ALTHOUGH the influence of acute regional ischemia on left ventricular function has been well defined in the previously normal animal, the response of the ventricle affected by a chronic metabolic or structural abnormality has not been described. Acute myocardial infarction has been reportedly associated with a greater incidence of pump failure and higher mortality in diabetes mellitus. Although indices of contractility did not differ, stroke volume and work reductions were significantly greater in diabetics, despite the fact that coronary blood flow was reduced to a similar extent. Size of the ischemic areas appeared comparable as judged by distribution of dye injected distal to the occlusion. Since potassium loss and sodium gain in the inner and outer layers of ischemic tissue did not differ between the two groups, the intensity of ischemia seemed similar. Glycogenolysis was unimpaired in the diabetic ischemic muscle but triglyceride levels remained elevated. Morphologically the diabetic myocardium was characterized by a diffuse accumulation of periodic acid-Schiff-positive glycoprotein in the interstitium, which was thought to limit diastolic filling of the ischemic ventricle and to contribute to the substantial reduction of ventricular performance.

In a previous study from this laboratory, we observed altered myocardial function in chronic diabetes mellitus in dogs, associated with accumulation of periodic acid-Schiff (PAS)-positive glycoprotein in the myocardial interstitium without coronary obstructive lesions; this morphological abnormality also has been observed in man. To examine the response of the diabetic myocardium during acute regional ischemia as compared to normal controls the following study was undertaken.

Methods

Two groups of healthy male mongrel dogs 2–4 years old and weighing 21–28 kg were studied. The dogs had no clinical evidence of disease for 6–8 weeks before admission to the study groups. Hematocrit and serum albumin were initially normal and both groups received the same diet consisting of 8% fat, 22% protein, 58% carbohydrate, 9% ash, and 3% crude fiber. One group (n = 10) served as controls with normal glucose tolerance by intravenous testing. The other group (n = 9) was made diabetic with low doses of alloxan at monthly intervals. To produce mild normoglycemic diabetes, alloxan monohy-
diate in sterile saline was administered intravenously in a
dose of 20 mg/kg over a 1-minute period. Two additional
doses were given at monthly intervals to maintain a rela-
tively steady state of glucose intolerance. Larger doses of
alloxan were avoided to prevent ketoacidosis. The diabetic
dogs and the controls were observed for an average period
of 9 months after the initial alloxan dose.

Glucose tolerance was measured before and every 3
months after the initial dose of alloxan. Glucose was
infused over 1 minute (1.05 g/kg, iv) through catheter
tubing in the relatively relaxed, unanesthetized dog. Blood
samples were taken at 1.2.4.6.10.20.30.45.60. and
120 minutes, and plasma glucose was analyzed by the
glucose oxidase method. The glucose clearance constant
was calculated to estimate the disappearance rate from the
vascular compartment and was derived by a semilogarith-
mic plot of glucose concentrations beginning with the
1-minute sample for calculation of slope. In the diabetic
dogs venous blood samples were obtained in the fasting
state at the onset of the study and at 3-month intervals for
determination of plasma lipids. Blood was placed in
chilled tubes containing ethylenediaminetetraacetic acid
(EDTA); after separation in a refrigerated centrifuge, the
plasma was stored at —20°C until assay. Duplicate deter-
minations of free fatty acid, triglyceride, and phospha-
lipid were made.

