Critical Role for Telomerase in the Mechanism of Flow-Mediated Dilation in the Human Microcirculation

Andreas M. Beyer, Julie K. Freed, Matthew J. Durand, Michael Riedel, Karima Ait-Aissa, Paula Green, Joseph C. Hockenberry, R. Garret Morgan, Anthony J. Donato, Refael Peleg, Mario Gasparri, Chris K. Rokkas, Janine H. Santos, Esther Priel, David D. Gutterman

Rationale: Telomerase is a nuclear regulator of telomere elongation with recent reports suggesting a role in regulation of mitochondrial reactive oxygen species. Flow-mediated dilation in patients with cardiovascular disease is dependent on the formation of reactive oxygen species.

Objective: We examined the hypothesis that telomerase activity modulates microvascular flow-mediated dilation, and loss of telomerase activity contributes to the change of mediator from nitric oxide to mitochondrial hydrogen peroxide in patients with coronary artery disease (CAD).

Methods and Results: Human coronary and adipose arterioles were isolated for videomicroscopy. Flow-mediated dilation was measured in vessels pretreated with the telomerase inhibitor BIBR-1532 or vehicle. Statistical differences between groups were determined using a 2-way analysis of variance repeated measure (n≥4; P<0.05).

L-NAME (Nω-nitro-L-arginine methyl ester; nitric oxide synthase inhibitor) abolished flow-mediated dilation in arterioles from subjects without CAD, whereas polyethylene glycol-catalase (PEG-catalase; hydrogen peroxide scavenger) had no effect. After exposure to BIBR-1532, arterioles from non-CAD subjects maintained the magnitude of dilation but changed the mediator from nitric oxide to mitochondrial hydrogen peroxide (% max diameter at 100 cm H2O: vehicle 74.6±4.1, L-NAME 37.0±2.0*, PEG-catalase 82.1±2.8; BIBR-1532 69.9±4.0, L-NAME 84.7±2.2, PEG-catalase 36.5±6.9*). Conversely, treatment of microvessels from CAD patients with the telomerase activator AGS 499 converted the PEG-catalase-inhibitable dilation to one mediated by nitric oxide (% max diameter at 100 cm H2O: adipose, AGS 499 78.5±3.9; L-NAME 10.9±17.5*; PEG-catalase 79.2±4.9). Endothelial-independent dilation was not altered with either treatment.

Conclusions: We have identified a novel role for telomerase in re-establishing a physiological mechanism of vasodilation in arterioles from subjects with CAD. These findings suggest a new target for reducing the oxidative milieu in the microvasculature of patients with CAD. (Circ Res. 2016;118:856-866. DOI: 10.1161/CIRCRESAHA.115.307918.)

Key Words: coronary artery disease ■ flow-mediated dilation ■ microvascular dysfunction ■ mitochondria ■ reactive oxygen species ■ telomerase activity ■ vascular biology

Telomerase is classically known as an enzyme that maintains telomere length in nuclear DNA. Telomerase activity is inversely related to endothelial senescence, especially in atherosclerotic prone vessels, and may attenuate premature coronary artery disease (CAD) and myocardial infarction. Increased telomerase activity can also protect against reactive oxygen species (ROS)–induced endothelial dysfunction.
such as hydrogen peroxide (H$_2$O$_2$) or prolonged hypoxia.\textsuperscript{6-8} Yet no physiological effects of this translocation have been described in connection with cardiovascular phenotypes. Once translocated to the mitochondria, TERT can suppress mitochondrial ROS (\textsubscript{mtROS}) production,\textsuperscript{5,7,9} TERT-overexpressing cells demonstrate lower mitochondrial superoxide (O$_2^-$) production,\textsuperscript{8} whereas in conditions of reduced telomerase activity, \textsubscript{mtROS} increases, contributing to cellular oxidative stress.\textsuperscript{6,10} Whether this system is active in the vascular endothelium and whether it contributes to redox regulation of vasodilation is not known. We have shown previously that in atrial and adipose vessels from human subjects without CAD, nitric oxide (NO) is the primary mediator of endothelium-dependent dilation in response to increased shear stress (flow-mediated dilation [FMD]).\textsuperscript{11,12} In CAD, FMD persists but is mediated almost exclusively by H$_2$O$_2$ released from the mitochondria, independent of nitric oxide synthase (NOS).\textsuperscript{13,14} Telomerase can modulate the cellular redox state by upregulation of mitochondrial antioxidant enzymes\textsuperscript{15} and by directly reducing \textsubscript{mtROS} production,\textsuperscript{16} making it ideally suited for contributing to the switch from NO to mitochondrial-derived H$_2$O$_2$ as the mediator of FMD in subjects with CAD.

