

## It's the Metabolism That Makes Macrophages Detectable in the Magnetic Resonance Scanner

### Immune Cell Detection by Hyperpolarized <sup>13</sup>C Magnetic Resonance Imaging

Wolfgang R. Bauer, Constantin Lapa, Theresa Reiter

Modulation of the immune response becomes more and more attractive for therapeutic interventions of cardiovascular diseases, aiming to improve healing and prevent remodeling after myocardial infarction or in myocarditis.<sup>1,2</sup> However, there are also conflicting and sometimes disappointing results.<sup>3,4</sup> Although a huge body of knowledge concerning immunologic pathways and networks is available, translation to the human setup and clinical progress are rather slow. One reason for this discrepancy might be that most of the latter is—roughly speaking—based on in vitro studies, which, for example, led to the concept of distinct classes of proinflammatory and reparative macrophages. However, there is increasing evidence that immune cells as macrophages behave differently in tissue. They are plastic and their polarization into different states strongly depends on the environment, which is determined by the affected organ and type of pathology.<sup>5</sup>

#### Article, see p 1084

Hence, it is of paramount importance that immunologic processes and targets should be assessed in their natural surroundings—at best in vivo, which demands the development of appropriate imaging techniques. Such techniques should provide new insights into basic processes and might guide therapeutic interventions in humans.

In their article, Lewis et al<sup>6</sup> apply a cutting-edge technology of nuclear magnetic resonance imaging which makes macrophage metabolism visible in vivo, namely <sup>13</sup>C hyperpolarization. Though relevant <sup>13</sup>C-labeled metabolites as pyruvate and lactate are in principle also detectable by conventional magnetic resonance imaging (MRI), the low signal amplitude implies a long acquisition time, which impedes its application for in vivo imaging. This obstacle may be overcome by hyperpolarization of <sup>13</sup>C nuclear magnetization, which increases the signal by magnitudes and, therefore, allows its spatial resolved detection in

living objects. In a small and large animal model of myocardial infarction, the authors elegantly demonstrate that macrophages are responsible for an increased lactate signal in the necrotic area, which means that lactate may serve as a biomarker for this type of inflammatory cells. In addition, the authors also provide underlying mechanisms for this observation. After stimulation by lipopolysaccharides—assumed to mimic polarization of macrophages in the damaged myocardial environment—an increased lactate signal is observed which is because of metabolic reprogramming toward the glycolytic pathways. Finally, the authors suggest an immune-modulatory therapy by inhibition of macrophage glycolysis by 2-deoxyglucose application. As 2-deoxyglucose is administered early, only the inflammatory but not the reparative component of macrophage infiltration is attenuated as biomarkers of myocardial repair revealed. Remodeling was moderately but still significantly affected as treated hearts showed an increased ejection fraction.

Hyperpolarization techniques are from a technical point of view fascinating and aim, as shown in the article of Lewis et al,<sup>6</sup> at a relevant biomarker, namely the specific metabolic shift of the macrophages. The practical application, however, is demanding and requires an elaborated infrastructure. For this reason, the question arises whether there are alternatives. In general, 2 approaches are conceivable. Contrary to the presented investigation of metabolic markers, the detection of immune cells, such as macrophages, aims at the effectors themselves. This can be achieved by conventional MRI, for example, with iron oxide nanoparticles which shorten the relaxation time in their vicinity, or <sup>19</sup>F MRI. The latter technique is based on the application of <sup>19</sup>F-perfluorocarbon emulsions.<sup>7,8</sup> Circulating monocytes ingest these substances before migration toward the damaged tissue, where they differentiate into macrophages followed by appropriate polarization. Because fluorine is not present in most tissues, its detection by <sup>19</sup>F MRI is unambiguous. <sup>19</sup>F MRI can, from a technical point of view, be easily combined with conventional <sup>1</sup>H MRI, allowing for simultaneous acquisition of morphological, functional, and immunologic information. It has been demonstrated in reperfused myocardial infarcts that severe microvascular damage resulting in microvascular obstruction is associated with an attenuated macrophage invasion in these areas and an aggravated remodeling, when compared with animals with infarcts of the same size and without microvascular obstruction. This partly relativizes the statement of Lewis et al,<sup>6</sup> who claim that reduction of inflammatory activity is associated with a better functional outcome. Although some years ago, some <sup>19</sup>F-perfluorocarbon substances were already

The opinions expressed in this article are not necessarily those of the editors or of the American Heart Association.

From the Departments of Internal Medicine I (W.R.B., T.R.) and Nuclear Medicine (C.L.), University Hospital Würzburg, Germany; and Comprehensive Heart Failure Center, University Würzburg, Germany (W.R.B., C.L., T.R.).

Correspondence to Wolfgang R. Bauer, Department of Internal Medicine I, University Hospital Würzburg, Oberdürrbacher Straße 6–8, 97080 Würzburg, Germany. E-mail Bauer\_W@ukw.de

(*Circ Res.* 2018;122:1039-1040.)

