer/tetramer equilibrium is 1000-fold lower in deoxy-Hb), it was not possible to directly measure the oxygen saturation of exogenous Hbs. Instead, we measured plasma SNO-Hb concentration versus time following infusion using a DAF-2 assay. The fitting of individual experimental data with exponential decay curves (\(R^2=0.87\pm 0.05\) and \(0.90\pm 0.03\) for normoxic and hyperoxic conditions, respectively) revealed a 2-fold increase in the half-life of the S-nitrosyl group in SNO-Hb in the plasma of hyperoxic versus normoxic rats (Figure 2, \(P < 0.01\), Student’s \(t\) test). We found no influence of O\(_2\) on the rate of SNO-Hb protein clearance (data not shown). Spectrophotometric measurements revealed that hyperoxia did not affect the rate of protein clearance during the period of the experiments (half-life \(\approx 30\) minutes, independent of O\(_2\) supply) (data not shown). Thus, our results, together with previous observations,\(^3\) suggest that the effects of O\(_2\) are mediated by an allosteric mechanism that promotes the R structure.

The effects of hyperoxia alone on tumor and muscle perfusion ranged from a transient decrease (that resumed within 5 minutes, i.e., before drug infusion) to no change. As expected from these observations (and in agreement with prior publications),\(^2,3\) hyperoxia abolished the reduction in tumor perfusion that we observed after i.v. infusion of SNO-Hb in normoxic rats (Figure 3A, \(P < 0.005\), 2-way ANOVA). In contrast to SNO-Hb, and native Hb reduced tumor perfusion independently of O\(_2\) supply (Figure 3B, \(P > 0.05\), 2-way ANOVA).

Figure 1. Time course of oxy-Hb- and SNO-Hb-induced decrease in tumor blood flow. Changes in muscle perfusion were determined by inserting Laser Doppler probes into the quadriceps muscle of rats breathing room air. (A) At time 0, animals were infused i.v. with albumin (\(\circ\), \(N=3\)), oxy-Hb (\(\bullet\), \(N=4\)) or SNO-Hb (\(\blacksquare\), \(N=7\)). The same protocol was used to measure tumor perfusion in rats infused i.v. with (B) oxy-Hb (\(\bullet\), \(N=5\), \(n=10\)) or albumin (\(\circ\), \(N=3\), \(n=3\)); or (C) SNO-Hb (\(\blacksquare\), \(N=7\), \(n=13\)), or SNO-albumin (\(\blacklozenge\), \(N=7\), \(n=14\)). *\(P < 0.05\), **\(P < 0.01\) between \(t=0\) and \(t+20\) (2-way ANOVA).

Figure 2. Oxygen breathing stabilizes the SNO moiety on plasma SNO-Hb in vivo. The concentration of SNO proteins was determined over time in the plasma of rats following i.v. infusion of SNO-Hb. The half-life of SNO-Hb is shown for rats breathing room air or 100% O\(_2\). (\(N=5\)). *\(P < 0.01\) (Student’s \(t\) test).
Hyperoxia Unmasks the Systemic Pressor Activity of Cell-Free SNO-Hb

We then sought to identify the relative contribution of changes in vascular activity versus systemic hemodynamics in the tumor perfusion response to SNO-Hb. In window chambers, i.v. infusion of SNO-Hb induced no significant change in the diameter of feeding arterioles in normoxic or hyperoxic rats (Figure 4A, \( P > 0.05 \), 2-way ANOVA). As reported in our previous studies,20,22 hyperoxia alone had no vascular effect (compare t5 to t=0 in Figure 4A, white boxes). Interestingly, in the same rats, we simultaneously documented that SNO-Hb induced a potent decrease in tumor perfusion in normoxic animals, and that this decrease was prevented in hyperoxic rats (data not shown). Thus, these results were identical to what we observed in flank tumors (Figure 3A).