Eitan et al have recently described a specific activator of telomerase that reduces \textsubscript{mtROS}. The small molecule AGS 499 was confirmed to act in a telomerase-dependent manner because in telomerase-depleted cells (by shRNA or knockout), AGS 499 did not elicit protection against increased oxidative stress.\textsuperscript{17,18} These studies also showed that AGS 499 does not act nonspecifically as an antioxidant but specifically increases extranuclear levels of TERT, including mitochondrial TERT.\textsuperscript{4} AGS 499 is beneficial in a variety of disease conditions characterized by elevations in \textsubscript{mtROS}.\textsuperscript{17,18} We hypothesized that a reduction in telomerase activity, insufficient to reduce telomere length, evokes \textsubscript{mtROS} production and reduces NO synthesis, recapitulating the phenotype observed in CAD where H$_2$O$_2$ mediates FMD. Conversely, in vessels from subjects with CAD and elevated \textsubscript{mtROS}, we tested whether short-term telomerase activation could restore NO-mediated dilation.

Methods

Tissue Acquisition and General Protocol

All protocols were approved by the Institutional Review Board of the Medical College of Wisconsin and Froedtert Hospital. Sections of human atrial and adipose (visceral and subcutaneous) tissue were obtained in deidentified fashion as otherwise discarded tissue at the time of surgery and placed in cold 4°C HEPES (NaCl 275 mmol/L, KCl 7.99 mmol/L, MgSO$_4$ 4.9 mmol/L, CaCl$_2$·2H$_2$O 3.2 mmol/L, KH$_2$PO$_4$ 2.35 mmol/L, EDTA 0.07 mmol/L, glucose 12 mmol/L, HEPES acid 20 mmol/L; adipose) or cardioplegic (atrial) buffer solution. Tissue from subjects without known cardiovascular risk factors or clinical diagnosis of CAD was used for non-CAD groups.

Cannulated Artery Preparation

Coronary and adipose arterioles (50–200 μm inner diameter) were cleaned of fat and connective tissue and prepared for continuous measurements of diameter.\textsuperscript{13,14} Briefly, in an organ chamber, both ends of the vessel were cannulated with glass micropipettes filled with physiological saline solution and pressurized (60 mm Hg) for videomicroscopy of diameter change as we have described previously.\textsuperscript{19,20}

Vascular Response to Flow and Pharmacological Interventions

FMD was measured as described by Kao et al\textsuperscript{21} using pipettes of identical impedance. Changing the height of each reservoir in equal and opposite directions generates flow without changes in vessel central pressure.\textsuperscript{22} Data are reported as diameter at a given pressure gradient. Pressure gradients of 5 to 100 cm H$_2$O were generated, assessing steady-state diameter and flow after each change, representing estimated shear rates of 5 to 25 dynes/cm$^2$. Two flow-response curves were generated for each vessel comparing no treatment (vehicle) to effects of pharmacological inhibitors (N$^\omega$-nitro-L-arginine methyl ester [L-NAME; 100 μmol/L]; polyethylene glycol-catalase (PEG-catalase; 500 U/mL); rotenone [1 μmol/L]; 2-(4-carboxyphenyl)-4,5-dihydro-4,4,5,5-tetramethyl-1H-imidazol-1-yl-oxy-3-oxide (c-PTIO) [1 μmol/L]; actinomycin D (1.59 μmol/L), and MitoTempol (10 μmol/L). In some cases, vessels were incubated for 15 to 20 hours in endothelial cell growth medium containing 5% serum (Lonza) containing either vehicle (DMSO 2% by volume), BIBR-1532 (10 μmol/L), or AGS 499 (20 μmol/L). After incubation, media was replaced with Krebs buffer, intraluminal pressure was raised from 30 mm Hg to 60 mm Hg, and vessels were constricted with ET-1 (endothelin-1) for assessment of FMD.

All pharmacological agents were added to the external bathing solution (volume added was <1% of the circulating external bath solution). Concentrations are stated as final concentrations in organ bath.

Vessels were constricted with ET-1 (0.1–1 μmol/L) to achieve a 20% to 50% stable reduction in passive diameter. Dose response curves to the endothelial-dependent vasodilator acetylcholine (ACh; 1 mmol/L to 100 μmol/L) and the endothelium-independent vasodilator papaverine (0.1 mmol/L to 100 μmol/L) were performed to evaluate specific endothelial and smooth muscle–diameter dependent dilation. At the end of each experiment, papaverine (100 μmol/L) was used to determine the maximal (passive) diameter at 60 mm Hg.

