Molecular Mechanisms Mediating Inhibition of Human Large Conductance Ca\(^{2+}\)-Activated K\(^{+}\) Channels by High Glucose

Tong Lu, Tongrong He, Zvonimir S. Katusic, Hon-Chi Lee

Abstract—Diabetic vascular dysfunction is associated with an increase in reactive oxygen species (ROS). In this study, we hypothesized that hyperglycemia-induced ROS generation would impair the function of large conductance Ca\(^{2+}\)-activated K\(^{+}\) (BK) channels, which are major determinants in vasorelaxation. We found that when cultured in high glucose (HG) (22 mmol/L), HEK293 cells showed a reduction in expressed hSlo current densities, as well as slowed activation and deactivation kinetics. When human coronary smooth muscle cells were cultured in HG, similar findings were observed for the BK currents. HG enhanced superoxide dismutase and suppressed catalase (CAT) expression in HEK293 cells, leading to a significant increase in intracellular ROS. The effects of HG were mimicked by hydrogen peroxide (H\(_2\)O\(_2\)), and hSlo functions were restored by CAT gene transfer. Peroxynitrite inhibited hSlo current density but did not change channel kinetics. The hSloC911A mutant was insensitive to the effects of HG and H\(_2\)O\(_2\). Hence, imbalance of antioxidant enzymes plays a critical role in ROS generation in HG, impairing hSlo functions through H\(_2\)O\(_2\)-dependent oxidation at cysteine 911. This may represent an important fundamental mechanism that contributes to the impairment of vasodilation in diabetes. (Circ Res. 2006;99:607-616.)

Key Words: large conductance Ca\(^{2+}\)-activated K\(^{+}\) channels ■ hyperglycemia ■ catalase ■ reactive oxygen species ■ gene transfer

Diabetes is associated with a 2- to 4-fold increase in the risk of developing coronary artery disease and stroke, and there has been a 75% increase in mortality from cardiovascular complications over the last decade in the United States.1,2 Nearly 21 million Americans have diabetes, with another 41 million considered prediabetic. Multiple factors contribute to the development of diabetic vasculopathy.1,2 Endothelial dysfunction, reduced nitric oxide (NO) availability, and enhanced production of reactive oxygen species (ROS), such as superoxide anion (O\(_2^-\)) and hydrogen peroxide (H\(_2\)O\(_2\)), are thought to be major mechanisms that underlie the development of cardiovascular complications in diabetes.3 O\(_2^-\) is rapidly oxidized to H\(_2\)O\(_2\) by superoxide dismutase (SOD), and H\(_2\)O\(_2\) is further reduced to H\(_2\)O by catalase (CAT) and glutathione peroxidase (GPX).3 In addition, O\(_2^-\) reacts with NO extremely efficiently to form peroxynitrite (OONO\(^-\)). H\(_2\)O\(_2\) and OONO\(^-\) are highly reactive ROS. Abnormal ROS metabolism, leading to cellular oxidative stress, plays a central role in the progression of diabetic vascular dysfunction.1,3 The large conductance Ca\(^{2+}\)-activated K\(^{+}\) (BK) channel, abundantly expressed in vascular smooth muscle cells, plays a critical role in controlling vascular tone.4 The BK channel is composed of 4 pore-forming a subunits, each with 7 transmembrane domains (S0 to S6), including a highly conservative pore region between S5 and S6 and a voltage sensor located at S4.5 The cytoplasmic terminus has 4 hydrophobic segments (S7 to S10) that contain critical channel regulatory sites, including 2 regulators of K\(^{+}\) conductance (RCK1 and RCK2) domains and the Ca\(^{2+}\) sensor located at S4.5 The cytoplasmic terminus has 4 hydrophobic segments (S7 to S10) that contain critical channel regulatory sites, including 2 regulators of K\(^{+}\) conductance (RCK1 and RCK2) domains and the Ca\(^{2+}\) binding motif (Ca\(^{2+}\) bowl).6,7 Movement of the RCK domains affects Ca\(^{2+}\) binding affinity.7 There is evidence that BK channel–mediated vasodilation is impaired in hyperglycemia; however, the molecular mechanism has not been fully delineated.8,9 Hyperglycemia-induced ROS overproduction in diabetes is well documented.10 In vascular smooth muscle cells, a major source of ROS is NAD(P)H oxidase,11 which is present in HEK293 cells.11 OONO\(^-\) inhibits vascular BK channels in a dose-dependent manner, whereas O\(_2^-\) itself has no effect on BK channels.12,13 H\(_2\)O\(_2\) may either inhibit15,16 or activate BK channels through direct effects17,18 or through cGMP- and cAMP-dependent signaling pathways.19,20 Cysteine 911 of hSlo, which encodes the BK channel, is thought to be involved in channel regulation by redox and cysteine-modification reagents,14 although studies have produced conflicting results.21 Most of the previous studies used exogenously applied H\(_2\)O\(_2\) from 10 \(\mu\)mol/L to 167 nmol/L.14,16,17 Whether endogenously produced H\(_2\)O\(_2\) in hyperglycemia can

Original received January 25, 2006; resubmission received June 14, 2006; revised resubmission received July 24, 2006; accepted August 16, 2006.
From the Departments of Internal Medicine (T.L., H.-C.L.) and Anesthesiology (T.H., Z.S.K.), Mayo Clinic, Rochester, Minn.
Correspondence to Tong Lu, MD, PhD, Division of Cardiovascular Diseases, Department of Internal Medicine, Mayo Clinic, 200 First St SW, Rochester, MN 55905. E-mail lu.tong@mayo.edu.
© 2006 American Heart Association, Inc.

Circulation Research is available at http://circres.ahajournals.org
DOI: 10.1161/01.RES.0000243147.41792.93

607
modulate BK channel function is unknown. In this study, we hypothesized that a high glucose (HG)-induced increase in ROS modulates hSlo channel function by oxidation of specific residues. We examined changes in the properties of hSlo channels, which encode human BK channel α subunits, in HG. The results may help elucidate the molecular mechanisms underlying diabetes-associated vascular dysfunction and the role of ROS in the regulation of BK current functions in blood vessels.