HEMODYNAMIC STUDIES

Dogs were anesthetized with morphine sulfate (2 mg/
kg) and sodium pentobarbital (12 mg/kg, iv) and studies
were performed with the chest intact. Ventilation was
regulated by a Harvard pump via a cuffed endotracheal
tube to maintain pH and P0 2 within normal range. Cathe-
ters were placed in the pulmonary artery, left ventricle,
and ascending aorta and maintained patent with infusion
or intermittent flushing with small volumes of saline. The
50-cm Goodale-Lubin catheters were connected directly
to a Statham strain gauge transducer (P23Gb) and re-
corded on a multichannel oscilloscope recorder (Electron-
ics for Medicine). The first derivative of left ventricular
pressure was recorded at high sensitivity and the
mean left ventricular systolic pressure minus end-
diastolic pressure was recorded at mm Hg per second. Left ventricular end-
diastolic pressure was recorded at high sensitivity and the
average of 4–5 end-expiratory pressures were calculated.
To evaluate accuracy of the fluid-filled system used in this
study for measuring left ventricular pressure, four separate
dogs were studied. Left ventricular pressure recordings
were obtained with a no. 8 catheter-tip transducer (Micro-
Tip, Millar Instrument) and compared with pressure re-
corded via a 50-cm-long Goodale-Lubin catheter con-
nected directly to a P23Gb Statham strain gauge trans-
ducer positioned at the midthoracic level. Thirty-two si-
multaneous determinations of left ventricular end-diastolic
pressure during a wide range of hemodynamic interven-
tions showed close agreement, with a correlation coeffi-
cient of 0.98 (y = 1.019x — 0.409) in accord with a prior
report.

Cardiac output and left ventricular volumes were deter-
mined in duplicate by the thermal indicator-dilution
method. Cardiac output by this technique has been found to
correlate well with the dye-dilution technique. and
end-diastolic volume has been correlated with the an-giographic method. A Swan-Ganz thermodilution cathet-
ter tip was placed in the main pulmonary artery and 10 ml
of normal saline at room temperature was injected as a
bolus into the right atrium through the proximal lumen.
Change of temperature was detected by the thermodilutor
in the pulmonary artery and displayed on the recorder
through a Wheatstone bridge; the area of the curve was
computed manually for cardiac output calculation. Left
ventricular injection fraction was obtained in duplicate
from the left ventricular washout curves obtained by ther-
omilution method. Cold saline (5 ml) was injected as a
bolus at the inflow site of the ventricle and the step change
of temperature was determined from a thermodilutor-tip cath-
ter placed just above the aortic valve. Adequate mixing
appears to be present at this injection site, since prior
studies showed good correlation of ejection fractions de-
rived sequentially from left atrial and ventricular injection
sites at the level of the inflow tract or apex. The total
amount of saline infused for the measurement of cardiac
output and ejection fraction was approximately 150 ml in
each dog during the course of study. We have previously
observed that 180 ml/hour infused in dogs of equivalent
size produced no significant hemodynamic effects.

End-diastolic volume was calculated from the ratio of
stroke volume and ejection fraction and expressed per
kilogram of body weight. The ejection fraction values
were lower than the mean of 38% observed by the indica-
tor-dilution technique in conscious sedated dogs. How-
ever, this is within the range observed after the adminis-
tration of sodium pentobarbital anesthesia, apparently
related to the myocardial depressant effects of the anes-
thetic agent.

Stroke work index in gram-meters per kilogram was
calculated from the product of stroke volume per kilogram
and the mean left ventricular systolic pressure minus end-
diastolic pressure times 1.36. The ratio of left ventricular
diastolic pressure and volume was used as a simple index
of wall stiffness at end-diastole. To evaluate the contractile
state of the ventricle in vivo we have used an index that normalizes left ventricular dp/dmax, for (1) the
maximal isovolumic pressure (MIP); (2) circumferential
fiber length (2mr), assuming a spherical shape at the end of
the systolic isovolumic period and deriving the radius from
the end-diastolic volume. The formula is (dp/dmax/MIP)/
2mr. Using the same end-diastolic fiber length and pres-
sure, the end-diastolic tension in dynes x 10 6/beats was
calculated as end-diastolic pressure x 2mr x 4,188; the
latter is derived from the end-diastolic volume has been correlated with the angio-
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amount of saline infused for the measurement of cardiac
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each dog during the course of study. We have previously
observed that 180 ml/hour infused in dogs of equivalent
size produced no significant hemodynamic effects.