Quantitative Real-Time Polymerase Chain Reaction

Total mRNA was harvested from human umbilical vein endothelial cells (HUVECs) using an Ambion PureLink RNA Kit. Approximately 1500 ng of RNA was used to synthesize cDNA using the Applied Biosystems High Capacity cDNA Reverse Transcription Kit. Gene expression was quantified by real-time-quantitative polymerase chain reaction using primers (Hs_TERT_1.5_SG QuantiTect Primer Assay QT00073409) and SYBR green from Qiagen in a BioRad CFX96 Touch Real-Time PCR Detection System. Expression levels were normalized to 18S rRNA (Hs_18S5_1.5_SG QT00199367).

Cell Culture Experiments

HUVECs were freshly isolated from umbilical cords collected from local hospitals, processed by the hybridoma core facility of the Blood Research Institute of Wisconsin, and supplied at passage 3 to 5. Cells were exposed to AGS 499 for identical time and conditions as isolated vessels. For stressing cells in culture, heat shock treatment was used wherein culture dishes were sealed with Parafilm and immersed in a water bath at 42°C for 30 minutes. Subsequently, the cells were
put into a 37°C incubator for 16 hours of recovery. Control cells were sealed for 30 minutes but remained in the incubator.

Normal human fibroblasts were cultured under standard conditions as described previously. In brief, cells were transiently transfected with either whole cell TERT or R3E/R6E TERT ( ∆ TERT). Oxidative stress was generated by 30 minutes of 200 μmol/L H₂O₂ treatment. Purity of isolated mitochondria preparation was tested using the standard subcellular markers tubulin (cytoplasmic), mitochondrial heat shock proteins 70, and Ku80 (nuclear isoform [higher molecular weight] and mitochondrial isoform). Mitochondria were labeled with Mitotracker Red using manufacturer’s specifications.

Measurement of Mitochondrial Reactive Oxygen Species
In vessels, Mito Peroxy Yellow 1 (MitoPY1) was used to evaluate microvessel generation of H₂O₂. As previously described, after cannulation in a warmed chamber (37°C) containing HEPES buffer at pH 7.4, arterioles were perfused intraluminally with MitoPY1 (5 μmol/L, 1 hour) at low levels of flow, below the threshold for dilation, until the luminal surface was bathed in MitoPY1 containing buffer. Next the transvascular pressure gradient was changed from zero to 100 cm H₂O, and fluorescence measured after 5 minutes. Experiments were performed in the presence or absence of PEG-catalase (500 U/mL). Fluorescence was evaluated with a Nikon Eclipse TE200 microscope using a krypton/argon laser at excitation wavelength of 488 nm and measured emission between 530 and 590 nm. All comparisons were made using vessels studied at the same session with constant microscope image display settings. In cultured cells, mitoSox was used to measure superoxide generation. Cells were incubated for 30 minutes with 10 μmol/L mitoSox. Relative average fluorescence intensity was normalized to background fluorescence and presented as percent change from baseline.

Telomere Length
Genomic DNA from left ventricle tissue or isolated atrial arterioles was prepared by the Medical College of Wisconsin (MCW) pathology tissue bank using Qia spin column according to manufacturer’s specifications. Polymerase chain reaction reaction was performed under following conditions: 0.75 SYBR Green I (Invitrogen), 10 mmol/L Tris–HCl pH8.3, 50 mmol/L KCl, 3 mmol/L MgCl₂, 0.2 mmol/L each dNTP (deoxynucleotides), 1 mmol/L DTT (dithiothreitol), and 1 mol/L betaine (US Biochemicals). For 25 μL reaction, 0.625U AmpliTaq Gold DNA polymerase (Applied Biosystems, Inc.) was used. Multiplex QPCR primer pair final concentrations were 900 nmol/L each. Telomere primers were telg, TGGTTAGGTATCCATCCCTCATCCCTATCCCTATCCCTAAACA human albumin primers albu: CCGGCCGCGGGCCGCGGC- GGCTGGGCGG-aaatgctgcacagaatccttg; albd: GCCCGCCCGC- CGCGCCCGTGTCGCCGG-gaaatgctgcacagaatccttg. Polymerase chain reaction was performed as follows: step 1 (1X): 95°C for 15 minutes; Step 2 (36X): 98°C for 2 s, 48°C for 1 min, 74°C for 15 s (data acquisition), 84°C for 30 s, 85°C for 15 s (data acquisition) as previously described by Morgan et al.