Changes in the tumor blood flow were paralleled by changes in the perfusion of the leg quadriceps muscle: the decrease in muscle blood flow after delivery of SNO-Hb under normoxia was prevented under hyperoxia (Figure 4B,
Intra-arterial infusion preserves the pressor activity of SNO-Hb but accounts for loss of O2-dependence. SNO-Hb was infused i.v. or i.a. in rats at t=0. When indicated, rats breathed O2 from t=−5 (arrowheads). Changes in perfusion were simultaneously determined using Laser Doppler probes in rat tumors A, or in the quadriceps muscle B. (A) SNO-Hb was infused i.v. in animals breathing room air, N=7, n=13 or 100% O2, N=7, n=14, or i.a. with room air, N=7, n=14 or 100% O2, N=7, n=13. Note the loss of O2-dependence for i.a. infusions. (B) SNO-Hb was infused i.v. in animals breathing room air, N=7 or 100% O2, N=6. C, shows changes in the mean arterial pressure (MAP) and/or the heart rate (HR) in the heart rate (HR) following i.a. infusion of SNO-Hb. Rats breathed room air, N=7 or 100% O2, N=7. (C-D): P<0.05 versus values at t=0 (Student’s t test).

**Figure 5.** Intra-arterial infusion preserves the pressor activity of SNO-Hb but accounts for loss of O2-dependence. SNO-Hb was infused i.v. or i.a. in rats at t=0. When indicated, rats breathed O2 from t=−5 (arrowheads). Changes in perfusion were simultaneously determined using Laser Doppler probes in rat tumors A, or in the quadriceps muscle B. (A) SNO-Hb was infused i.v. in animals breathing room air, N=7, n=13 or 100% O2, N=7, n=14, or i.a. with room air, N=7, n=14 or 100% O2, N=7, n=13. Note the loss of O2-dependence for i.a. infusions. (B) SNO-Hb was infused i.v. in animals breathing room air, N=7 or 100% O2, N=6. C, shows changes in the mean arterial pressure (MAP) and/or the heart rate (HR) in the heart rate (HR) following i.a. infusion of SNO-Hb. Rats breathed room air, N=7 or 100% O2, N=7. (C-D): P<0.05 versus values at t=0 (Student’s t test).

**The Route of Administration Impacts the Pressor Activity of Cell-Free SNO-Hb**

The route of administration is a key factor in determining the pressor activity of cell-free SNO-Hb. In normoxic animals, the pressor effect of i.v. SNO-Hb is significantly greater compared to i.a. infusion. However, in i.a. infusion of SNO-Hb, the pressor activity of i.a. SNO-Hb was not modified. This suggests that the route of administration can alter the pressor effect of SNO-Hb, potentially due to differences in tissue oxygenation and perfusion between i.v. and i.a. routes.

**Discussion**

Despite molecular modifications that successfully overcome the toxicity of unstable Hb tetramers, vasoconstriction remains a major limitation in the clinical use of Hb-based blood substitutes. Cell-free Hb preparations are devoid of the SNO that normally serves to counter their vasoconstrictive effects in vivo. We therefore investigated whether SNO reconstitution of Hb could reverse the vasoconstrictor activity of oxy-Hb, and whether SNO-Hb could be manipulated allosterically to maximize O2 delivery. In rats breathing room air (normoxia), SNO-Hb induced a greater decrease in tissue oxygenation compared to oxy-Hb, suggesting that SNO-Hb has a greater pressor effect. This observation highlights the importance of optimizing the route of administration and molecular modifications to enhance the clinical utility of SNO-Hb in oxygen delivery systems.
perfusion than native Hb (Figure 1A and Table 1). These results suggested that oxy-Hb and SNO-Hb operate by different mechanisms. Whereas the effects of oxy-Hb could be readily explained by NO scavenging (the “oxyhemoglobin,” or met-Hb forming-reaction), those of SNO-Hb were consistent with other NO donors, which paradoxically also decrease tissue perfusion. NO-mediated dilatation of healthy blood vessels upstream of tumors creates shunts that divert blood away from the tumors (vascular steal). By raising the concentration of the inhaled O2, SNO-Hb bioactivity can be targeted to more distally blood vessels, and elicits improvements in tumor blood flow.