**Materials and Methods**

An expanded Materials and Methods section containing details for cell culture, transfection, vasoreactivity, Western blotting, and electrophysiology is available in the online data supplement at http://circres.ahajournals.org.

**Cell Culture and hSlo, CAT Transfection**

HEK293 cells were cultured in DMEM containing normal glucose (NG) (5 mmol/L) or HG (22 mmol/L). Recombinant adenoviruses, AdCAT carrying CAT and AdLacZ carrying β-galactosidase, were used for gene transfer.

**Animals and Vasoreactivity Measurements**

Diabetes mellitus was produced in male Sprague–Dawley rats (200 g, Harlan, Inc, Ind) by streptozotocin (60 mg/kg, IP). Control rats received vehicle injection. Blood glucose in excess of 300 mg/dL was considered diabetic. Animals were used for vasoreactivity experiments 4 weeks after development of diabetes. Handling and care of animals, as well as all animal procedures, were approved by the Institutional Animal Care and Use Committee, Mayo Foundation.

On the day of the experiment, rats were anesthetized with pentobarbital (50 mg/kg IP). The heart was quickly removed from the thorax. Small coronary arteries were isolated from control and diabetic rats and were used for gene transfer.

**Electrophysiology**

hSlo currents were recorded in HEK293 cells 72 hours after hSlo transfection using standard patch clamp techniques at room temperature (23°C).

**Intracellular ROS Measurement**

Products of intracellular H$_2$O$_2$ and O$_2^-$ were detected as previously described. Briefly, attached HEK293 cells were exposed to 5 μmol/L 5,6-chloromethyl-2,7′-dichlorodihydrofluorescein diacetate (CM-H$_2$DCF-DA) for 30 minutes at 37°C in the dark, washed twice with PBS, and incubated in NG or HG medium for another 30 minutes. Cells were trypsinized (0.1%) and washed with PBS. After centrifugation (1200 rpm for 10 minutes), the cell pellet was suspended in 200 μL of PBS. Formation of ROS was detected by signals from the fluorescent reaction products of CM-DCF and ethidium using flow cytometry (Becton Dickinson FACSCan System).

**Western Blot Analysis**

Protein expressions of manganese SOD (MnSOD), copper zinc SOD (CuZnSOD), CAT, and GPX-1 in HEK293 cells cultured with NG and HG were determined using Western blot.

**Chemicals**

OONO$^-$ was purchased from Upstate Cell Signaling and Solutions (Lake Placid, NY). All other chemicals were obtained from Sigma-Aldrich (St Louis, Mo).

**Statistical Analysis**

Data are presented as mean±SEM. Student $t$ test was used to compare data between 2 groups. Paired $t$ test was used to compare data before and after treatment. One-way ANOVA, followed by Tukey test analysis was used to compare data from multiple groups using SigmaStat software (Jandel, San Rafael, Calif). Statistical significance was defined as $P<0.05$.

**Results**

**Inhibition of hSlo Currents by HG**

Acute exposure of hSlo expressed in HEK293 cells to medium containing NG or HG had no effects on current density or kinetics. However, when hSlo was expressed in HEK293 cells that had been exposed to HG for 1 week, reduction in current density and slowing of channel activation were observed. These changes became stable in cells after 2 weeks culture with HG. Figure 1A shows representative whole-cell currents of hSlo 72 hours after transfection in HEK293 cells that had been cultured in NG or HG for 2 weeks. HG significantly reduced current amplitudes (Figure 1A) and current/voltage (I-V) curves showed that HG reduced the maximal current density of hSlo by 47.1%, with 671.6±105.1 pA/pF (n=10) in NG and 355.6±102.3 pA/pF in HG (+200 mV; n=10; P=0.045) (Figure 1B). Normalized conductance/voltage (g-V) curves of hSlo in NG and HG were fitted using a Boltzmann equation (Figure 1C). The voltage at half-maximal activation ($V_{1/2}$) of hSlo activation was not different between NG and HG (113.5±10.8 mV in NG versus 97.5±9.2 mV in HG; n=10 for both; $P=NS$), and neither was the slope factor $k$ (34.8±1.6 mV/e-fold in NG versus 31.4±2.3 mV/e-fold in HG; n=10 for both; $P=NS$).

Single-channel recordings showed that cells in HG reduced hSlo open probability ($P_{o}$) by 60%, prolonging channel closed time constants ($\tau_c$) by 100% without changing channel open time constants ($\tau_o$) (Figure 1D; Table I in the online data supplement). HG slowed the time course of channel activation and increased the activation time constants ($\tau_a$) compared with those in NG (detailed analysis is presented in supplemental Table II).

HG also reduced hSlo tail current amplitudes (Figure 2A) and slowed the time course of channel deactivation. The activation time constant ($\tau_a$) and deactivation time constant ($\tau_d$) was fitted from the hSlo currents in NG and HG (detailed analysis of the channel deactivation kinetics is presented in Table II and Figure III of the online data supplement). The relationships between time constants ($\tau$) and voltages ($\tau$-V curve) were fitted using the voltage-dependent relaxation equation, as shown in Figure 2B. Increases in $\tau_a$ and $\tau_d$ by HG resulted in shifting the $\tau$-V curve upward. The voltage-dependent free energy ($\Delta G$) required for transition from channel closed state (C) to open state (O) was calculated by curve fitting using rate theory analysis to be 2.6 kJ/mol in NG and 4.9 kJ/mol in HG, an increase of 1.9-fold in HG, and $\Delta G$ from O to C was increased by 1.3-fold in HG (4.5 kJ/mol in NG and 5.8 kJ/mol in HG), suggesting that channel activation and deactivation transitions in HG are less favorable thermodynamically compared with those in NG. Similar results were obtained from the BK channels of human coronary artery smooth muscle cells cultured with HG (see the online data supplement). In addition, vasoreactivity studies showed that epoxyeicosatrienoic acid (EET)-induced vasodilation through BK channel activation was impaired in...
coronary arteries of streptozotocin-induced diabetic rats (Figure 2C). In controls, BK channel activation accounted for 39.3% of the 11,12-EET–mediated (1 μmol/L) vasodilation (63.8 ± 4.2% at baseline versus 24.5 ± 2.1% with 100 nmol/L iberiotoxin [IBTX]; n = 5). In diabetic vessels, BK channels contributed only 9.5% of the EET-mediated vasodilation, with 31.9 ± 2.1% dilation at baseline (n = 5; P < 0.01 versus control) and 22.4 ± 2.0% with IBTX (n = 5; P = NS versus baseline). These results suggest that BK channel dysfunction contributes to the impairment of vasodilation in diabetes (Figure 2C).