DOI: 10.1161/CIRCRESAHA.118.312901.)

© 2018 American Heart Association, Inc.

*Circulation Research* is available at <http://circres.ahajournals.org>

DOI: 10.1161/CIRCRESAHA.118.312901

on the threshold for application in patients as blood substitutes, it is still open whether they will be available for patients in the near future.

Besides MRI, nuclear techniques exist, which visualize inflammatory activity. Because increased glucose metabolism is a hallmark of inflammation, positron emission tomography using the radiolabeled glucose analog [<sup>18</sup>F]-2-deoxy-2-fluoro-D-glucose (FDG) is the standard diagnostic test for nuclear imaging of inflammation. However, specificity of FDG is hampered by physiological myocardial glucose uptake. Recently, a number of promising, more specific alternatives have been introduced, including various radiolabeled nanoparticles (mainly evaluated in preclinical models) as well as small molecules targeting SSTR (somatostatin receptors) and CXCR4 (C-X-C chemokine receptor 4), which are both expressed on the cell surface of activated macrophages<sup>9</sup> and have been transferred to the clinical setting.

Radiolabeled somatostatin analogs like [<sup>68</sup>Ga]-DOTA-TATE or -TOC detect macrophage activity in inflammatory conditions, including acute myocardial infarction and sarcoidosis in pilot human studies,<sup>10,11</sup> whereas preclinical experiments in mice questioned the usefulness of this approach.<sup>12</sup> In atherosclerosis, specific SSTR II expression on proinflammatory M1 macrophages in plaques was demonstrated,<sup>13</sup> suggesting that SSTR-directed positron emission tomography might serve as a marker of atherosclerotic inflammation. However, conflicting data have been published to date in humans.<sup>14</sup> Another target on macrophages for noninvasive imaging is CXCR4, which is a regulator of leukocyte trafficking. First small human studies suggested that this approach is feasible in acute myocardial infarction and atherosclerosis.<sup>15,16</sup> Because CXCR4 antagonists have gained market authorization, chemokine receptor-directed positron emission tomography might serve as readout of target expression and help to monitor therapy.

In summary, the article of Lewis et al<sup>6</sup> is another important step which will advance our knowledge of inflammation and repair in the cardiovascular system. The data from the presented animal studies suggest a legitimate position of <sup>13</sup>C hyperpolarization among other novel imaging techniques. Further research to assess the value of this technique—especially in comparison to alternative imaging modalities and with regard to its prognostic value for myocardial repair and remodeling—is warranted.

### Sources of Funding

This study was supported by Deutsche Forschungsgemeinschaft (SFB688, TBB05 to W.R. Bauer) and the Bundesministerium für Bildung und Forschung (BMBF01 EO1504 to W.R. Bauer, C. Lapa, T. Reiter).

### Disclosures

None.

### References

1. Weirather J, Hofmann UD, Beyersdorf N, Ramos GC, Vogel B, Frey A, Ertl G, Kerkau T, Frantz S. Foxp3+ CD4+ T cells improve healing

after myocardial infarction by modulating monocyte/macrophage differentiation. *Circ Res*. 2014;115:55–67. doi: 10.1161/CIRCRESAHA.115.303895.