Materials
The telomerase modulators BIBR-1532 (Tocris Bioscience) and AGS 499 (Ester Priel, PhD, Israel) and the mitochondrial ROS-detecting dyes MitoPY1 (Cayman chemicals) and mitoSox (life technologies) were prepared in DMSO. ET-1 (Peninsula Laboratories) was prepared in saline with 1% bovine serum albumin. All other chemicals were obtained from Sigma-Aldrich and prepared in distilled water or physiological salt solution. All concentrations represent the final concentrations in the organ bath.

Statistical Methods
Data are presented as means±SEM. For all flow response curves, differences between groups at each concentration were determined using a 2-way, repeated-measures analysis of variance. A post hoc Tukey’s test was used for individual comparisons. A probability value of P<0.05 was considered to be statistically significant. For fluorescence studies, telomere length, Western blot, or expression analysis, either a paired t test or 1-way analysis of variance with post hoc Tukey’s test was used to identify individual differences within groups. Values outside of 2 standard deviations of the mean were considered statistical outliers and excluded from analysis.

Results
Tissue was collected from a total of 58 subjects. Results are derived from arterioles of 36 patients without CAD and 18 patients with CAD. Detailed patient demographic information is shown in Table.

Effect on FMD of Inhibiting Telomerase
After incubating atrial or adipose arterioles for 15 to 20 hours in the specific telomerase inhibitor BIBR-1532, the magnitude of FMD was not changed (Figure 1A). However, the mediator of FMD shifted from NO to H₂O₂ (PEG-catalase inhibitable and L-NAME-insensitive) in both vessel types after treatment with BIRB1532 for 15 to 20 hours (Figure 1B and 1C).

| Table. Patient Characteristics for Microvessels Used in Study |
|-------------------|---------------------------------|--------|---------|
|                   | Adipose (n=38)                  | CAD (n=8) |
| Characteristics   | Non-CAD | CAD | Non-CAD | CAD |
| Sex, M/F          | 14/24  | 7/1 | 4/6     | 5/5  |
| Age, y (average±SEM) | 49.1±2.6  | 65.6±2.6* | 56.2±4.1 | 62.6±2.8* |
| BMI (average±SEM) | 28.9±1.3 | 31.8±1.8 | 27.6±1.4 | 29.4±1.5 |
| BMI >30           | 8 of 38  | 3 of 8 | 2 of 10 | 3 of 5 |
| Underlying diseases/risk factors | | | |
| Coronary artery disease | 0 | 8 | 0 | 10 |
| Hypertension | 1 | 7 | 5 | 7 |
| Hypercholesterolemia | 1 | 4 | 1 | 4 |
| Diabetes mellitus | 0 | 3 | 1 | 2 |
| Congestive heart failure | 0 | 1 | 0 | 0 |
| None of the above | 37  | 0 | 5 | 0 |

BMI indicates body mass index; and CAD, coronary artery disease.

*P<0.05 t test. Compared with non-CAD.
†Total of 7 subjects with no information on height and weight (3 non-CAD and 4 CAD).
Endothelial-dependent dilation to ACh was also reduced after telomerase inhibition (Figure 1D). Maximal endothelium-independent dilation (assessed with papaverine) was not altered by BIBR-532 (Figure 1E).

Subjects With CAD Have Reduced Cardiac and Vascular TERT Levels but Normal Telomere Length

To establish whether the difference in vasodilator mechanism between CAD and non-CAD subjects correlates with telomere length, we measured telomere length in heart and microvessels from individuals with and without CAD. No difference in telomere length was observed between vessels from patients with or those without CAD. TERT protein expression was significantly higher in non-CAD subjects versus those with CAD (Figure 2), although CAD subjects tended to be older (66±4 versus 48±7 years). Immunohistochemistry of coronary vessels from subjects with and without CAD revealed expression of TERT in both smooth muscle and endothelial cells (Online Figure I).

Role of Mitochondria as the Source of H₂O₂ and the Site of TERT Action in FMD

Using MitoPY1, a mitochondrial-specific fluorescent indicator for H₂O₂, we observed that for 15 to 20 hours, BIBR-1532 treatment in vessels from subjects without CAD caused stimulated mitochondrial release of H₂O₂ in response to flow (Figure 3A and 3B).