MODEL FOR ISCHEMIA

A double-lumen, 5F catheter with a distal lumen was posi-
tioned in the proximal 1.5 cm of the left anterior
descending coronary artery under fluoroscopic control.
The balloon was inflated gradually over a period of 60
seconds. Aortic pressure, peripheral coronary pressure,
and electrocardiogram (ECG) lead I were continuously monitored. Complete coronary occlusion was evidenced by a sustained reduction of mean coronary pressure to approximately 25 mm Hg and appearance of an injury potential on standard lead I in all the dogs studied.

To reduce mortality due to the high incidence of arrhythmias in the initial 15 minutes after ischemia, procainamide (10 mg/kg) was administered intravenously to dogs developing ventricular tachycardia (four in the nondiabetic group and three of the diabetics). No antiarrrhythmic agent was administered after the initial 15 minutes. Since the circulatory effects of procainamide so administered are considered to last a matter of minutes, it is improbable that the antiarrhythmic drug contributed to the hemodynamic differences observed in these two groups. Measurement of coronary blood flow to the ischemic area was obtained by injections of $^{85}$Kr distal to the occlusion site. This inert gas method appears to give valid flow measurements over a wide range of tissue perfusion. Approximately 100 $\mu$Ci of $^{85}$Kr was injected at 6- to 10-minute intervals in duplicate before ischemia and 12- to 20-minute intervals thereafter. Blood flow was calculated from the decay slopes obtained by precordial scintillation counting.

At 60 minutes of ischemia, the chest was opened and the heart was arrested with iced Ringer’s solution. To delineate the area profused by the left anterior descending coronary artery (LAD) distal to the occlusion, Evans blue dye was injected via the LAD catheter just prior to arrest. A transmural section was rapidly excised from the central ischemic area and the nonischemic posterior wall at least 1 cm from the former. Both were frozen in liquid nitrogen for glycogen assay. The remainder of the dyed area was excised, weighed, and related to the total left ventricle and septum. Sections were taken from ischemic and nonischemic areas of left ventricle for electrolyte and lipid analysis as well as histochemical examination. The ventricle was divided into inner and outer layers; the latter was carefully trimmed of epicardial adipose tissue. Samples were homogenized in phosphate buffer and the lipids were extracted in chloroform-methanol to determine free fatty acid, triglyceride, and phospholipid. Separate samples were homogenized and extracted for 48 hours in distilled water. Potassium and sodium were determined on an AutoAnalyzer system (Technicon) with flame attachment. Water content was obtained by drying samples in an oven at 100°C to constant weight. A group of normal intact anesthetized dogs without ischemia underwent similar tissue studies for comparison with the two experimental groups.

Histochemical examination included PAS staining after treating twice with diastase to exclude staining of glycogen. Statistical data were expressed as means ± standard errors; the paired or nonpaired Student's $t$-test was applied as appropriate.

**Results**

Both groups remained healthy over the approximate 9-month period prior to the induction of ischemia. Body weight was maintained; the initial hematocrit of $45 \pm 1.7$ in the control group and $44 \pm 2.0$ in the diabetics was not significantly changed during this period. Prior to the terminal study in the controls, the mean fasting blood sugar was $83 \pm 3$ mg/100 ml and the mean glucose clearance constant was $3.7 \pm 0.1$. In the alloxan-diabetic dogs the mean fasting plasma glucose was increased significantly from a control of $84 \pm 4$ mg/100 ml to $104 \pm 2$ ($P < 0.001$) prior to the final study, although the mean value of plasma glucose for the group was within normal limits. The glucose clearance constant was reduced from $3.3 \pm 1$ prior to alloxan to a level of $2.1 \pm 0.1$ ($P < 0.001$), before the terminal study.