HG Produced Antioxidant Enzyme Imbalance and Increased Intracellular ROS Production in HEK293 Cells

Western blot analysis showed that CAT expression was reduced by 44% in HG cells (0.37 ± 0.04 versus 0.66 ± 0.07 in NG cells; n = 3 for both; P = 0.023), whereas CuZnSOD expression increased by 36% (0.83 ± 0.06 versus 0.61 ± 0.05 in NG cells; n = 3 for both; P = 0.048) (Figure 3A and 3B). The level of GPX-1 expression in HEK293 cells was very low. There was no significant difference in MnSOD and GPX-1 expression between NG and HG cells. This altered antioxidant enzyme profile in HG cells could result in reduced ability to scavenge H₂O₂ sufficiently, favoring the accumulation of H₂O₂.

Production of total peroxides was measured in HEK293 cells after 2 weeks in NG and HG, using the fluorescent ROS indicator CM-H₂DCF-DA (Figure 3C). Cells cultured in NG (NG cells) showed fluorescence signals of 66.9 ± 9.1 arbitrary units (n = 4) in NG medium and 124.6 ± 21.3 arbitrary units (n = 4; P = 0.02) in HG medium, suggesting HG could instantaneously increase intracellular ROS production in NG cells. However, the fluorescence signals from cells cultured in HG (HG cells) were 153.0 ± 17.3 arbitrary units when measured in NG medium (n = 4; P = 0.002 versus NG cells in NG medium), comparable to that of NG cells measured in HG medium (n = 4; P = NS). The fluorescence signals in HG cells further increased to 219.6 ± 43.8 arbitrary units when placed in HG medium (n = 4; P = 0.048 versus NG cells with HG medium). Thus, intracellular ROS increased
3.3-fold in HG cells in HG medium, compared with NG cells in NG medium. These results suggest that intracellular ROS was persistently elevated in HG cells and the ability to scavenge intracellular ROS generated was impaired. Group data are summarized in Figure 3D.

Adenoviral Expression of CAT Reversed the Effects of HG on hSlo
To determine whether the effects of HG on hSlo could be alleviated by increase in CAT activities, we overexpressed CAT by adenoviral gene transfer into HG cells before transfection of hSlo. Figure 4A shows raw tracings of hSlo 72 hours after transfection in HG cells infected with adenoviral-CAT (AdCAT). There was a 4-fold increase in CAT expression in AdCAT-infected HG cells compared with HG cells infected with control adenovirus (AdLacZ) (Figure 4B). Interestingly, hSlo current density in HG cells was enhanced by AdCAT infection (890.7 ± 125.5 pA/pF, +200 mV; n = 6) versus 355.6 ± 102.3 pA/pF in HG control [n = 10]; P = 0.006), attaining levels similar to that in NG cells (dashed line) (n = 6; P = NS) (Figure 4C). Enhanced CAT expression had no effect on the g-V curve (Figure 4D). V_{1/2} and k were 93.0 ± 8.6 mV and 36.7 ± 1.8 mV/e-fold (n = 6; P = NS versus HG control for both), respectively, in HG cells infected with AdCAT, showing no difference from those in HG control. However, CAT overexpression reduced \( \tau_\alpha \) and \( \tau_0 \) in parallel (Figure 4E) to levels comparable to those in NG. Hence, enhanced CAT activity restored hSlo current density and kinetics to that of NG cells, indicating that reduced CAT activity is the major culprit in hSlo inhibition by HG.

Exogenous H\(_2\)O\(_2\) Mimicked the Effects of HG on hSlo Kinetics
NAD(P)H oxidase is present in HEK293 cells\(^{12} \) and intracellular H\(_2\)O\(_2\) in HEK293 cells is \(<40 \mu\text{mol/L} \) under physiological conditions.\(^{30} \) Because the ROS fluorescence signal increased 3.3-fold in HG, the average cellular concentration of ROS could be raised to the \(10^{-4}\) mol/L range in HG cells. Figure 5A shows that H\(_2\)O\(_2\) dose-dependently inhibited the inside-out macroscopic currents of hSlo with an IC\(_{50}\) of 4 mmol/L. The H\(_2\)O\(_2\) effect was reversed by 2 mmol/L dithiothreitol (DTT). Hence, in subsequent experiments, we applied 4 mmol/L H\(_2\)O\(_2\) to determine the effects of H\(_2\)O\(_2\) on hSlo kinetics. Similar to HG, H\(_2\)O\(_2\) reduced hSlo current density and increased \( \tau_\alpha \) and \( \tau_0 \) in a voltage-dependent manner (Figure 5B). I-V curves normalized to baseline showed that H\(_2\)O\(_2\) reduced the maximal current density by 44.7 ± 8.1% (+200 mV; n = 3; P = 0.01 versus baseline) (Figure 5C). V_{1/2} was unaltered by H\(_2\)O\(_2\) (131.0 ± 7.0 mV at baseline versus 134.6 ± 6.3 mV with H\(_2\)O\(_2\); n = 3; P = NS.
versus baseline), as was \(k\) (17.7±2.6 mV/e-fold at baseline versus 19.3±2.3 mV/e-fold with \(H_2O_2\); \(n=3; P=NS\)) (Figure 5D). Thus, the effects of \(H_2O_2\) on hSlo channel kinetics were similar to those of HG. The \(\Delta G\) for channel activation was calculated to be 2.0 kJ/mol at baseline and 4.4 kJ/mol with \(H_2O_2\), whereas the \(\Delta G\) for channel deactivation was 4.8 kJ/mol at baseline and 5.5 kJ/mol with \(H_2O_2\). These values are similar to those obtained in HG. The \(\tau-V\) curve and kinetics parameters in response to \(H_2O_2\) are shown in Figure 5, E, in the online data supplement.