Addition of the complex I inhibitor rotenone abolished FMD in vessels treated with BIBR-1532 (atrial and adipose) but had no effect on vehicle-treated vessels (Figure 3). Scavenging mitochondrial O₂⁻ with the mitochondrial-specific antioxidant mitoTEMPOL (10 μmol/L) reduced FMD in vessels treated with BIBR-1532 (Figure 3C). Taken together, these data suggest that the source of ROS modulated by the telomerase inhibitor BIBR-1532 is from the mitochondrial electron transport chain.

Because of the small tissue volume of human arterioles, we were not able to quantify protein content. Therefore, we sought direct evidence for TERT translocation to the mitochondria using cultured human cells transfected with TERT-eGFP (endothelial green fluorescent protein) expression vectors as previously published.⁹

In cells expressing wild-type (WT) TERT, protein fused to eGFP nuclear localization of TERT was reduced following stress (application of H₂O₂). A corresponding increase in mitochondrial localization of TERT-GFP (green fluorescent protein) was seen by colocalization with the mitochondrial probe mitotracker red (Figure 4A).

To test whether the mitochondrial localization of TERT limits mtROS production, we used a mutant version of TERT (nt TERT) that we have previously shown to be functional (elongation of telomeres) but excluded from the mitochondria.⁹ Western blots confirmed the mitochondrial localization of the WT protein and the lack of mitochondrial localization of nt TERT (Figure 4B). Whole cell TERT levels were comparable among WT and nt TERT groups but only WT TERT is detected in highly purified mitochondria (Figure 4B). Generation of mtROS (mitoSOX) depended on the subcellular localization of TERT, with mt TERT cells showing consistently higher mitoSOX signal (Figure 4C). To evaluate the effects of TERT localization on the response to stress, WT TERT- or nt TERT-expressing cells were plated for

Figure 1. Telomerase inhibition replicates the coronary artery disease (CAD) vascular phenotype in human adipose and atrial vessels from non-CAD subjects. Flow-mediated dilation (FMD) was evaluated in isolated microvessels treated with vehicle (A; adipose, BIBR-1532 (B; adipose), and BIBR-1532 (C; atrial). In BIBR-treated vessels, the mechanism of FMD changed from nitric oxide (NO) to hydrogen peroxide (H₂O₂). D, Acetylcholine (ACh)-induced dilation was virtually eliminated after telomerase inhibition. E, No change in endothelium-independent dilation to papaverine was observed. N=4 to 12 adipose, N=5 to 9 atrial. *P<0.05 2-way analysis of variance (ANOVA) RM Tukey post hoc analysis. L-NAME indicates N'-nitro-L-arginine methyl ester; and Peg-Catalase, polyethylene glycol-catalase.

Figure 2. Coronary artery disease (CAD) does not cause telomere shortening in microvessels or left ventricular tissue but decreases expression of the catalytic subunit catalytic subunit of human telomerase complex (TERT). A, Total average telomere length was evaluated in genomic DNA from microvessels (adipose) and left ventricular tissue from subjects with and without CAD. B, Expression of the TERT was evaluated in left ventricular tissue from subjects with and without CAD by Western blot. Values are normalized to β-actin loading control and expressed as fold change compared with non-CAD control. N=4 to 7. *P<0.05 t test.
16 hours then treated with 200 μmol/L H₂O₂ for 60 minutes. Cells expressing nucTERT (aka absence of mtTERT) show a significant decrease in the steady state amount of ATP produced compared with WT (Figure 4D), indicating that absence of mtTERT has direct effects on mitochondrial function by impairing ATP production capacity following stress.

Effects of Telomerase Inhibition Are Independent of Transcription

It is possible that the switch in mechanism of FMD based on TERT modulation is transcriptionally induced because TERT regulates gene expression (reviewed by Zhou et al.24). To test this possibility, the transcription inhibitor actinomycin D (2 μg/mL)25 was used. Actinomycin D had no effect on the % increase in fluorescence from baseline.

Figure 3. Mitochondrial hydrogen peroxide (H₂O₂)-mediated flow-mediated dilation (FMD) after inhibition of telomerase. H₂O₂ levels were analyzed using the H₂O₂-specific fluorescent probe PYI (peroxy yellow 1) targeted to mitochondria (MitoPYI) in vessels from non-coronary artery disease (CAD) subjects. A, Representative image; numbers represent fluorescent intensity above background. B, Summary of fluorescence intensity at 5 minutes after initiation of flow. Specificity of the probe for H₂O₂ was confirmed using polyethylene glycol-catalase (Peg-Catalase). C, An inhibitor of electron transport chain complex I (rotenone) or a mitochondrial-targeted reactive oxygen species (ROS) scavenger (MitoTempol) inhibited FMD after telomerase inhibition. N=4. *P<0.05 2-way analysis of variance (ANOVA) RM (dose response curve) or t test (fluorescence data) with Tukey post hoc.