We and recently others1,2,3 have proposed that SNO-Hb is formed in concert with O2 uptake in the lung, and that the release of vasodilator NO from SNO-Hb is favored at low O2. We demonstrated that plasma half-life of SNO in Hb was more than doubled during hyperoxia whereas protein clearance remained unaffected. Because of ongoing angiogenesis, most vessels downstream of feeding arterioles in fast-growing rodent tumors lack structural elements for vasoactivity, and they lack functional endothelial NO synthase.25-27 Hence, during normoxia, i.v. SNO-Hb must decrease tumor perfusion through vascular steal. This is consistent with the previous observation that, although oxy-Hb provokes a decrease in healthy muscle perfusion (as expected from a NO scavenger), it increases perfusion in muscle surrounding the R3230AC tumor (a NO donor-like response) consistent with a steal effect.25

To gain further mechanistic insight, we reasoned that manipulation of blood oxygenation would impact SNO-Hb bioactivity. As reported in our previous studies,20,22 normobaric 100% oxygen breathing (hyperoxia) induced a significant increase in venous and arteriolar pO2. Although the delivery of pure oxygen sometimes reduced tumor and muscle perfusion (see Figure 3A, 3B, 5A, and 5B between t = −5 and t = 0), this effect was transient and returned to baseline within <5 minutes. Thus, the vasoactive properties of oxygen per se did not affect the response to SNO-Hb or oxy-Hb. On i.v. infusion in hyperoxic animals, SNO modified Hb successfully opposed the reduction in perfusion created by Hb in tumors (compare Figure 3A and 3B). Increases in MAP were seen in response to SNO-Hb during 100% O2 breathing, but there was no change in tumor/muscle perfusion or HR (Table 1). By comparison, the reduction in tumor perfusion by native Hb remained unaffected by hyperoxia (Figure 3B). This is likely accounted for by lowered systemic perfusion and/or bradycardia, not by changes in MAP (Table 1).

Hb dissociation from tetramers to dimers depends on Hb concentration and oxygen tension, and NO release from dimers is unresponsive to allosteric effectors (eg, O2).27 Assuming a plasma concentration of ≈6.5 μmol/L, ≈35% of SNO-Hb would be tetramers (K0 for R and T conformers are 3 μmol/L and 3 nmol/L, respectively) at room air and the amount would increase precipitously as O2 tension declines - the majority would be tetramers at tissue pO2. Is there enough tetrameric SNO-Hb to sense oxygen and regulate NO delivery? Evidently yes, as we documented a finely tuned regulation of SNO-Hb bioactivity by oxygen tension (NO release depends on local pO2) that contrasted with the O2-independent behavior of oxy-Hb. We further demonstrated that the plasma half-life of SNO in Hb was more than doubled during hyperoxia whereas protein clearance remained unaffected. Thus, the prolonged half-life of SNO at high pO2 is consistent with the expectation that release of NO bioactivity is disfavored in the R structure.1,2 Collectively these data provide a definitive demonstration of allosteric regulation by O2 of NO delivery from SNO-Hb.

During hyperoxia SNO-Hb is greatly stabilized, surviving arteriovenous transit. SNO-Hb bioactivity was thus indistinguishable following hyperoxic i.v. and normoxic i.a. infusions (Table 1). To address a point of confusion, we note that SNO-Hb will exert activity at both high and low pO2 but that this activity will be potentiated at low pO2.23 Effects will

| TABLE 1. Hemodynamic Properties of Cell-free Human SNO-Hb and Oxy-Hb |
|-----------------|----------------|-----------------|----------------|
|                 | Tumor          | Vascular        | Systemic       |
|                 | Perfusion      | Diameter        | Perfusion      |
|                 |                |                 | MAP            |
|                 |                |                 | HR             |
| SNO-Hb          |                |                 |                |
| IV              | Room air       | ↓*              | 0              |
|                 | 100% O2        | 0               | 0 ↑            |
| IA              | Room air       | 0               | nd             |
|                 | 100% O2        | 0               | nd             |
| Oxy-Hb          | Room air       | nd              | 0              |
|                 | 100% O2        | nd              | 0 ↑            |

* refers to changes vs values before treatment: 0, unchanged; ↓, decreased; ↑, increased; nd, not determined. SNO-Hb, human S-nitrosohemoglobin; oxy-Hb, human oxyhemoglobin; IV, intravenous infusion; IA, intra-arterial infusion; MAP, mean arterial pressure; HR, heart rate.
Therefore manifest in deoxygenated vessels. Consistent with this interpretation, hyperoxia had no effect on i.a. infusions of SNO-Hb but markedly altered i.v. responses.