Exogenous OONO\(^-\) Inhibited hSlo Current Density but Not Channel Kinetics

Because OONO\(^-\) can also be overproduced in HG, we examined the effects of OONO\(^-\) on hSlo. We found that OONO\(^-\) (0.1 mmol/L) inhibited hSlo current density by as much as 4 mmol/L \(H_2O_2\) but did not alter channel activation or deactivation kinetics, and the effects of OONO\(^-\) were completely reversed by 2 mmol/L DTT (Figure 6A). Normalized I-V curves in the presence and absence of 0.1 mmol/L OONO\(^-\) are shown in Figure 6B. OONO\(^-\) inhibited the maximal hSlo activity by 31.4±16.7% (\(n=4; P=0.01\)), comparable to the effects of 4 mmol/L \(H_2O_2\) and HG (\(n=4; P=NS\)). Neither channel \(V_{1/2}\) (162.8±11.8 mV at baseline and 170.5±13.1 mV with OONO\(^-\); \(n=4; P=NS\)) nor \(k\) (21.5±1.7 mV/e-fold at baseline and 22.0±1.4 mV/e-fold with OONO\(^-\); \(n=4; P=NS\)) was altered by OONO\(^-\). Unlike HG and \(H_2O_2\), OONO\(^-\) had no effect on \(\tau_a\) and \(\tau_d\) (Figure 6C). A comparison between the effects of \(H_2O_2\) and OONO\(^-\) on hSlo channel kinetics is given in Figure VI, D, and Table II in the online data supplement.

hSloC911A Mutant Was Insensitive to \(H_2O_2\) and HG

Because the effects of HG on hSlo were mainly mediated by \(H_2O_2\) in HEK293 cells, we determined the role of cysteine 911 in HG- and \(H_2O_2\)-mediated hSlo modulations. hSlo with a single substitution at cysteine 911 by alanine (hSloC911A) was expressed in HEK293 cells. The time course of the effects of 4 mmol/L \(H_2O_2\) and 0.1 mmol/L OONO\(^-\) on hSloC911A inside-out macroscopic currents is illustrated in Figure 7A (top and bottom, respectively). Relative to hSlo wild type (wt), hSloC911A was insensitive to \(H_2O_2\) inhibition, although \(H_2O_2\) produced a very small but still significant inhibition in current density and the effect was reversed by 2 mmol/L DTT. However, the effects of OONO\(^-\) on
hSloC911A were unchanged from wt, suggesting that C911 is the molecular site for H2O2 modulation, but not for OONO−. Moreover, H2O2 failed to slow hSloC911A activation or deactivation (Figure 7B and 7D) and only produced a maximal current inhibition of 14.3±2.4% (+200 mV; n=4; P=0.048) (Figure 7C), compared with 44.7±8.1% in wt (n=4; P=0.011). H2O2 did not alter the channel V1/2 (30.3±16.1 mV at baseline and 137.3±27.4 mV with H2O2; n=4; P=NS) or k (17.7±2.6 mV/e-fold at baseline and 19.7±2.3 mV/e-fold with H2O2; n=4; P=NS). H2O2 did not

Figure 4. Effects of HG on hSlo kinetics after AdCAT infection. A, Raw current tracings of hSlo were recorded from cells in HG after AdCAT infection. AdCAT infection restored hSlo current density and channel kinetics in HG. B, Western blots showing that HG cells had a 4-fold increase in CAT expression 72 hours after AdCAT infection compared with those infected with AdLacZ. C, hSlo I-V curves in HG cells after AdCAT (●) and AdLacZ infection (○) (n=6). Dashed line represents the I-V curve of NG. *P<0.05 vs AdLacZ. D, Normalized g-V curves in cells infected with AdCAT (●) and AdLacZ (○) (n=6). E, τ-V curves from cells infected with AdCAT (●) and AdLacZ (○). Overexpression of AdCAT accelerated and normalized hSlo activation and deactivation kinetics in HG.

Figure 5. Exogenous H2O2 reproduced the effects of HG on hSlo kinetics. A, Time course of inside-out macroscopic currents of hSlo, recorded at +160 mV with a holding potential of −60 mV, showing the effects of various concentrations of H2O2 and 2 mmol/L DTT applied to the cytoplasmic surface of hSlo. IC50 of H2O2 on hSlo currents was ~4 mmol/L. B, Application of 4 mmol/L H2O2 suppressed hSlo current density and slowed its activation and deactivation kinetics. C, hSlo I-V curves at baseline (○) and after exposure to H2O2 (●) (n=4). Results were normalized to maximum current amplitude at baseline. D, τ-V curves composed of τ0 (~140 mV to +100 mV) and τ1 (~100 mV to +210 mV) before (○) and after (●) exposure to H2O2. H2O2 shifted the τ-V curves upwards, similar to the effects of HG. *P<0.05 vs baseline.
change the ΔG of hSloC911A activation (2.8 kJ/mol at baseline and 3.0 kJ/mol with H₂O₂) or deactivation (4.5 kJ/mol at baseline and 4.8 kJ/mol with H₂O₂). There was no difference between the current densities, V₁/₂, and channel kinetics of hSloC911A in cells in NG and HG, which were similar to wt recordings in NG (Figure 8). Hence, the C911A mutation prevented downregulation of channel activities in HG (926.0/11006193.2 pA/pF, 200 mV [n=6] versus 355.6±102.3 pA/pF in wt [n=10]; P=0.008) and maintained normal -V curves. The ΔG for hSloC911A activation and deactivation in HG was 2.9 kJ/mol and 4.9 kJ/mol, respectively, similar to those of wt channel in NG. Our results indicate that H₂O₂ is the major ROS produced by HEK293 cells in HG and C911 is the major molecular site of redox regulation in hSlo by HG.