**HEMODYNAMIC FINDINGS DURING ISCHEMIA**

The hemodynamic measurements were made in duplicate in the control state and during the course of ischemia in both groups. Baseline hemodynamic values prior to ischemia were not significantly different in the control and diabetic groups (Tables 1 and 2). Between 30 and 60 minutes of ischemia in the 10 dogs of the control group (Table 1), the stroke volume declined an average of 4%, ejection fraction by 21%, and stroke work by 16%. The end-diastolic pressure and volume increased by 80% ($P < 0.01$) and 25% ($P < 0.02$), respectively. Changes in heart rate and arterial pressure were not significant. In this nondiabetic group, three dogs developed ventricular fibrillation between 30 and 60 minutes of ischemia and the remaining seven survived the observation period. Since the seven surviving dogs exhibited ventricular responses at 30 minutes similar to those at 60 minutes, the three that succumbed early have been included.

The hemodynamic changes during ischemia in the diabetic group are indicated in Table 2. There were two patterns of hemodynamic response in this group. The two dogs that exhibited marked hypotension (group B, Table 2) progressed to shock and developed cardiac arrest between 30 and 60 minutes of ischemia. Group A was characterized by a modest but significant decline of arterial pressure and left ventricular failure and only one of the seven dogs developed late ventricular fibrillation.

In the seven dogs of group A, the end-diastolic pressure rose more than 3-fold while end-diastolic volume did not change significantly. Stroke volume and stroke work decreased by 35% ($P < 0.02$) and 55% ($P < 0.01$), respectively, a significantly greater decrease than in the nondiabetics (Table 2). In the normal control dogs, the cardiac index was maintained during ischemia and reduced insignificantly from a control of $119.9 \pm 6.9$ ml/kg to 112 ± 9.0. In contrast, during ischemia in the diabetic group, the cardiac index was reduced by almost one-third, (control, $114 \pm 7.9$ ml/kg to 78.5 ± 4.2; $P < 0.01$), a significantly different response from that of controls ($P < 0.02$).

Calculation of the normalized index of contractility to compare the relative performance of the contractile elements revealed no significant difference between normal dogs (1.10 ± 0.11) and diabetic dogs (1.26 ± 0.09) before ischemia; there was a similar small decline in both groups during ischemia to 0.86 ± 0.07 and 1.10 ± 0.19, respectively. Since the index of contractility did not differ in the
two groups, the reduced stroke volume appears related at least in part to the lack of end-diastolic volume increment. Calculated end-diastolic tension increased from a control of 0.29 ± 0.04 dyne x 10⁶ to 0.54 ± 0.08 (P < 0.01) in the normals, while in diabetes the rise of tension from a control of 0.22 ± 0.03 dyne x 10⁶ to 0.65 ± 0.06 during ischemia, was significantly greater than in the normals (P < 0.05). The ratio of end-diastolic pressure and volume was unaltered during ischemia in both groups but was significantly higher in the diabetic group (P < 0.005), suggesting enhanced wall stiffness in the diabetics.

Left ventricular function was further assessed by plotting stroke work index against end-diastolic pressure (Fig. 1A). In the control group during ischemia, the rising end-diastolic pressure was associated with a small decline of stroke work. In contrast, the diabetic group at 60 minutes of ischemia exhibited a substantial reduction of stroke work associated with a 3-fold increase of end-diastolic pressure. To determine whether the heart rate response may have affected this relationship, dogs with relatively small rate changes, within 16 beats/min of the control state, were compared in the two groups (Fig. 1B).