Figure 4. Mitochondrial translocation of catalytic subunit of human telomerase complex (TERT) after exposure to acute oxidative stress. A, Treatment of normal human fibroblasts (HF) with hydrogen peroxide (H₂O₂) resulted in translocation of TERT-GFP (green fluorescent protein) from the nucleus to the mitochondria (colocalized with Mitotracker Red). Cells were imaged using a 63× oil immersion objective. B, To confirm purity of isolated mitochondria of cells either transfected with whole cell (WC) TERT or R3E/R6E TERT (nucTERT) Western blots for tubulin (cytoplasmic marker), mitochondrial heat shock proteins 70 (mtHSP70), and Ku80 (nuclear isoform [higher molecular weight] and mitochondrial isoform) were performed. C, Expression of nucTERT increased mitochondrial superoxide production as measured with MitoSox. D, Expression of nucTERT decreased cellular adenosine triphosphate (ATP) production after external stress. n=3 to 5. *P<0.05 t Test.
mediator of FMD in vessels from patients without CAD treated 15 to 20 hours with vehicle or with the telomerase inhibitor BIBR-1532 (Figure 5). Efficacy of the dose of actinomycin D was demonstrated in endothelial cells treated for 2 hours with actinomycin D or vehicle and exposed for 1 hour to heat shock (42°C). Fifteen to 20 hours later, cells in the vehicle group showed an increase in HSP 27 and 70 compared with unstressed cells. Pretreatment with actinomycin D eliminated this effect (Figure 5C and 5D). These data indicate that nuclear transcriptional properties of telomerase are not involved in the functional and redox changes observed in this study.

**Activation of Telomerase Restores NOS-Mediated FMD in Vessels From Subjects With CAD**

Adipose and atrial vessels from subjects with CAD were exposed to AGS 499 (15–20 hours; 22 nmol/L), an activator of telomerase. AGS 499 restored the mediator of FMD from H$_2$O$_2$ to a NOS-derived product. Maximal dilation to flow (100 cm H$_2$O) in vessels from subjects with CAD treated with AGS 499 (15–20 hours; 22 nmol/L), an activator of telomerase. AGS 499 restored the mediator of FMD in vessels from patients without CAD treated with vehicle. AGS 499 was of similar magnitude to vehicle-treated vessels (% max diameter at 100 cm H$_2$O in atrial vessels (n=3–4): vehicle 64.5±8.7, L-NAME 12.6±15.3*, PEG-catalase 65.2±7.8, c-PTIO 22.9±17.3; adipose vessels (n=5–8): AGS 499+vehicle 79.5±3.9, L-NAME 10.9±17.2*, PEG-catalase 79.2±4.9; *P<0.05 2-way analysis of variance RM; Figure 6).

Inhibition of telomerase activity in vessels from subjects with CAD had no effect (Figure 5C). The dose of AGS 499 used was transcriptionally active because HUVECs treated with the same dose demonstrated an 4 fold increase of total TERT mRNA (Figure 6D).

To establish the effect of telomerase on endothelial nitric oxide synthase (eNOS), we used quantitative polymerase chain reaction and Western blotting. In HUVECs, AGS 499 increased eNOS message, whereas BIBR-1532 had no effect (Online Figure IIa). Levels of p-eNOS (ser 1117) increase after treatment with AGS 499 (Online Figure IIb). Similarly, Daf-2A (HPLC [high-performance liquid chromatography]; Methods in Online Data Supplement) significantly increased with AGS 499 treatment in HUVECs, whereas BIBR-1532 had no effect (Online Figure IIc).

**Discussion**

There are 3 primary novel findings of this study. First, we establish that inhibition of telomerase in vessels from subjects without CAD recapitulates the vascular phenotype seen in patients with CAD by converting the mediator of FMD from NO to H$_2$O$_2$. Activation of telomerase can restore an NOS-mediated FMD in vessels from patients with CAD. TERT protein levels are reduced in patients with CAD compared with those without CAD, and mtROS production is elevated when telomerase activity is reduced (mitoPY1). Second, the intracellular site for these actions of telomerase is the mitochondria and not the nucleus as demonstrated with novel mutant telomerase constructs that target either the whole cell (WT) or the nucleus alone (nucTERT). Third, the effects of telomerase inhibition are not dependent on changes in gene expression. Collectively, these data for the first time provide evidence for an extranuclear role for telomerase in modulating vascular function. Figure 7 illustrates our findings.

