Interaction of Bretylium Tosylate with Rat Cardiac Muscarinic Receptors
Possible Pharmacological Relevance to Antiarrhythmic Action

Gabriel Schreiber, Menachem Friedman, and Mordechai Sokolovsky
From the Department of Biochemistry, George S. Wise Faculty of Life Sciences, Tel-Aviv University, Tel-Aviv, Israel

SUMMARY. The interaction of the antifibrillatory antiarrhythmic drug, bretylium tosylate, with the muscarinic receptor in tissue homogenates from regions of rat brain and heart was investigated. Competition-binding experiments were carried out with the highly specific tritiated antagonist N-methyl-4-piperidyl benzilate. Bretylium tosylate competitively displaced the labeled antagonist from the muscarinic receptor. The binding of the drug to the two brain preparations was found to be best fitted by a one-site model in each case. On the other hand, in the case of both heart preparations, a two-site model yielded a significantly better fit for the binding data than that given by a single-site model. The low affinity-binding constants in the atrium and the ventricle were similar (~10 μM) to those in the brain regions examined, namely, the cortex and the medulla-pons. Sites with relatively higher affinity for the drug were detected in the heart only, with equilibrium-binding constants of 0.24 ± 0.12 μM and 0.97 ± 0.27 μM for the atrium and the ventricle, respectively. The drug also exerted anti-acetylcholine activity (Ki = 14 ± 2 μM) measured physiologically in the guinea pig atrium, which correlated well with the concentration of the drug observed to be efficacious clinically (~10 μM). (Circ Res 55: 653-659, 1984)

BRETYLIUM tosylate* (scheme 1), which was first introduced as an antihypertensive agent, but later rejected because of rapid development of tolerance, is an effective antiarrhythmic drug, recommended for treatment of refractory ventricular arrhythmias (Kumana and Hamer, 1979). The drug has profound effects on the sympathetic nervous system: it is taken up and concentrated in adrenergic nerve terminals (Iversen, 1973), initially releasing norepinephrine (Gilmore and Siegel, 1962), and, later, preventing its release (Boura and Green, 1959). Bacaner (1966) reported that bretylium tosylate increased the current threshold for electrical induction of ventricular fibrillation in dogs. Subsequent experimental (Logic, 1973; Knifin et al., 1975) and clinical (Bernstein and Koch-Weser, 1972; Sanna and Arcidiacono, 1978; Bigger et al., 1971; Cardinal and Sasyniuk, 1978) evaluations of the drug have confirmed its unique antifibrillatory action, while also demonstrating that it exerts various direct electrophysiological effects (Wit et al., 1970; Bigger et al., 1971; Cardinal and Sasyniuk, 1978). It was assumed that the drug did not affect the cardiac muscarinic cholinergic receptors (Bigger and Hoffman, 1980). In view of the diversity of the effects of bretylium tosylate and its complex interaction with the sympathetic nervous system, it was of interest to evaluate any effect it might exert directly on the para-

Methods

Materials

[3H]-4NMPB, 69.7 Ci/mmol, was prepared by catalytic tritium exchange as described elsewhere (Kalir et al., 1978). Its purity was >97%. Bretylium tosylate was the product of the Wellcome Foundation Ltd.

Methods

Adult male rats of the CD strain, obtained from Levinstein’s Farm, were maintained in an air-conditioned room

* 2-Bromo-N-ethyl-N,N-dimethylbenzenmethanaminium p-toluenesulfonate.
at 24 ± 2°C for 14 hour (5 a.m. to 7 p.m.) under fluorescent illumination, and in darkness for 10 hours. Food from Assia Maabarot Ltd. and water were supplied ad libitum. Rats aged 3–4 months and weighing 190–250 g were decapitated (between 10 a.m. and noon), and their brains or hearts were rapidly removed. The medulla-pons and the cortex were dissected out in a cold room after identification with the aid of a stereotaxic atlas (DeGroot, 1972).

**Binding Assay**

Atria and ventricles were cut up finely with scissors, mixed with 9 volumes of 0.32 M sucrose, homogenized at setting 7 on an Ultra-Turrax (Ika-Werk Instruments) with three 15-second bursts separated by 30-second pauses, and then filtered through three layers of cheesecloth and centrifuged twice in hypotonic sucrose solution (17,000 rpm, 15 minutes). Brain regions were homogenized, as previously described in detail (Kloog et al., 1979), to yield a 3% homogenate (wt/vol). Homogenates prepared from the medulla-pons (three rats for each experiment), the cortex (one rat for each experiment), and the atrium and ventricle (six rats each, for each experiment) were used for binding assays as follows: tissue preparation (50 μl) was incubated at 25°C in 2 ml of modified Krebs-Henseleit solution (25 mM Tris-HCl, 118 mM NaCl, 4.69 mM KCl, 1.9 mM CaCl₂, 0.54 mM MgCl₂, 1.0 mM NaH₂PO₄, 11.1 mM glucose), pH 7.4, containing varying amounts of the labeled ligand. After incubation for 40 minutes, ice-cold Krebs solution (3 ml) was added and the contents were passed rapidly by suction through a glass filter (Whatman GF/C, 25 mm in diameter). The filters were washed three times in 3 ml of ice-cold Krebs solution. All procedures were completed within less than 10 seconds. Binding assays were performed in triplicate, together with triplicate control samples containing 5 × 10⁻⁵ M unlabeled atropine. The filters were placed in vials containing 4 ml of scintillation liquid (Hydro-Luma, Lumac Systems Inc.) and were maintained at 25°C for 30 minutes; the radioactivity then was measured by liquid scintillation spectrometry (Packard Prias model PL) with a counting efficiency of 40–45%.