### Table 1: Left Ventricular Response to Acute Ischemia in Normal Dogs

<table>
<thead>
<tr>
<th>No.</th>
<th>Heart rate (beats/min)</th>
<th>Systolic/diastolic aortic pressure (mm Hg)</th>
<th>Left ventricular end-diastolic pressure (mm Hg)</th>
<th>Volume (ml/kg)</th>
<th>EDP/EVI</th>
<th>Stroke volume (ml/kg)</th>
<th>Stroke work (g-m/kg)</th>
<th>Ejection fraction</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>C</td>
<td>E</td>
<td>C</td>
<td>E</td>
<td>C</td>
<td>E</td>
<td>C</td>
<td>E</td>
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<tr>
<td>1</td>
<td>123 161 137/118 163/135</td>
<td>10 2.80 2.36 2.15 4.23 0.83 0.50 1.49 0.88</td>
<td>0.294 0.213</td>
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<tr>
<td>2</td>
<td>115 102 123/105 128/100 8 12 4.08 4.51 1.97 3.08 0.71 0.79 1.11 1.26</td>
<td>0.174 0.175</td>
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</tr>
<tr>
<td>3</td>
<td>112 143 174/135 170/138 14 28 5.34 5.50 2.63 5.30 1.23 0.94 2.68 1.82</td>
<td>0.230 0.171</td>
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<tr>
<td>4*</td>
<td>140 130 186/146 149/118 16 23 4.67 4.78 3.43 4.82 1.00 1.03 2.30 1.76</td>
<td>0.214 0.215</td>
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<tr>
<td>5</td>
<td>144 149 122/64 104/40 10 17 4.95 7.80 2.03 2.18 1.08 1.08 1.64 1.29</td>
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<td>7 13 3.29 5.12 2.13 2.55 0.68 1.00 1.52 1.42</td>
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<tr>
<td>7*</td>
<td>110 130 139/110 134/110 8 13 4.48 8.30 1.79 1.57 1.04 1.15 1.83 1.90</td>
<td>0.222 0.138</td>
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<tr>
<td>8</td>
<td>126 123 150/122 159/130</td>
<td>5 17 6.20 6.46 0.81 2.64 0.92 0.90 1.81 1.73</td>
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<td>9*</td>
<td>137 133 150/112 140/108 6 11 3.96 5.06 1.52 2.18 0.87 0.81 1.56 1.38</td>
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<tr>
<td>10</td>
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<td>0.230 0.167</td>
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</table>

Mean 129 134 150/116 143/110 8.6 15.3 4.37 5.47 1.98 3.03 0.92 0.89 1.76 1.48 0.216 0.171

C = control data; E = value obtained at 60 minutes of ischemia; EDP/EVI = ratio of end-diastolic pressure and end-diastolic volume index.

* E values obtained between 30 and 60 minutes of ischemia in nonsurvivors.

† P = paired t-test comparing C vs. E.

### Table 2: Left Ventricular Response to Acute Ischemia in Diabetic Dogs

<table>
<thead>
<tr>
<th>No.</th>
<th>Heart rate (beats/min)</th>
<th>Systolic/diastolic aortic pressure (mm Hg)</th>
<th>Left ventricular end-diastolic pressure (mm Hg)</th>
<th>Volume (ml/kg)</th>
<th>EDP/EVI</th>
<th>Stroke volume (ml/kg)</th>
<th>Stroke work (g-m/kg)</th>
<th>Ejection fraction</th>
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<td>E</td>
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<td>E</td>
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<tr>
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<td>126 132 149/112 119/95</td>
<td>8 24 2.37 4.87 3.37 4.92 0.69 0.59 1.34 0.78</td>
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<tr>
<td>3</td>
<td>168 138 165/136 117/109</td>
<td>6 32 4.31 3.90 1.39 8.20 0.80 0.57 1.70 1.01</td>
<td>0.185 0.146</td>
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<tr>
<td>4</td>
<td>144 128 140/105 133/84</td>
<td>4 20 3.75 2.17 1.06 9.21 0.80 0.61 1.60 0.71</td>
<td>0.213 0.280</td>
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<tr>
<td>5*</td>
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<tr>
<td>6</td>
<td>150 132 149/126 107/87</td>
<td>4 17 3.33 2.93 1.20 5.80 0.62 0.51 1.22 0.70</td>
<td>0.186 0.174</td>
<td></td>
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</tr>
<tr>
<td>7</td>
<td>81 128 188/143 160/135</td>
<td>10 31 4.52 4.50 2.21 6.88 1.30 0.68 3.22 0.92</td>
<td>0.287 0.151</td>
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</tr>
</tbody>
</table>

Mean 127 122 159/124 130/103 7.1 23.8 3.69 3.71 1.97 6.65 0.98 0.66 2.11 0.95 0.264 0.186

C = control data; E = value obtained at 60 minutes of ischemia.