On close inspection, our data unravel a central effect of SNO-Hb on control of hemodynamics. An increase in MAP was observed following i.a. infusion during normoxia, and both i.v. and i.a. infusions during hyperoxia. Native Hb (hyperoxia, i.v.) did not produce increases in MAP (Table 1) under any condition. Baroreceptor activity opposes increases in MAP and resistance to flow. Native Hb activated the baroreceptor, which opposed changes in MAP by decreasing the HR. In contrast, strikingly, SNO-Hb raised the MAP without altering perfusion (in the organs that we monitored) or HR. The systemic pressor activity of SNO-Hb results from a direct inhibition of the baroreceptor reflex. Indeed, at higher concentrations (1 μmol/kg, i.v., normoxia), it has been reported that both SNO-Hb and oxy-Hb overcome baroreceptor buffering capacities (presumably indirectly through NO scavenging and peripheral vasoconstriction). Thus, SNO-Hb activity in hyperbaric hyperoxia is sufficient to inhibit the baroreceptor. As a neurotransmitter, NO is known to regulate baroreceptor activity through sympathetic afferent nerve fibers. Exogenous NO has an inhibitory action on the baroreceptor, albeit only at high concentrations, and under such circumstances that the baroreceptor activity recovers within seconds. Unlike classical NO activity, SNO-Hb bioactivity is mediated by species distinct from NO itself (eg, low-mass SNOs). It is noteworthy that other endogenous S-nitrosothiols (eg, SNO-cysteine) can suppress baroreceptor activity independently of cGMP generation (the mediator of classical nitrosovasodilator activity). Stereoselective recognition sites in the baroreceptor vasculature could mediate baroreceptor inhibition by S-nitrosylated species such as SNO-cysteine and SNO-Hb. Expanded discussion is available online.

In conclusion, we have shown that SNO reconstitution of Hb successfully overcomes the reduction in tumor perfusion created by Hb itself. This activity involves the allosteric control of NO release by O2. The hemodynamic effects of SNO-Hb are a composite of central activity (baroreceptor inhibition) and allostry-facilitated NO release in the peripheral circulation. In contrast, native Hb, at concentrations characteristic of hemolytic states, has no central pressor effect. These results are summarized in Table 2.

Whereas the concentration of Hb we used is low from the O2 delivery standpoint, it greatly exceeds the concentration of any endogenous NO. In light of our results, development of safe Hb-based blood substitutes might include approaches that not only preserve Hb allostergy (eg, crosslinking to prevent dissociation into dimmers) but also reconstitute SNO content. Reoxygenation of hypoxic tissues may benefit from both the central pressor and blood flow increasing effects elicited by SNO-Hb. Further, the treatment of trauma or septic patients might involve manipulation of SNO-Hb allostery to limit the excessive release of NO.

Acknowledgments

This work was supported by grants from the NIH/NCI CA40355 (M.W.D.), the Howard Hughes Medical Institute (J.S.S.), the NIH Division of Research Resources COBRE University of Puerto Rico Protein Research Center (Grant 5P20RR016439) (J.B.), and a collaborative travel award from the North Atlantic Treaty Organization (NATO) (G.T.). P.S. is a fellow of the Belgian American Educational Foundation (BAEF) and a ‘Fonds National de la Recherche Scientifique’ (FNRS) post-doctoral fellow detached from the University of Louvain Medical School (UCL, Unit of Pharmacology and Therapeutics, Brussels, Belgium). The authors appreciate the assistance of Christine Baudelet and Pelzer O. Doar.

**TABLE 2. Bioactivity of Cell-free SNO-Hb and Oxy-Hb**

<table>
<thead>
<tr>
<th>Vascular pO2 at Injection Site</th>
<th>NO-related Chemical Fate</th>
<th>Peripheral Effect</th>
<th>Baroreceptor Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>SNO-Hb</td>
<td>Low</td>
<td>Met-Hb + iron-nitrosyl Hb</td>
<td>Vasoconstriction*</td>
</tr>
<tr>
<td>uzione down</td>
<td>High</td>
<td>SNO-Hb</td>
<td>Vasodilation*</td>
</tr>
<tr>
<td>Oxy-Hb</td>
<td>Any</td>
<td>Met-Hb + iron-nitrosyl Hb</td>
<td>Vasoconstriction</td>
</tr>
</tbody>
</table>

*SNO-Hb, S-nitrosohemoglobin; oxy-Hb, oxyhemoglobin; NO, nitric oxide; Met-Hb, methemoglobin.