Protein was determined by the method of Lowry et al. (1951), using bovine serum albumin as a standard.

Specific binding was defined as the total binding minus the nonspecific binding, i.e., binding in the presence of 5 × 10⁻⁵ M unlabeled atropine. Direct binding studies and competition experiments were carried out as described in detail in previous reports (Kloog et al., 1979; Avissar et al., 1981).

**Data Analysis**

The competition curves (Fig. 1) were analyzed by a nonlinear least-square curve-fitting procedure, using a model for either one or two binding sites. Theoretical competition curves were fitted to the experimental data points using the nonlinear least-square regression computer program BMDPAR (November 1978 revision), developed at the Health Science Computing Faculty (University of California, Los Angeles, CA).

The goodness of fit was evaluated by comparison of the predictive error, given by the weighted sum of squared residuals, with the experimental error. The criterion for rejecting the one-site model in the cardiac preparations was a predictive error, which was significantly ($P < 0.01$, $F$-test) greater than the experimental error.

The statistical significance of the differences between the IC₅₀ values given for the cardiac preparations and those given for the brain preparations was tested using Student’s $t$-test. The same test was employed to evaluate whether the deviation from unity of the $n_H$ values in heart preparations is significant, and whether they differ significantly from the $n_H$ values found for the brain preparations.

**Effects on Isolated Organ**

Electrically stimulated left atria were mounted in a 14-ml organ bath filled with McEwen’s solution (McEwen, 1956) maintained at 37°C and gassed with 95% O₂ and 5% CO₂. Left atria were stimulated by a square wave stimulator at supramaximal voltage, 10-msec pulse duration, and a frequency of 5 Hz. Inotropic responses of the atria were recorded as described by Clark and Mitchelson (1976). Dose ratios and inhibition constants were calculated from the dose-response curves for acetylcholine in the presence of different concentrations of the drug tested, as described for the guinea pig ileum preparation previously (Amitai et al., 1980).

**Results**

A representative [³H]-4NMPB saturation binding curve to homogenates from rat atrium is shown in Figure 2. The Scatchard representation (see inset) yields a straight line. The dissociation equilibrium constant and the total binding capacity, calculated from three sets of experiments, were: $K_D = 0.8 ± 0.1$ nM and $R_T = 250 ± 20$ pmol/gr protein, respec-

**Figure 1.** Binding of 2 nM [³H]-4NMPB to homogenates of different brain and heart regions (25°C) in the presence of various concentrations of bretylium tosylate. Each point is the mean of four to six experiments whose standard error was less than 5%. Each experiment was performed in triplicate. The displacement by bretylium tosylate in the cortex and medulla reached 100% (data not shown).
Schreiber et al. /Bretylium Tosylate as a Muscarinic Ligand 655

The possible interaction of bretylium tosylate with the muscarinic receptor was tested by means of competition experiments with [3H]-4NMPB (2 nM). This concentration was selected because it produces approximately 80% occupancy of the sites (Fig. 2). Figure 1 shows the results of competition experiments with homogenates from brain tissue (cortex and medulla-pons), as well as from cardiac tissue (atrium and ventricle). Bretylium tosylate inhibited the binding of [3H]-4NMPB, thus indicating that it may act as an anticholinergic drug. The behavior of the drug was clearly different in the two types of tissues; this was evident from the variation in its IC50 values by a factor of 5-10 (Table 1), as well as from the differences in shape of the binding curves obtained with the brain and the heart. In cardiac tissue, the binding curves, unlike those for the brain, were flattened, and yielded Hill slopes of significantly less than unity (Table 1). The flattened curves and the different IC50 values can be explained by assuming the presence of multiple muscarinic binding sites in the heart characterized by different affinities for bretylium tosylate. Scatchard representation of the data (Fig. 3) does indeed appear to confirm the complex nature of displacement of [3H]-4NMPB by bretylium tosylate in heart homogenates. Whereas in both the cortex and the medulla-pons a straight line was obtained, the data for both the atrium and the ventricle yielded curvilinear lines deviating markedly from linearity.