* E values obtained between 30 and 60 minutes of ischemia in nonsurvivors.

† P = paired t-test comparing C vs. E.

‡ P = unpaired t-test comparing the responses in normals vs. diabetics.
Figure 1 Left ventricular (LV) function before and during ischemia. The stroke work index is plotted against end-diastolic pressure in both the nondiabetic and the diabetic group. The changes from the preischemic control are depicted by arrowhead. Panel A, which represents the data (mean ± SEM) for all the dogs in each group, illustrates that the impaired ventricular function in response to ischemia is significantly greater in the diabetic group (P < 0.01). Panel B represents the data for six dogs of the control group and four dogs of the diabetic group with heart rate change within 16 beats/min of the control state. The responses were qualitatively similar to those observed in the groups as a whole, indicating that these changes were independent of heart rate.

Coronary blood flow to the ischemic site, measured by the $^{85}$Kr clearance technique, was similarly reduced in both groups (Fig. 2). The ischemic area, as ascertained by injection of Evans blue dye, was 31.0 ± 2.4% of the total left ventricle in the control group and 32.7 ± 1.6% in the diabetic group.

To examine the tissue response to ischemia, the transmural concentrations of potassium and sodium in the outer and inner layers of ischemic myocardium were assayed (Fig. 3). In the control group, a significant reduction of potassium and an increase of sodium concentrations were present in the inner and outer layers of ischemic tissue. Diabetic dogs exhibited a similar K* reduction and sodium gain in the two layers of ischemic myocardium. Tissue water was 78.1 ± 0.73% in normals, and 82.9 ± 0.38% and 81.7 ± 0.44% in the inner and outer layers, respectively, of nondiabetics during ischemia; the respective values were 83.0 ± 0.63% and 82.5 ± 0.85% in the diabetic group. The nonischemic muscle exhibited small but non-significant changes of K* and Na* concentration in both groups with ischemia. However, tissue water was elevated in the nonischemic posterior wall of diabetics to 81.3 ± 0.42% and 81.4 ± 0.42% in the inner and outer layers, respectively, compared to 79.7 ± 0.32% (P < 0.01) and 79.0 ± 0.52 (P < 0.01) in nondiabetics. The increment of tissue water is more apparent when expressed in terms of grams of H2O per gram of dry weight. In the inner layers, where the largest changes were observed during ischemia, water increased to 4.85 ± 0.09 g/g dry weight in nondiabetics, and 4.88 ± 0.19 for diabetics, compared to 3.56 ± 0.21 in normals. The nonischemic posterior wall was increased to 4.35 ± 0.09 in diabetics and 3.92 ± 0.9 in nondiabetics (P < 0.01).