**Figure 5. Change in vascular phenotype is independent of changes in gene expression.** Flow-mediated dilation (FMD) was evaluated in isolated adipose microvessels from subjects without coronary artery disease (CAD) treated with actinomycin D, a transcriptional inhibitor. A, Act D+vehicle. B, Act D+BIBR-1532. Mechanism of FMD changed from NO (Act D)→H$_2$O$_2$ (Act D+BIBR-1532). C and D, Act D was sufficient to prevent transcriptional activation of heat shock proteins 27 and 70 after acute exposure to 42°C. Sample blot shows 2/5 total treatments. *P<0.05 2-way analysis of variance (ANOVA) RM Tukey post hoc analysis (functional studies). N=4 to 5. *P<0.05 vs untreated control t Test Tukey post hoc analysis (protein levels). CO indicates control; HS, heat shock; L-NAME, N$\omega$-nitro-L-arginine methyl ester; and Peg-Catalase, polyethylene glycol-catalase.
An extranuclear role for telomerase suggests noncanonical sites of actions. Based on our subcellular localization experiments and prior studies in other cell types, we postulate an effect on mitochondria. The short timeframe of the observed changes (hours) supports such a telomere-independent mechanism. Our study provides a physiological role for the mitochondrial ROS-suppressing effects of TERT. Because TERT suppresses ROS production in vascular endothelial cells, reduced TERT, as seen in CAD, would support ROS production necessary for FMD in human arterioles. TERT as a regulator of ROS is important in other diseases as well. Transcriptional activation of TERT via AGS 499 delayed the onset of amyotrophic lateral sclerosis in mice by increasing cell survival after oxidative stress. The same mechanism of action was shown in human bone marrow mesenchymal stem cells which were protected from oxidative stress by TERT.

The role of telomerase in chronic disease is multifactorial. Traditionally, TERT protects tissues, including the vasculature, from chromosomal rearrangements by maintaining telomere length over time. For example, shortened telomeres and decreased telomerase activity are associated with atherosclerosis and activation of senescence pathways. Of recent, it has been shown that telomere uncapping without change in telomere length which can also invoke cellular aging and inflammation or novel data show a role of TERT as a regulator of gene expression by modulating chromatin. In addition, some of the pathological effects of reduced telomerase related to loss of telomerase outside the nucleus, including the increase of mitochondria-free radical formation, have been established. Reduced ROS production changes inversely with expression of TERT in the mitochondria, an action that involves extranuclear TERT.

As observed previously, the magnitude of FMD in human microvessels is maintained but the mechanism changes in response to stress, whereas agonist induced dilation to ACh is simply lost. The reason for this difference is not known, but it has been speculated that ACh is but one of many agonists which can stimulate NO release, whereas shear stress-induced increases in NO involves multiple signaling pathways (ie, ACh binds to muscarinic receptors to induce dilation while redundant pathways may exist for FMD). We have not determined the precise mechanism by which reduced telomerase activity modulates ROS. However, this

![Figure 6. Pharmacological activation of telomerase restores nitric oxide (NO)-mediated dilation in response to flow. Flow-mediated dilation (FMD) was evaluated in isolated adipose and atrial microvessels from subjects with coronary artery disease (CAD) treated with the selective transcriptional telomerase activator AGS 499 (22 nmol/L). Vehicle-treated vessels are historic controls.](Image)

![Figure 7. Proposed mechanism of TERT in regulating balance of mitochondrial ROS and NO to maintain FMD in human microvessels. AGS 499 indicates telomeres activator; ATP, adenosine triphosphate; CAD, coronary artery disease; FMD, flow-mediated dilation; H2O2, hydrogen peroxide; mt, mitochondrial; nuc, nuclear; NAD+, nicotinamide adenine dinucleotide; NO, nitric oxide; O2·−, superoxide; ROS, reactive oxygen species; and TERT, catalytic subunit of human telomerase complex.](Image)
modulation is independent of nuclear transcriptional events and from telomere elongation, suggesting a novel nontelomeric site of action. However, we cannot exclude a role for telomere uncapping, a focus for future investigations.