References


Oxygen Regulation of Tumor Perfusion by S-Nitrosohemoglobin Reveals a Pressor Activity of Nitric Oxide


_Circ Res._ published online May 5, 2005;

_Circulation Research_ is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231

Copyright © 2005 American Heart Association, Inc. All rights reserved.

Print ISSN: 0009-7330. Online ISSN: 1524-4571

The online version of this article, along with updated information and services, is located on the World Wide Web at:

http://circres.ahajournals.org/content/early/2005/05/05/01.RES.0000168740.04986.a7.citation

Data Supplement (unedited) at:

http://circres.ahajournals.org/content/suppl/2005/05/05/01.RES.0000168740.04986.a7v1.DC1
http://circres.ahajournals.org/content/suppl/2005/06/06/01.RES.0000168740.04986.a7.DC1

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in _Circulation Research_ can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:

http://www.lww.com/reprints

Subscriptions: Information about subscribing to _Circulation Research_ is online at:

http://circres.ahajournals.org//subscriptions/
Online data supplement.

**Oxygen Regulation of Tumor Perfusion Unravels the Central Pressor Activity of Cell-Free Human S-Nitrosohemoglobin**

Pierre Sonveaux*, PhD; Andrew M. Kaz*, MD; Stacey A. Snyder, BS; Rachel A. Richardson, DVM; L. Isabel Cárdenas-Navia, BS; Rodney D. Braun, PhD; John R. Pawloski, MD, PhD; Gillian M. Tozer, PhD; Joseph Bonaventura, PhD; Timothy J. McMahon, MD, PhD; Jonathan S. Stamler, MD; Mark W. Dewhirst, DVM, PhD.

**Corresponding author:**

Mark W. Dewhirst, DVM, PhD,

Department of Radiation Oncology,

Box 3455, Room 201 MSRB,

Duke University Medical Center,

Research Drive,

Durham, NC, 27710.

E-mail: dewhirst@radonc.duke.edu

* Both authors contributed equally to this study.
**Expanded methods.**

**SNO-Hb synthesis**

Solutions of purified human hemoglobin A₀ (Apex Bioscience, Durham, NC) were dialyzed overnight at 4°C against 2% (w/v) aerated sodium tetraborate pH 9.2, 0.5 mmol/L EDTA. For SNO-Hb synthesis, L-cysteine hydrochloride (0.55 mol/L in 0.5 mol/L HCl, 0.5 mol/L EDTA) was first reacted with an equal volume of 0.5 mol/L NaNO₂ to yield Cys-NO.¹ Then, a 10-fold molar excess of Cys-NO was reacted with dialyzed oxy-Hb at a 1:20 (v/v) ratio for 10 min. The reaction was terminated by centrifugation through a fine Sephadex G-25 chromatography column (Pharmacia Biotech, Uppsala, Sweden) equilibrated with PBS, pH 7.4, 0.5 mmol/L EDTA. Under these conditions, the yield of oxy-Hb S-nitrosylation is consistently 1.9 S-nitroso-cysβ93 per tetramer.¹²

**SNO-Hb Assay**

Briefly, blood samples were collected in heparinized tubes just before infusion and at 1 min, 5 min, and every 5 min after infusion of SNO-Hb i.v. for 30 min. Plasma (100 µl) was immediately mixed with DAF-2 (30 µmol/L final concentration). Half of this solution was then reacted for 10 min with an equal volume of HgCl₂ 1.2 mmol/L. The reaction was terminated by centrifugation in filter tubes (Nanosep, Pall, East Hills, NY). Upon light excitation at 485 nm, DAF-2-associated light emission was read at 520 nm. SNOs were quantified as the difference in fluorescence signals generated in the presence and absence of HgCl₂, which specifically cleaves SNOs, yielding nitrite.
Expanded Results.

Although the method we used for the preparation of SNO-Hb was designed to selectively nitrosylate thiols\(^1\), reaction in alkaline buffer did not fully prevent heme oxidation. By comparison to unreacted oxy-Hb which is typically 100% oxy-Hb(FeII), SNO-Hb solutions contain small amounts of SNO-Hb(FeIII). In our hands, the extent of heme oxidation had fallen further under normoxic and hyperoxic conditions as soon as 30 minutes after i.v. SNO-Hb administration (\(P>0.05\), \(n=5\), Student’s \(t\) test). The rate of heme reduction was independent of the oxygen concentration delivered to rats. Moreover, there was no correlation between SNO content and FeII/FeIII ratio in Hb over time (\(R^2 = 0.31\)).