The expected decrease of glycogen levels in the ischemic tissue of the anterior wall in normals occurred to a similar extent in the diabetics (Table 3), supporting the view that the severity of ischemia was comparable in the two groups. Myocardial triglycerides were elevated in the inner and outer layers of both ischemic and nonischemic regions in the diabetics as compared to the corresponding area in the controls. Myocardial phospholipid and free fatty acid were not significantly different in the two groups. In the diabetics the plasma lipid classes in the fasting conscious state were within normal limits through the 9-month observa-
ventricular end-diastolic pressure in diabetics is consistent over a wide range of pressure and volume values in these in vivo experiments, the significantly higher ratio of end-diastolic pressure to end-diastolic volume in diabetics is consistent with an interpretation of reduced compliance compared to the nondiabetic group. A prior study of ischemia in dogs with a scar in the nonischemic myocardium indicated a response similar to that of these diabetic dogs. This was characterized by a 4-fold rise of filling pressure and a more profound reduction of stroke volume and work, hypotension analogous to that in the diabetics of group B occurred in some dogs. Thus, alteration of a portion of the nonischemic myocardium may substantially affect the response to ischemia even if hypertrophy does not occur. Since the interstitial accumulation of PAS-positive glycoprotein is a diffuse process in the diabetic myocardium (cf. Fig. 4 of Regan et al.), the relative contribution of ischemic and nonischemic ventricle cannot be defined with certainty. However, the behavior of the myocardium in dogs with scar strongly supports the view that chronic compositional changes in the nonischemic zone of the left ventricle may be the basis for the observed differences in response to ischemia compared to controls.

The nonischemic area is known to undergo mechanical and metabolic alterations, associated with those of the ischemic tissue. A modest reduction in glycogen levels was similar in diabetics and nondiabetics (Table 3). Slight reductions of potassium and gain of sodium in this area of muscle previously have been shown to be related to an enlarged extracellular space. However, our diabetic dogs exhibited a significant increase in water content of the nonischemic myocardium, with an interpretation of reduced compliance compared to the nondiabetic group. A prior study of ischemia in dogs with a scar in the nonischemic myocardium indicated a response similar to that of these diabetic dogs. This was characterized by a 4-fold rise of filling pressure and a more profound reduction of stroke volume and work, hypotension analogous to that in the diabetics of group B occurred in some dogs. Thus, alteration of a portion of the nonischemic myocardium may substantially affect the response to ischemia even if hypertrophy does not occur. Since the interstitial accumulation of PAS-positive glycoprotein is a diffuse process in the diabetic myocardium (cf. Fig. 4 of Regan et al.), the relative contribution of ischemic and nonischemic ventricle cannot be defined with certainty. However, the behavior of the myocardium in dogs with scar strongly supports the view that chronic compositional changes in the nonischemic zone of the left ventricle may be the basis for the observed differences in response to ischemia compared to controls.

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ject to ischemia. This was also manifested in the stroke work to end-diastolic pressure relationship, which was more abnormal than in nondiabetics and was independent of heart rate change (Fig. 1).

The modest reduction of aortic pressure in the diabetics was largely due to the reduction in stroke volume, since there was no significant difference in the calculated mean peripheral resistance when this group was compared to the nondiabetics. In two of nine diabetic dogs that exhibited marked hypotension, the drop in arterial pressure was in excess of the decline in stroke volume, so that the vasomotor response of the peripheral vasculature appeared to be inappropriate.

Coronary blood flow to the ischemic myocardium when the coronary artery is completely occluded is presumed to represent collateral flow. In this study the perfusion level in ischemic tissue appeared to be comparable in both normal and diabetic groups undergoing ischemia. Although morphological abnormalities have been observed in the media of intramural vessels in this animal model,6 this did not appear to effect luminal narrowing. The observation of collateral flow levels which were comparable to the nondiabetic group at this stage of diabetes implies a functionally normal microvasculature in response to the vasodilator stimulus of ischemia. The fact that the transmural distribution of potassium and sodium ions in the ischemic area was similarly altered in both groups supports the view that blood perfusion of the inner and outer wall was quantitatively similar.

In addition to the anatomical alterations of the ventricle in the diabetic dogs, the altered response to ischemia may be related to changes in energy production and utilization. Enhanced glycolgenolysis is a feature of the metabolic response in ischemic muscle.20 In view of the similarly reduced glycogen levels in diabetics and in nondiabetics after 60 minutes of ischemia, the glycolylic response does not appear to be impaired by diabetes of this degree and duration. On the other hand, endogenous lipolysis does not seem to contribute significantly to substrate avail-

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