L-Type Calcium Channels
Highs and New Lows

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Voltage-gated calcium channels are essential for coupling membrane depolarization to the influx of calcium in all excitable cells. The calcium that flows into excitable cells through voltage-gated calcium channels serves a dual function, generating both electrical and chemical signals. The intracellular events controlled by calcium are diverse and many. Excitable cells can select from a number of functionally distinct voltage-gated Ca\textsuperscript{2+} channel subunits, whose activities are precisely tuned to support specific tasks. These include excitation-contraction coupling in muscle, excitation-secretion coupling in neurons, hair cells, and endocrine cells, and regulation of gene expression. Ten genes encode the main Ca\textsubscript{v}1.3 subunit of the voltage-gated calcium channel complex in mammals. Sequence comparisons among Ca\textsubscript{v}1.3 genes from several genomes reveal three major families, Ca\textsubscript{v}1\alpha\textsubscript{1}, Ca\textsubscript{v}2\alpha\textsubscript{1}, and Ca\textsubscript{v}3\alpha\textsubscript{1}.

Even before the availability of selective toxins, several investigators demonstrated that multiple, functionally distinct classes of voltage-gated calcium channels are expressed in a variety of cell types including heart. This division was based on the presence of two distinct classes of calcium channels that differed significantly in their voltage dependence of activation. The concept of low voltage-activated and high voltage-activated calcium channels was established, and although simple, this remains a useful and informative way for distinguishing among different classes of calcium channels.

Certain features have emerged from studies of voltage-gated calcium channels in heart and neurons that have established a set of standard criteria to define the presence of a specific Ca\textsuperscript{2+} channel subtype. Low voltage-activated, T-type, Ca\textsuperscript{2+} channels that contain Ca\textsubscript{v}3\alpha\textsubscript{1} subunits (\alpha\textsubscript{5G, \alpha\textsubscript{1H, \alpha\textsubscript{T}}) activate rapidly, deactivate slowly, exhibit pronounced voltage-dependent inactivation, and are insensitive to dihydropyridines and several other toxins that inhibit neuronal calcium channels. Evidence supporting this conclusion is presented in both the Striessnig and Chiamvimonvat studies by comparing properties of native currents in wild-type and Ca\textsubscript{v}1.3\alpha\textsubscript{1} knockout mice. Others studies characterizing the functional properties of recently cloned Ca\textsubscript{v}1.3\alpha\textsubscript{1} subunits isolated from neurons and endocrine cells provide additional support.

Striessnig and colleagues recorded from inner hair cells of the cochlea of Ca\textsubscript{v}1.3\alpha\textsubscript{1} mouse and showed selective loss of a low-threshold activating Ca\textsuperscript{2+} current. From this they inferred the presence of a similar current in SA node cells to explain the observed abnormalities in pacemaking in the same mice. Chiamvimonvat and colleagues now test this hypothesis directly by recording from the SA node and from isolated cells of wild-type and Ca\textsubscript{v}1.3\alpha\textsubscript{1} mice. As reported in this issue of Circulation Research, the absence of Ca\textsubscript{v}1.3\alpha\textsubscript{1} is associated with a reduced rate of SA node firing, diastolic depolarization rate slowing at relatively hyperpolarized voltages (−40 and −45 mV), and the loss of calcium current in isolated SA node cells that activates at relatively hyperpolarized membrane potentials. These new studies offer strong support that Ca\textsubscript{v}1.3\alpha\textsubscript{1} ablation, SA node dysfunc-

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tion, and the loss of a low-threshold activating Ca$^{2+}$ current in SA node cells are intimately linked.

Do all L-type Ca$^{2+}$ channels that contain Ca$_{1.3}$ subunits activate at hyperpolarized voltages? The answer is probably yes, based on recent functional analyses of recombinant Ca$_{1.3}$ channels. The Figure compares peak current voltage relationships of Ca$_{1.3}$ L-type channels to high voltage-activated Ca$_{1.2}$ L-type, and to low voltage-activated Ca$_{3.1}$ T-type channels. The large difference in voltage dependence of activation between the two L-type Ca$^{2+}$ channels is as striking as the similarity in the activation thresholds of Ca$_{1.3}$ L-type and Ca$_{3.1}$ T-type channels. While properties of calcium channels are influenced by several factors including association with specific auxiliary subunits, the similar features of Ca$_{1.3}$ subunits cloned from different tissues, combined with two gene ablation studies in mice, favor the conclusion that low voltage-dependent activation is an intrinsic feature of Ca$_{1.3}$-containing L-type Ca$^{2+}$ channels. Clearly, significant functional differences exist among L-type Ca$_{1,3}$ genes.

If Ca$_{1.3}$-containing L channels activate at hyperpolarized membrane potentials, it is rather surprising that this feature has not been highlighted in previous studies of cloned and heterologously expressed channels. Although other factors almost certainly influence channel properties, the concentration of extracellular divalent cation has large effects on the voltage dependence of activation as a result of charge screening and is a factor that differs significantly among studies. For unknown reasons, achieving high expression levels from Ca$_{1.3}$ clones has until recently been problematic. To compensate for low current densities, concentrations of extracellular calcium and barium up to 40 mmol/L have been used. As suggested by Zhang et al., this likely contributes to the discrepancy between properties of recombinant Ca$_{1.3}$ channels and the activation range expected from functional analyses of native currents in SA node cells. The use of high concentrations of extracellular divalent cations in earlier studies of cloned channels probably obscured the unusually hyperpolarized activation range of Ca$_{1.3}$ L channels. It is notable that the Ca$_{1.3}$ L-type current-voltage relationship is shifted toward voltages $\approx$20 mV more depolarized and into the range of a high voltage-activated L-type channel when 40 mmol/L barium is used.

Future studies will be needed to address the relative importance of Ca$_{1.3}$-containing L-type Ca$^{2+}$ channels in pacemaking in the heart. Although Ca$_{1.3}$ mRNA is present in atrial myocytes, recent studies suggest that levels are very low in the SA node, particularly when compared with Ca$_{3.1}$ T-type mRNA. The availability of a selective inhibitor of Ca$_{1.3}$-containing L channels would prove a useful tool to determine the relative contribution of this channel to SA node function. Classic L-type Ca$^{2+}$ channel blockers are not useful in this regard. Recent studies of recombinant Ca$_{1.3}$ L-type channels suggest a relatively low sensitivity to block by dihydropyridines compared with Ca$_{1.2}$ L-type channels. It will be of interest to establish whether a unique splice isoform of Ca$_{1.3}$ is expressed in the SA node. There is evidence for some level of atrial-specific splicing of Ca$_{1.3}$ RNA in the S3–S4 linker of domain IV of the channel. Splicing at this site shifts the voltage dependence of activation by $<$10 mV and does not seem to influence dihydropyridine binding. Finally, given the emphasis placed on similarities between Ca$_{1.3}$ and T-type Ca$^{2+}$ channels in terms of their activation thresholds, it is worth noting features that distinguish these channels. Whereas T-type Ca$^{2+}$ channels undergo prominent voltage-dependent inactivation, Ca$_{1.3}$ L-type Ca$^{2+}$ channels show weak voltage-dependent, but strong calcium-dependent, inactivation. Further, Ca$_{1.3}$ L-type Ca$^{2+}$ channels deactivate rapidly compared with T-type Ca$^{2+}$ channel subtypes that dominate in heart.

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