Discussion
In this study, we have made several important findings. First, HG reduced hSlo current density and altered channel kinetics. Second, HG enhanced SOD expression but suppressed CAT expression, resulting in a 3.3-fold increase in ROS generation in HEK293 cells. Third, H₂O₂ but not OONO⁻ mimicked the effects of HG on hSlo channel kinetics. Fourth, hSloC911A was insensitive to HG or H₂O₂ modulation, suggesting that C911 in hSlo is the major molecular target of redox regulation by HG. We believe this is the first report identifying the molecular mechanism associated with impaired BK channel function in hyperglycemia.

Vascular K⁺ channels are effectors of endothelium-derived relaxation factors (EDRF) and endothelium-derived hyperpolarizing factors (EDHF) and are critical determinants of vascular tone.⁴,³¹ Impaired BK channel–mediated vasodilation is known to be associated with vascular dysfunction in diabetic animals. However, the mechanisms through which vascular BK channels are modulated by hyperglycemia have not been delineated. In high fructose diet–induced, insulin-resistant rats, BK channel density in mesenteric smooth
muscle cells was reduced, but the channel $\text{Ca}^{2+}$ sensitivity and the voltage sensitivity were unchanged. In early-stage type 2 diabetic rats, impaired coronary arterial smooth muscle BK channel activation by arachidonic acid was attributable to a decrease in prostacyclin bioavailability. In this study, we demonstrated that enhanced $\text{H}_2\text{O}_2$ formation in HEK293 cells cultured with HG directly inhibited channel activity through redox modulation at C911 on the BK channel subunit. HG not only directly enhanced NAD(P)H oxidase, but also interrupted the mitochondria electron transport chain at complex III, resulting in accumulation of $\text{O}_2^\cdot$, which in turn formed $\text{H}_2\text{O}_2$ and OONO$. However, hSlo activity was not inhibited by acute exposure to HG but by prolonged culture in HG that reached stable effects after 14 days. This time frame was required for HG to produce the imbalance in SOD and CAT activities, resulting in the accumulation of $\text{H}_2\text{O}_2$. This was supported by direct measurements of intracellular ROS, showing a 3.3-fold increase in HEK293 cells after 14 days of culture in HG. We believe that the major ROS product in our system was $\text{H}_2\text{O}_2$. First, SOD expression was increased by 36%, whereas CAT expression was decreased by 44% in HG cells, hence resulting in net $\text{H}_2\text{O}_2$ accumulation. Second, gene transfer of CAT into HG cells abolished the HG effects on hSlo current density and channel kinetics. Third, the application of $\text{H}_2\text{O}_2$, but not OONO$, mimicked the effects of HG, suggesting that OONO$ was not the key ROS participant in modulating hSlo properties in HEK293 cells. Whereas BK channel kinetics were not affected by OONO$, we found that 0.1 mmol/L OONO$ produced as much hSlo current density inhibition as 4 mmol/L $\text{H}_2\text{O}_2$. Because OONO$ is highly reactive and generated by both endothelial and vascular smooth muscle cells, we believe that in diabetic vessels, BK channels are also modulated by OONO$. Why relatively high concentrations of exogenous $\text{H}_2\text{O}_2$ are required to reproduce the HG results is not immediately fully understood. We believe that $\text{H}_2\text{O}_2$ acts directly on hSlo and not on membrane lipids because a single C911A mutation is sufficient to eliminate the HG and $\text{H}_2\text{O}_2$ effects.

The mechanism underlying BK channel regulation by $\text{H}_2\text{O}_2$ is complex. A putative EDHF, $\text{H}_2\text{O}_2$ has been shown to activate vascular BK channels directly or through second messenger signaling pathways, leading to vasodilation. However, activation of BK channels by $\text{H}_2\text{O}_2$ is observed only when applied extracellularly, whereas intracellularly applied...
H₂O₂ inhibits BK channels.¹⁴,¹⁶,¹⁷ Hence, the paracrine and autocrine functions of H₂O₂ on BK channels have divergent outcomes. Contrary to its physiological signaling role, prolonged excessive production of H₂O₂ is detrimental to the maintenance of cellular homeostasis, resulting in the development of diseases such as diabetes and arteriosclerosis. Cysteine modification of hSlo is involved in channel regulation by H₂O₂. However, the molecular identity of the cysteine residue in hSlo that is the functional target of redox modulation has been controversial. Tang et al reported that C⁹¹¹ near the Ca²⁺/H¹¹₀₀₁ bowl in the RCK2 domain is the major target of cysteine-modification reagents such as MTSEA and of cytoplasmic application of H₂O₂.¹⁴ Oxidation of C⁹¹¹ also reduces channel Ca²⁺ sensitivity as a result of an increase in the free energy for Ca²⁺ binding, which was only observed in the presence of high free Ca²⁺ (≥1 μmol/L).¹⁴ Zhang and Horrigan showed that C⁴³⁰ in the RCK1 domain but not C⁹¹¹ was the functional residue modified by oxidation.²¹ The crystal structure of the RCK domains on the Ca²⁺/H¹¹₀₀₁ channel, MthK, shows that RCK1 is coupled to the channel pore through the M2 transmembrane domain, which corresponds to S6 in the BK channel.⁷ Thus, oxidation of C⁴³⁰ may impair RCK1 movement and reduce channel Ca²⁺ sensitivity.²¹ Both studies presented strong evidence that a single mutation virtually eliminates the effects of the cysteine-modification reagents on hSlo channel Ca²⁺ sensitivity and voltage sensitivity. H₂O₂ only efficiently oxidizes cysteine residues that are deprotonated, whereas most cysteine residues in proteins have a pKa of 8.5, so they would not be targeted by H₂O₂ unless they are located in the vicinity of highly positively charged residues. The Ca²⁺ bowl contains a high density of aspartic acids surrounded by positively charged amino acids. This critical location may explain why C⁹¹¹ is the prime target for H₂O₂ modulation. Our results show that HG and H₂O₂ inhibit hSlo mainly through C⁹¹¹ oxidation, because the C⁹¹¹A mutation abrogated the effects of HG and eliminated 80% of the H₂O₂ effects. The effects of cysteine-modifying reagents such as MTSET, MTSES, and NEM on hSlo channel kinetics can shift the steady-state I-V curves either leftward or rightward, depending on whether the channel activation or deactivation gate is modified by these reagents.²¹ Slowing channel activation can shift the steady-state I-V or g-V curves leftward, whereas slowing channel deactivation favors channels to stay in open state and shifts the curves rightward. We found that HG and H₂O₂ do not shift the I-V curve, perhaps because of a balanced slowing of channel activation and deactivation by HG and H₂O₂. Another explanation is that hSlo currents were recorded with 200 nmol/L intracellular free Ca²⁺, which may be insufficient to change the free energy of Ca²⁺ binding.¹⁴