To estimate affinities for bretylium tosylate from competitive experiments and statistically verify the possible existence of two muscarinic receptor binding sites for bretylium tosylate in the heart, we performed a nonlinear regression analysis. The data were analyzed using a model for either one or two binding sites, as described previously (Avissar et al., 1981). The binding of bretylium tosylate to the two brain preparations was found to be best fitted by a one-site model, in each case. On the other hand, in

![Figure 2. Saturation curve of the specific binding of [3H]-4NMPB to rat atria homogenate. Each point is the mean of triplicate samples whose standard error was less than 5%. Inset: Scatchard representation of the data.](image)

| Table 1 Diversity of Bretylium Binding to Heart and Brain Regions* |
|----------------------|------------------|------------------|
| Region               | IC50 (M)         | nH               |
| Cortex               | (1.8 ± 0.08) × 10^-3 | 0.96 ± 0.06     |
| Medulla-pons         | (1.3 ± 0.06) × 10^-3 | 0.98 ± 0.05     |
| Atrium               | (2.8 ± 0.11) × 10^-4 | 0.79 ± 0.06‡    |
| Ventricle            | (4.6 ± 0.20) × 10^-4 | 0.67 ± 0.04‡    |

* IC50 is the concentration of bretylium tosylate causing 50% reduction in binding of [3H]-4NMPB under the experimental conditions described in Methods and in Figure 2. The IC50 raw values were calculated according to IC50 raw = IC50/(1 + [C]/Kd), where Kd and [C] represent the dissociation constant and the concentration of [3H]-4NMPB, respectively. nH = slopes of Hill plots. Kd values for [3H]-4NMPB binding to cortex and medulla-pons preparations were 0.4 and 0.8 nM, respectively (Kloog et al., 1979). Values for the atrium and ventricle were 0.8 nM (Fig. 1).

The average values of the binding parameters were determined from the values obtained separately in four to six independent experiments, each performed in triplicate. The SE values were calculated from the parameter values obtained at the separate experiment.

† Significantly different from 1 (P < 0.05, Student's t-test).
‡ Significantly different from 1 (P < 0.05, Student's t-test).
Also significantly different from the nH values given for the brain (P < 0.05, Student's t-test).
FIGURE 3. Scatchard analysis of the displacement of [3H]-4NMPB by various concentrations of bretylium tosylate in different brain and heart regions.

The dissociation equilibrium constants of bretylium tosylate presented in Table 2 were obtained by nonlinear regression analysis of the competition curves shown in Figure 1. Since such curves are obtained with only one concentration of [3H]-4NMPB, it was necessary to verify the results for other concentrations as well. This is shown in the insets to Figure 4, where the slopes of the lines of the double reciprocal plots obtained for the cortex (Fig. 4A) and the ventricle (Fig. 4B) are depicted as functions of bretylium tosylate concentration. In the presence of a single binding site for bretylium tosylate, such a presentation would be expected to yield a straight line intersecting the ordinate at KD with a slope of KD/Ki. The Ki value, 1.20 × 10^-5 M, calculated from the slope of the line, is in accord with the value of 1.37 × 10^-5 M for Ki of bretylium tosylate in the cortex, shown in Table 2. The existence in the heart of two sites with different Ki values for bretylium tosylate is also supported by the curvature of the line obtained (Fig. 4B, inset).

In driven isolated left atria, bretylium tosylate (1.4–0.14 μM) inhibited the negative inotropic responses to acetylcholine. At the concentration range
investigated, bretylium tosylate by itself did not induce any inotropic effects on the responses of the atria to the electric stimulation. A parallel shift of the dose-response curves of acetylcholine in the presence of different concentrations of bretylium tosylate was obtained. Schild plot (Fig. 5) of the inhibitory effect curves yielded a straight line with a slope of \( \approx 45^\circ \), indicating competitive antagonism, with an inhibition constant \( K_i = 14 \pm 2 \mu M \).

**Discussion**

The present study demonstrates the ability of bretylium tosylate to interact directly with the muscarinic parasympathetic system. Bretylium tosylate was able to displace the muscarinic antagonist 4-NMPB from its receptor-ligand complexes in various brain regions, as well as in the heart. Specific sites with relatively higher affinity for bretylium tosylate were detected among the muscarinic receptors in the heart. Their dissociation equilibrium constants, which were in the range of 0.3–1.0 \( \mu M \), were 10 to 25 times lower than those of the corresponding low affinity binding sites (see Table 2); the latter were generally similar for all the brain and-heart preparations examined (Table 2). The finding that bretylium tosylate also interacts with brain muscarinic receptors is of interest, since it demonstrates that the interaction of the drug with muscarinic receptors is not a unique feature of these receptors in the heart. However, the effect on the brain is probably without physiological or therapeutic significance, since bretylium tosylate being a quaternary amine, does not cross the blood-brain barrier. The anticholinergic effect of the drug was verified physiologically with the guinea pig atrium preparation. The \( K_i \) value obtained, 14 \( \mu M \), was in good correlation with the low affinity binding constants found in the corresponding experiments.