Supplemental Table. Blood Gas and Hemoglobin Saturation Measurements.

<table>
<thead>
<tr>
<th></th>
<th>Femoral artery</th>
<th>Femoral vein</th>
<th>Tumor artery</th>
<th>Tumor vein</th>
</tr>
</thead>
<tbody>
<tr>
<td>pO(_2) (mm Hg)</td>
<td>82.4±0.4</td>
<td>57.5±1.2</td>
<td>67.7±2.4</td>
<td>76±2</td>
</tr>
<tr>
<td>pCO(_2) (mm Hg)</td>
<td>49.6±1.3</td>
<td>50.6±1.7</td>
<td>76±2</td>
<td>69±2</td>
</tr>
<tr>
<td>HbO(_2) (%)</td>
<td>88.8±0.4</td>
<td>67.7±2.4</td>
<td>76±2</td>
<td>69±2</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>Femoral artery</th>
<th>Femoral vein</th>
<th>Tumor artery</th>
<th>Tumor vein</th>
</tr>
</thead>
<tbody>
<tr>
<td>pO(_2) (mm Hg)</td>
<td>435.5±34.7*</td>
<td>69.8±3.0*</td>
<td>75.8±3.4*</td>
<td>89±2*</td>
</tr>
<tr>
<td>pCO(_2) (mm Hg)</td>
<td>56.9±1.7*</td>
<td>72.0±2.1*</td>
<td>78±3*</td>
<td>89±2*</td>
</tr>
<tr>
<td>HbO(_2) (%)</td>
<td>98.2±0.1*</td>
<td>75.8±3.4*</td>
<td>78±3*</td>
<td>89±2*</td>
</tr>
</tbody>
</table>

* \(P<0.05\) between value and corresponding room air value (paired Student’s \(t\) test, \(N=4\)). HbO\(_2\), endogenous hemoglobin oxygen saturation.
**Expanded discussion.**

In regard to the direct inhibition of the baroreceptor reflex by SNO-Hb, one can further envisage a mechanism independent of SNO generation. Indeed, unlike oxy-Hb, synthesized SNO-Hb contains small amounts of SNO-Hb(FeIII), which can scavenge NO’. Few studies have addressed the role of NO’ present in the vasculature on the baroreceptor in systems independent of peripheral hemodynamics. Recently however, Meyrelles et al showed that gene transfer of the endothelial NO’ synthase to the carotid sinus induced NO’-mediated vasodilation that inhibited the baroreceptor reflex; these effects were reversed by local NO’ synthase inhibition. If local changes in the availability of NO’ species are the main trigger for direct baroreceptor inhibition by SNO-Hb, it likely results from SNO donation rather than NO’ scavenging. Furthermore, the lack of correlation between SNO and FeIII content in SNO-Hb indicates that SNO donation is independent of the oxidation state in Hb.

**References.**


Online data supplement.

Oxygen Regulation of Tumor Perfusion Unravels the Central Pressor Activity of Cell-Free Human S-Nitrosohemoglobin

Pierre Sonveaux*, PhD; Andrew M. Kaz*, MD; Stacey A. Snyder, BS; Rachel A. Richardson, DVM; L. Isabel Cárdenas-Navia, BS; Rodney D. Braun, PhD; John R. Pawloski, MD, PhD; Gillian M. Tozer, PhD; Joseph Bonaventura, PhD; Timothy J. McMahon, MD, PhD; Jonathan S. Stamler, MD; Mark W. Dewhirst, DVM, PhD.