To summarize, we have provided compelling evidence that HG inhibits BK channel function through H₂O₂ oxidation of C⁹¹¹. These results indicate that diabetes alters antioxidant enzyme profiles, enhances ROS generation and compromises BK channel function. Gene transfer of CAT ameliorates these functional impairments and has therapeutic potential in the treatment of diabetic vascular dysfunction.
Acknowledgments

We thank Dr Larry W. Oberley (The University of Iowa, Iowa City) for providing AdCAT. The hSloC911A mutant was kindly provided by Dr Toshinori Hoshi (University of Pennsylvania, Philadelphia).

Sources of Funding

This work was supported by grants from the NIH (HL-63154 and HL-74180) and the American Heart Association (0265472Z).

Disclosures

None.

References


Disclosures

None.

References

Molecular Mechanisms Mediating Inhibition of Human Large Conductance Ca$^{2+}$-Activated K$^{+}$ Channels by High Glucose

Tong Lu, Tongrong He, Zvonimir S. Katusic and Hon-Chi Lee

_Circ Res._ 2006;99:607-616; originally published online August 24, 2006; doi: 10.1161/01.RES.0000243147.41792.93

_Circulation Research_ is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2006 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7330. Online ISSN: 1524-4571

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circres.ahajournals.org/content/99/6/607

Data Supplement (unedited) at:
http://circres.ahajournals.org/content/suppl/2006/08/24/01.RES.0000243147.41792.93.DC1

**Permissions:** Requests for permissions to reproduce figures, tables, or portions of articles originally published in _Circulation Research_ can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

**Reprints:** Information about reprints can be found online at:
http://www.lww.com/reprints

**Subscriptions:** Information about subscribing to _Circulation Research_ is online at:
http://circres.ahajournals.org/subscriptions/
Molecular Mechanisms Mediating Inhibition of Human Large Conductance $\text{Ca}^{2+}$-Activated $\text{K}^+$ Channels by High Glucose

Tong Lu, MD., PhD.*
Tongrong He, MD., PhD.†
Zvonimir S. Katusic, MD., PhD.†
Hon-Chi Lee, MD., PhD.*
METHODS

hSlo and CAT cDNA transfection in HEK 293 cells

hSlo wt and C911A mutant cDNA in pTracerCMV-2 were transfected into HEK293 cells using a FuGENE 6 transfection kit as previously described \(^1\). The iberiotoxin (IBTX, 100 nmol/L)-sensitive component (BK currents) accounts for at least 90% of total K\(^+\) currents in HEK293 cells 72 h after transfection (Figure 1 supplement). AdLacZ (2x10\(^{10}\) pfu/ml) containing the histochemical marker β-galactosidase, and AdCAT (9x10\(^{10}\) pfu/ml) carrying the catalase gene were transfected into HEK293 cells with a multiplicity of infection (MOI) of 50 for 4 h before hSlo cDNA was added to the culture medium \(^2\).

Vasoreactivity Measurements

Hearts were rapidly excised and placed in ice-cold Krebs solution that contained (in mmol/L): NaCl 118.3, KCl 4.7, CaCl\(_2\) 2.5, MgSO\(_4\) 1.2, KH\(_2\)PO\(_4\) 1.2, NaHCO\(_3\) 25, and dextrose 11.1. The secondary and tertiary branches (50-200 μm in intraluminal diameter) of the right and left coronary arteries from the epicardial surface, as well as branches of the septal coronary arteries were carefully dissected and isolated free of surrounding myocardium and connective tissue under a dissecting microscope (Olympus SZ4045 Stereo Microscope, Olympus America Inc., Melville, NY). Isolated small coronary arteries (1-2 mm in length) were transferred to a custom-made vessel chamber filled with Krebs solution. The arteries were mounted and secured between two borosilicate glass micropipettes (30 μm diameter tips) with 10-O ophthalmic suture. The lumen of the vessel was filled with Krebs solution through the micropipettes and
maintained at a constant pressure (no flow) of 60 mmHg. The vessel chamber was transferred to an inverted light microscope stage (Olympus CK40) coupled to a video measurement system (VIA-100, Boeckeler Instruments, Inc., AZ) equipped with a video camera, monitor and calibrated video calipers for visualization and recording of the intraluminal diameter as described previously. Vessels were equilibrated for at least 30 min in oxygenated (20% O₂, 5% CO₂, balanced with N₂, 37°C) Krebs solution, which was continuously circulated through the vessel bath. Vessels were unacceptable for experiments if they demonstrated leaks, failed to produce >30% constriction to 60 mmol/L KCl or to graded doses of endothelin-1 (ET-1), or failed to produce 80% dilation with nitroprusside (100 µmol/L).

All compounds were added abluminally. During a 30 min equilibration period, some vessels were pretreated with 100 nmol/L of iberiotoxin (IBTX). Vessels were then constricted to 30-60% of baseline diameter with endothelin-1 and the effect of 1 µM 11,12-epoxyeicosatetraenoic acid (11,12-EET) on vessel diameter was measured.