![Figure 4](image4.png)

**Figure 4.** Double reciprocal plots of the specific binding of various concentrations of \( [\text{H}] \)-4NMPB to cortex homogenate (panel A) and ventricle preparation (panel B) in the presence of bretylium tosylate. \( R_m \) = maximal binding capacity of \( [\text{H}] \)-4NMPB; \( R_A \) = bound \( [\text{H}] \)-4NMPB. Binding was performed with different concentrations of the drug as indicated. Each point is the mean of four to six experiments whose se was less than 5%. Each experiment was performed in triplicate. Total receptor concentrations were 250 and 45 pmol/g protein for the atrium and ventricle, respectively, and 300 and 1000 pmol/g protein for the medulla-pons and cortex, respectively.

![Figure 5](image5.png)

**Figure 5.** Schild plot of the inhibitory effect of bretylium tosylate on the negative inotropic effect of acetylcholine in left atria preparation from guinea pig. Dose ratio is the ED\(_{50}\) ratio of the negative inotropic effect induced by acetylcholine in the presence and in the absence of bretylium tosylate. The experiment was repeated three times. The se for ED\(_{50}\) values was \( \pm 8\% \).
It is of interest to note that the first report on the antifibrillatory action of bretylium tosylate by Leveque (1965) was based on his special method of inducing arrhythmia, namely, through acetylcholine-induced atrial fibrillation. It is likely that parasympathetic influences are intimately involved in the induction and facilitation of a number of arrhythmias (Higgins et al., 1973). Parasympathetic activity may induce arrhythmia by depressing the activity of the sinus pacemaker cells, by blocking or slowing atrioventricular conduction, or by promoting nonuniform repolarization of adjacent regions of the myocardium (Higgins et al., 1973). Alessi et al. (1958) demonstrated, in the canine heart, that both direct and reflex vagal stimulation increases the nonuniformity of duration of the refractory period of various atrial sites, an action which favors the development of atrial fibrillation and other arrhythmias. However, it was assumed that bretylium tosylate had no effect on the muscarinic receptors of the heart (Bigger and Hoffman, 1980).

Namm et al. (1976) have determined the uptake of bretylium tosylate and norepinephrine by perfused hearts from normal and immunosympathectomized rats. At a concentration of 0.1–1.0 μM, the uptake of bretylium tosylate by the ventricles was reduced by only 10% following sympathectomy, compared to a reduction of 80% in norepinephrine uptake. Interestingly, this range of bretylium tosylate concentrations at which its uptake was independent of the sympathetic nervous system was close to the value of the high affinity binding constant for bretylium tosylate found in the present study (see Table 2). However, the total direct binding of radiolabeled bretylium tosylate to heart plasma membranes of immunosympathectomized rats was much higher (50 nmol/g) than the total concentration of muscarinic receptors in the ventricles found in this and other studies (Fields et al., 1978) (~50 pmol/g). These relatively high total amounts of directly bound drug suggest that bretylium tosylate is also bound to other nonspecific sites which are distinct from the adrenergic nerve terminals, as well as from the muscarinic receptor.

In view of the known antiaarrhythmic action of bretylium tosylate and the involvement of the parasympathetic muscarinic system in the induction and facilitation of cardiac arrhythmias, the present finding of a direct interaction between the drug and the muscarinic receptor in the heart may be relevant, at least in part, to the therapeutic effects of the drug.

This work was supported in part by the Recanati Fund for Medical Research (Israel).

Address for reprints: Dr. Mordechai Sokolovsky, Department of Biochemistry, George S. Wise Faculty of Life Sciences, Tel-Aviv University, 69978 Tel-Aviv, Israel.

Received November 18, 1983; accepted for publication August 13, 1984.
References


McEwen LM (1956) The effect on the isolated rabbit heart of vagal stimulation and its modification by cocaine, hexamethonium and ouabain. J Physiol (Lond) 131: 678-689


INDEX TERMS: Antiarrhythmic action • Muscarinic receptors • Drug • Multiple binding • Acetylcholine
Interaction of bretylium tosylate with rat cardiac muscarinic receptors. Possible pharmacological relevance to antiarrhythmic action.
G Schreiber, M Friedman and M Sokolovsky

doi: 10.1161/01.RES.55.5.653

_Circulation Research_ is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 1984 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7330. Online ISSN: 1524-4571

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circres.ahajournals.org/content/55/5/653

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in _Circulation Research_ can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to _Circulation Research_ is online at:
http://circres.ahajournals.org/subscriptions/