Corresponding author:
Mark W. Dewhirst, DVM, PhD,
Department of Radiation Oncology,
Box 3455, Room 201 MSRB,
Duke University Medical Center,
Research Drive,
Durham, NC, 27710.
E-mail: dewhirst@radonc.duke.edu

* Both authors contributed equally to this study.
**Expanded methods.**

**SNO-Hb synthesis**

Solutions of purified human hemoglobin A₀ (Apex Bioscience, Durham, NC) were dialyzed overnight at 4°C against 2% (w/v) aerated sodium tetraborate pH 9.2, 0.5 mmol/L EDTA. For SNO-Hb synthesis, L-cysteine hydrochloride (0.55 mol/L in 0.5 mol/L HCl, 0.5 mol/L EDTA) was first reacted with an equal volume of 0.5 mol/L NaNO₂ to yield Cys-NO.¹ Then, a 10-fold molar excess of Cys-NO was reacted with dialyzed oxy-Hb at a 1:20 (v/v) ratio for 10 min. The reaction was terminated by centrifugation through a fine Sephadex G-25 chromatography column (Pharmacia Biotech, Uppsala, Sweden) equilibrated with PBS, pH 7.4, 0.5 mmol/L EDTA. Under these conditions, the yield of oxy-Hb S-nitrosylation is consistently 1.9 S-nitroso-cysβ93 per tetramer.¹ ²

**SNO-Hb Assay**

Briefly, blood samples were collected in heparinized tubes just before infusion and at 1 min, 5 min, and every 5 min after infusion of SNO-Hb i.v. for 30 min. Plasma (100 µl) was immediately mixed with DAF-2 (30 µmol/L final concentration). Half of this solution was then reacted for 10 min with an equal volume of HgCl₂ 1.2 mmol/L. The reaction was terminated by centrifugation in filter tubes (Nanosep, Pall, East Hills, NY). Upon light excitation at 485 nm, DAF-2-associated light emission was read at 520 nm. SNOs were quantified as the difference in fluorescence signals generated in the presence and absence of HgCl₂, which specifically cleaves SNOs, yielding nitrite.
Expanded Results.

Although the method we used for the preparation of SNO-Hb was designed to selectively nitrosylate thiols, reaction in alkaline buffer did not fully prevent heme oxidation. By comparison to unreacted oxy-Hb which is typically 100% oxy-Hb(FeII), SNO-Hb solutions contain small amounts of SNO-Hb(FeIII). In our hands, the extent of heme oxidation had fallen further under normoxic and hyperoxic conditions as soon as 30 minutes after i.v. SNO-Hb administration ($P>0.05$, $n=5$, Student’s $t$ test). The rate of heme reduction was independent of the oxygen concentration delivered to rats. Moreover, there was no correlation between SNO content and FeII/FeIII ratio in Hb over time ($R^2 = 0.31$).

Supplemental Table. Blood Gas and Hemoglobin Saturation Measurements.

<table>
<thead>
<tr>
<th></th>
<th>Femoral artery</th>
<th>Femoral vein</th>
<th>Tumor artery</th>
<th>Tumor vein</th>
</tr>
</thead>
<tbody>
<tr>
<td>$pO_2$ (mm Hg)</td>
<td>82.4±0.4</td>
<td>57.5±1.2</td>
<td>67.7±2.4</td>
<td>76±2</td>
</tr>
<tr>
<td>$pCO_2$ (mm Hg)</td>
<td>49.6±1.3</td>
<td>50.6±1.7</td>
<td>72.0±2.1</td>
<td>89±2*</td>
</tr>
<tr>
<td>$HbO_2$ (%)</td>
<td>88.8±0.4</td>
<td>67.7±2.4</td>
<td>75.8±3.4*</td>
<td>78±3*</td>
</tr>
</tbody>
</table>

* $P<0.05$ between value and corresponding room air value (paired Student’s $t$ test, $N=4$). $HbO_2$, endogenous hemoglobin oxygen saturation.
**Expanded discussion.**

In regard to the direct inhibition of the baroreceptor reflex by SNO-Hb, one can further envisage a mechanism independent of SNO generation. Indeed, unlike oxy-Hb, synthesized SNO-Hb contains small amounts of SNO-Hb(FeIII), which can scavenge NO’. Few studies have addressed the role of NO’ present in the vasculature on the baroreceptor in systems independent of peripheral hemodynamics. Recently however, Meyrelles et al showed that gene transfer of the endothelial NO’ synthase to the carotid sinus induced NO’-mediated vasodilation that inhibited the baroreceptor reflex; these effects were reversed by local NO’ synthase inhibition. If local changes in the availability of NO’ species are the main trigger for direct baroreceptor inhibition by SNO-Hb, it likely results from SNO donation rather than NO’ scavenging. Furthermore, the lack of correlation between SNO and FeIII content in SNO-Hb indicates that SNO donation is independent of the oxidation state in Hb.

**References.**