**Electrophysiology**

hSlo transfected cells were identified by GFP expression under a UV microscope (Olympus, IX70). Whole-cell and inside-out single channel currents were recorded with an integrating amplifier (Axon patch 200B, Axon Instruments) and filtered with an 8-pole low pass Bessel filter (902 LPF, Frequency Devices Inc.) with a bandwidth (-3 dB) of 2 kHz and sampling rate of 50 kHz (12-bit resolution). Pipette resistance when filled with the pipette solution was 0.5~1 MΩ for whole-
cell and macroscopic current recordings, and 8~10 MΩ for single channel recordings. The pipette solution for whole-cell recordings contained (in mmol/L): KCl 140, MgCl₂ 0.5, Na₂ATP 5.0, Na₂GTP 0.5, HEPES 1.0, EGTA 1.0, CaCl₂ 0.465 (200 nmol/L free Ca²⁺), pH 7.35. The pipette solution for inside-out macroscopic current recordings contained (in mmol/L): NaCl 145, KCl 5.6, MgCl₂ 1.0, CaCl₂ 0.5, HEPES 10, pH 7.4. The bath solution for whole-cell recordings contained (in mmol/L): NaCl 145, KCl 5.6, MgCl₂ 1.0, CaCl₂ 0.5, HEPES 10, glucose 5.0 or 22, pH 7.4. The bath solution for inside-out macroscopic current recordings contained (in mmol/L): KCl 140, MgCl₂ 0.5, HEPES 1.0, EGTA 1.0, CaCl₂ 0.465, pH 7.35. The pipette solution for single channel recording contained (in mmol/L) KCl 140, CaCl₂ 1.0, HEPES 10, MgCl₂ 0.5, pH 7.4. The bath solution for single channel recording contained (in mmol/L): KCl 140, HEPES 10, MgCl₂ 1.0, EGTA 1.0, CaCl₂ 0.685 (free Ca²⁺~0.5 μM), pH 7.35.

To determine channel activation, hSlo currents were elicited from a holding potential of -60 mV with 50 ms duration testing pulses from -40 mV to +210 mV in 10 mV increments. To determine channel deactivation, hSlo tail currents were recorded using a protocol with a holding potential of -60 mV and a conditioning pulse of 50 ms at +160 mV, followed by testing potentials from -140 mV to +90 mV in 10 mV increments.

The conductance-voltage relationship (g-V curve) and the current-voltage relationship (I-V curves) of hSlo channels were characterized by the Boltzmann equation:

\[
g / g_{\text{max}} = g \left\{ \frac{1}{1 + \exp \left[ \frac{V_{1/2} - V_m}{k} \right]} \right\}
\]
where \( g_{\text{max}} \) represents the maximal \( K^+ \) conductance (g), \( V_{1/2} \) is the voltage at which g reaches half of \( g_{\text{max}} \), \( V_m \) is the membrane potential, and \( k \) is the slope factor associated with the Boltzmann distribution.

Channel activation (\( \tau_A \)) and deactivation (\( \tau_D \)) time constants were fitted using a single exponential function. According to a two-state (closed state ↔ open state) model, the time constant and voltage relationship (\( \tau-V \) curve) was composed of \( \tau_D \) (−140 mV to +60 mV) and \( \tau_A \) (+70 mV to +210 mV) and was fitted using the voltage-dependent relaxation equation:

\[
\tau (V) = \frac{1}{\alpha \exp \left( \frac{V z F}{RT} \right) + \beta \exp \left( \frac{-V z F}{RT} \right)}
\]

where \( \alpha_0 \) represents the forward rate constant from closed state to open state, and \( \beta_0 \) represents the backward rate constant from open state to closed state at 0 mV. \( z_\alpha \) and \( z_\beta \) represent the forward and the backward charge movements. \( V \) is the membrane potential, \( F \) is the Faraday constant, \( R \) is the universal gas constant, and \( T \) is the absolute temperature.

The free energy (\( \Delta G \)) required to overcome the energy barrier during transitions between closed-state and open-state was calculated using rate theory analysis:

\[
1 / \tau = k_B T / h \exp(-\Delta G \pm z \delta F / 2RT)
\]

where \( k_B \) is the Boltzmann constant, \( T \) is the absolute temperature, \( h \) is the Planck constant, \( z \) is the charge movement, and \( \delta \) is the position of the energy barrier with a single binding site in membrane, assuming the peak is halfway at an electrical distance (\( \delta / 2 \)) from the beginning. \( V \) is the membrane potential, \( F \) is the Faraday constant, and \( R \) is the universal gas constant.
Single channel kinetics were analyzed using TAC software (Bruxton, Inc.) The components of channel open times ($\tau_o$) and closed time constants ($\tau_c$) were fitted with the sums of exponential probability density functions using the maximum likelihood with simple optimization. Any additional exponential component was added only when the probability exceeded 0.95.$^7$

RESULTS

Effects of high glucose in native BK channel activities
We examined the effects of HG on native IBTX-sensitive BK currents of human coronary artery smooth muscle cells, as shown in Figure 2 supplement. Similar to expressed hSlo channels, HG reduced BK current densities from $63.3 \pm 7.6$ pA/pF (+200 mV, n=3) in NG to $38.1 \pm 5.2$ pA/pF (+200 mV, n=4, p=0.036), and slowed the channel activation and deactivation time courses.

Effects of high glucose on hSlo channel activation kinetics
HG not only reduced hSlo current density but also slowed the time course of channel activation and deactivation (tail currents) compared to NG (Figure 3A supplement). The activation time constant ($\tau_A$) was increased from 0.72 ms in NG to 1.87 ms in HG at +200 mV, and from 1.41 ms in NG to 2.27 ms in HG at +100 mV. There was no difference between NG and HG in $\tau_A$ at voltages below +50 mV. The reciprocal of $\tau_A$ ($1/\tau_A$) as a function of voltage in NG and HG (n=10) is shown in Figure 3B supplement.

Effects of high glucose on hSlo channel deactivation kinetics
The hSlo current deactivation time constant ($\tau_D$) was fitted from the hSlo tail currents in NG and HG. HG reduced hSlo tail current amplitudes (Figure 2A in
text) and slowed the time course of channel deactivation (Figure 4A supplement).