2. Lewis DR, Petersen LK, York AW, Ahuja S, Chae H, Joseph LB, Rahimi S, Uhrich KE, Haser PB, Moghe PV. Nanotherapeutics for inhibition of atherogenesis and modulation of inflammation in atherosclerotic plaques. *Cardiovasc Res*. 2016;109:283–293. doi: 10.1093/cvr/cvv237.
3. Saxena A, Russo I, Frangogiannis NG. Inflammation as a therapeutic target in myocardial infarction: learning from past failures to meet future challenges. *Transl Res*. 2016;167:152–166. doi: 10.1016/j.trsl.2015.07.002.
4. Honold L, Nahrendorf M. Resident and monocyte-derived macrophages in cardiovascular disease. *Circ Res*. 2018;122:113–127. doi: 10.1161/CIRCRESAHA.117.311071.
5. Nahrendorf M, Swirski FK. Abandoning M1/M2 for a network model of macrophage function. *Circ Res*. 2016;119:414–417. doi: 10.1161/CIRCRESAHA.116.309194.
6. Lewis AJM, Miller JJ, Lau AZ, Curtis MK, Rider OJ, Choudhury RP, Neubauer S, Cunningham CH, Carr CA, Tyler DJ. Noninvasive immunometabolic cardiac inflammation imaging using hyperpolarized magnetic resonance. *Circ Res*. 2018;122:1084–1093. doi: 10.1161/CIRCRESAHA.117.312535.
7. Ye YX, Basse-Lüsebrink TC, Arias-Loza PA, Kocoski V, Kampf T, Gan Q, Bauer E, Sparka S, Helluy X, Hu K, Hiller KH, Boivin-Jahns V, Jakob PM, Jahns R, Bauer WR. Monitoring of monocyte recruitment in reperfused myocardial infarction with intramyocardial hemorrhage and microvascular obstruction by combined fluorine 19 and proton cardiac magnetic resonance imaging. *Circulation*. 2013;128:1878–1888. doi: 10.1161/CIRCULATIONAHA.113.000731.
8. Flögel U, Ding Z, Hardung H, Jander S, Reichmann G, Jacoby C, Schubert R, Schrader J. In vivo monitoring of inflammation after cardiac and cerebral ischemia by fluorine magnetic resonance imaging. *Circulation*. 2008;118:140–148. doi: 10.1161/CIRCULATIONAHA.107.737890.
9. Armani C, Catalani E, Balbarini A, Bagnoli P, Cervera D. Expression, pharmacology, and functional role of somatostatin receptor subtypes 1 and 2 in human macrophages. *J Leukoc Biol*. 2007;81:845–855. doi: 10.1189/jlb.0606417.
10. Lapa C, Reiter T, Li X, Werner RA, Sannick S, Jahns R, Buck AK, Ertl G, Bauer WR. Imaging of myocardial inflammation with somatostatin receptor based PET/CT - A comparison to cardiac MRI. *Int J Cardiol*. 2015;194:44–49. doi: 10.1016/j.ijcard.2015.05.073.
11. Lapa C, Reiter T, Kircher M, Schirbel A, Werner RA, Pelzer T, Pizarro C, Skowasch D, Thomas L, Schlesinger-Irsch U, Thomas D, Bundschuh RA, Bauer WR, Gärtner FC. Somatostatin receptor based PET/CT in patients with the suspicion of cardiac sarcoidosis: an initial comparison to cardiac MRI. *Oncotarget*. 2016;7:77807–77814. doi: 10.18632/oncotarget.12799.
12. Thackeray JT, Bankstahl JP, Wang Y, Korf-Klingebiel M, Walte A, Wittneben A, Wollert KC, Bengel FM. Targeting post-infarct inflammation by PET imaging: comparison of (68)Ga-citrate and (68)Ga-DOTATATE with (18)F-FDG in a mouse model. *Eur J Nucl Med Mol Imaging*. 2015;42:317–327. doi: 10.1007/s00259-014-2884-6.
13. Li X, Bauer W, Kreissl MC, Weirather J, Bauer E, Israel I, Richter D, Riehl G, Buck A, Sannick S. Specific somatostatin receptor II expression in arterial plaque: (68)Ga-DOTATATE autoradiographic, immunohistochemical and flow cytometric studies in apoE-deficient mice. *Atherosclerosis*. 2013;230:33–39. doi: 10.1016/j.atherosclerosis.2013.06.018.
14. Tarkin JM, Joshi FR, Evans NR, et al. Detection of atherosclerotic inflammation by 68Ga-DOTATATE PET compared to [18F]FDG PET imaging. *J Am Coll Cardiol*. 2017;69:1774–1791. doi: 10.1016/j.jacc.2017.01.060.
15. Reiter T, Kircher M, Schirbel A, Werner RA, Kropf S, Ertl G, Buck AK, Wester HJ, Bauer WR, Lapa C. Imaging of C-X-C motif chemokine receptor CXCR4 expression after myocardial infarction with [68Ga]pentixafor-PET/CT in correlation with cardiac MRI. *JACC Cardiovascular Imaging*. 2018;pii: S1936-878X(18)30007-X. doi: 10.1016/j.jcmg.2018.01.001.
16. Hyafil F, Pelisek J, Laitinen I, et al. Imaging the cytokine receptor CXCR4 in atherosclerotic plaques with the radiotracer 68Ga-pentixafor for PET. *J Nucl Med*. 2017;58:499–506. doi: 10.2967/jnumed.116.179663.

KEY WORDS: Editorials ■ cardiovascular disease ■ magnetic resonance imaging ■ macrophage ■ myocardial infarction ■ wound healing

# Circulation Research

JOURNAL OF THE AMERICAN HEART ASSOCIATION



## **It's the Metabolism That Makes Macrophages Detectable in the Magnetic Resonance Scanner: Immune Cell Detection by Hyperpolarized <sup>13</sup>C Magnetic Resonance Imaging** Wolfgang R. Bauer, Constantin Lapa and Theresa Reiter

*Circ Res.* 2018;122:1039-1040

doi: 10.1161/CIRCRESAHA.118.312901

*Circulation Research* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231

Copyright © 2018 American Heart Association, Inc. All rights reserved.

Print ISSN: 0009-7330. Online ISSN: 1524-4571

The online version of this article, along with updated information and services, is located on the World Wide Web at:

<http://circres.ahajournals.org/content/122/8/1039>

**Permissions:** Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Circulation Research* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the [Permissions and Rights Question and Answer](#) document.

**Reprints:** Information about reprints can be found online at:  
<http://www.lww.com/reprints>

**Subscriptions:** Information about subscribing to *Circulation Research* is online at:  
<http://circres.ahajournals.org/subscriptions/>