$\tau_D$ increased from 0.5 ms in NG to 2.54 ms in HG at +90 mV, and from 0.48 ms to 1.54 ms at +40 mV; there was no difference between NG and HG at voltages below 0 mV. The finding that $\Delta G$ is increased for channel transitions from open state to closed state and from closed state to open state indicated that hSlo has to overcome higher energy barriers in HG both for channel activation and deactivation (Figure 4B supplement).
Figure 1 (supplement): hSlo channel expression in HEK293 cells

Continuous recordings of whole-cell K⁺ currents elicited at +100 mV from a holding potential of -60 mV (left panel) show the response to 100 nmol/L IBTX, a specific BK channel blocker. The time course of the IBTX effects is displayed in the right panel. The IBTX-sensitive component (BK currents) accounts for at least 90% of total K⁺ currents in HEK293 cells 72 h after transfection.
Figure 2 (supplement): Effects of high glucose on native BK channels of cultured human coronary artery smooth muscle cells

A: Representative IBTX-sensitive BK currents recorded from human coronary artery smooth muscle cells cultured with NG and HG for 2 weeks. B: Normalized BK currents elicited at +200 mV in NG and HG. C: I-V curves showing voltage-dependent inhibition of BK channel density by HG. * p<0.05 vs. NG.
Figure 3 (supplement):  A: Normalized hSlo current activation recorded at +50 mV, +100 mV and +200 mV in NG (solid lines) and HG (dashed lines). HG slowed the time course of channel activation at voltages greater than +50 mV. B: $1/\tau_A$ of NG (○) and HG (●) plotted as a function of voltage. The solid lines represent the best fit according to the rate theory model with a single binding site. * p<0.05 vs. NG.
Figure 4 (supplement): A: Normalized hSlo tail currents at –10 mV, +40 mV and +90 mV in NG (solid lines) and in HG (dashed lines). HG slowed the time course of channel deactivation in a voltage-dependent manner. B: Plots of $1/\tau_D$ as a function of voltage in NG (○) and HG (●) cells. The solid lines represent the best fit according to rate theory analysis. * p<0.05 vs. NG.
Table 1 (supplement)  Effects of high glucose on the open time and closed time constants of hSlo wt single channel kinetics

<table>
<thead>
<tr>
<th></th>
<th>Open Times Constants $\tau_O$ (ms)</th>
<th>Closed Time Constant $\tau_C$ (ms)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$\tau_{O1}$ ($A_{O1}$)</td>
<td>$\tau_{O2}$ ($A_{O2}$)</td>
</tr>
<tr>
<td>NG</td>
<td>2095</td>
<td>0.66</td>
</tr>
<tr>
<td></td>
<td>(0.16)</td>
<td>(0.64)</td>
</tr>
<tr>
<td>HG</td>
<td>2142</td>
<td>0.66</td>
</tr>
<tr>
<td></td>
<td>(0.24)</td>
<td>(0.61)</td>
</tr>
</tbody>
</table>

$\tau_{O1}$, $\tau_{O2}$ and $\tau_{O3}$ represent the slow, intermediate and fast components of channel open time constants and $A_{O1}$, $A_{O2}$ and $A_{O3}$ represent the corresponding weights respectively. Similarly, $\tau_{C1}$, $\tau_{C2}$, $\tau_{C3}$ and $\tau_{C4}$ represent the very slow, slow, intermediate and fast components of channel closed time constants. $A_{C1}$, $A_{C2}$, $A_{C3}$ and $A_{C4}$ are the relative weights. Here, $A_{O1}+A_{O2}+A_{O3}=1$ and $A_{C1}+A_{C2}+A_{C3}+A_{C4}=1$. Compared to NG, culture with HG prolonged each components of $\tau_C$ without changing the $\tau_O$ components.
Table 2 (supplement)  Activation and deactivation kinetics of hSlo wt and C911A mutant channels in response to high glucose, H$_2$O$_2$, OONO$^-$, and high glucose with CAT over-expression

<table>
<thead>
<tr>
<th></th>
<th>NG</th>
<th>HG</th>
<th>CAT+HG</th>
<th>C911A+HG</th>
<th>Baseline</th>
<th>H$_2$O$_2$</th>
<th>OONO$^-$</th>
<th>C911A+H$_2$O$_2$</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>whole-cell recordings</td>
<td>inside-out recordings</td>
<td></td>
</tr>
<tr>
<td>$\alpha_0$ (s$^{-1}$)</td>
<td>235.5</td>
<td>187.5</td>
<td>298.7</td>
<td>275.6</td>
<td>109.8</td>
<td>88.5</td>
<td>113.7</td>
<td>164.2</td>
</tr>
<tr>
<td>$\beta_0$ (s$^{-1}$)</td>
<td>800.6</td>
<td>758.7</td>
<td>926.8</td>
<td>1024</td>
<td>981.5</td>
<td>686.8</td>
<td>875.4</td>
<td>883.7</td>
</tr>
<tr>
<td>$z_\alpha$ (e)</td>
<td>0.65</td>
<td>0.52</td>
<td>0.71</td>
<td>0.59</td>
<td>0.49</td>
<td>0.42</td>
<td>0.48</td>
<td>0.46</td>
</tr>
<tr>
<td>$z_\beta$ (e)</td>
<td>0.43</td>
<td>0.59</td>
<td>0.38</td>
<td>0.47</td>
<td>0.53</td>
<td>0.58</td>
<td>0.50</td>
<td>0.54</td>
</tr>
</tbody>
</table>

$\alpha_0$ represents the forward rate constant from closed to open state, $\beta_0$ represents the backward rate constant from open to closed state in a 2 state model, $z_\alpha$ represents the charge movement from closed state to open state, and $z_\beta$ represents the charge movement from open state to closed state.
REFERENCES


7. Lu T, Katakam PV, VanRollins M, Weintraub NL, Spector AA, Lee HC. Dihydroxyeicosatrienoic acids are potent activators of Ca(2+)-activated