In this study, we used a physiological stimulus for endothelium-dependent dilation, shear stress. The mechno-chemical signal transduction during shear that leads to dilation has been studied extensively. Depending on the tissue and species, the signaling pathway involves glyocalyx, TRPV4, or other mechano-sensitive ion channels, and focal adhesions tethering endothelial cells to the underlying basement membrane by cytoskeletal elements. Secondary pathways include tyrosine kinases or calcium signaling to activate eNOS and other vasoactive mediators. In human vessels from subjects with CAD, a different signaling pathway is invoked with stimulation of mitochondrial H$_2$O$_2$ production. The present study provides novel evidence that telomerase lies at the intersection of these pathways and regulates the switch between shear-induced mediators. We speculate that augmentation of telomerase activity may improve the coronary and peripheral vascular phenotypes in patients with CAD. However, the potential adverse effects associated with increasing telomerase activity, including accelerated tumor growth, have to be carefully considered.

Both NO and ROS are signaling pathways for mitochondrial–nuclear communication (reviewed by Kotiadis et al and Valerio and Nisoli), and TERT contains both mitochondrial and nuclear targeting sequences. Site-specific phosphorylation of TERT initiates either mitochondrial or nuclear translocation. This puts telomerase in a position to serve as a critical mediator of nuclear-mitochondrial cross-talk and implicates involvement of AKT, which phosphorylates TERT at its nuclear localization site, increasing binding affinity for the nuclear transporter importin-α, promoting translocation. AKT also targets mitochondrial biogenesis and NO signaling (reviewed by Miyamoto et al and Finley and Haigis). A critical role for non-nuclear telomerase activity in regulating this interplay represents a new field of study with implications for diseases in which ROS play a prominent role, such as diabetes mellitus and neurodegenerative diseases and cardiovascular responses to chemotherapies, such as doxorubicin and PK inhibitors.

It is well established that telomerase activity and telomere length decrease during the natural aging process. A parallel reduction in NO bioavailability is seen after the 4th decade of life, reducing vasodilator capacity. The shift toward a more oxidative environment could explain the reduction in NO bioavailability and increase in ROS production observed with CAD and represents a common feature that contributes to the development of other disease phenotypes. The telomerase activator AGS 499, which normalized FMD and ROS production in microvessels from subjects with CAD, also delays onset of ALS in rodent models. Thus, the ability of TERT to regulate NO production could be important beyond vasodilation.

Compared with age-matched controls, subjects with CAD show a similar degree of FMD. Importantly, the underlying mechanism of FMD changes from NO to H$_2$O$_2$. Inhibition of telomerase can initiate this pathological phenotype (H$_2$O$_2$-mediated FMD) in vessels from subjects without CAD in <24 hours. Conversely, in subjects with chronic CAD, stimulation of TERT activity is sufficient to subacutely restore NOS as critical for FMD, likely via generation of NO (L-NAME and c-PTIO inhibitable). Induced upregulation of telomerase activity may be beneficial in CAD because NO is vasoprotective, whereas H$_2$O$_2$ is proinflammatory in the vasculature. With growing evidence that the microcirculatory dynamics influence cardiovascular outcomes (reviewed by Lockhart et al), understanding mechanisms of arteriolar endothelium-dependent dilation is of both physiological and clinical importance.

We have previously demonstrated that in a matter of minutes, an acute increase in intraluminal pressure can evoke the same change in mediator of FMD after telomerase inhibition or as seen in CAD subjects. Using ceramide as a physiological stressor, Freed et al showed a similar shift in vasodilator mechanism as we see with telomerase inhibition. Ceramide is known to suppress TERT expression; thus, it is intriguing to speculate that ceramide activation of ROS is TERT-dependent. Future studies will evaluate whether ceramide and TERT are part of the same pathway modulating mitochondrial redox function.

**Study Limitations**

Our functional studies point toward a critical role of NOS and NO in arteriolar FMD under normal conditions. We cannot rule out a role of prostacyclins, EETs (epoxygenase products), or other EDHFs (endothelial-derived hyperpolarizing factor); however, our prior studies argue against a role of EETs because the enzyme responsible for their production, CYP450 epoxygenase, is directly inhibited by H$_2$O$_2$. Although it is not technically feasible to directly assess NO bioavailability in the small tissue samples used, we used DAF-FM (4-amino-5-methylamino-2',7'-difluorofluorescein diacetate) as a surrogate marker for NO production in cultured human endothelial cells. We observed an elevation in eNOS message and p-eNOS (phospho-endothelial nitric oxide synthase; ser 1117) after activation of TERT with AGS 499. NO production showed a statistically significant increase in cultured HUVECs after treatment with AGS 499. Our combined data in cultured human endothelial cells and isolated vessels strongly suggests that NOS-derived NO is responsible for FMD in vessels from patients without CAD and in vessels from patients with CAD treated with AGS 499 15 to 20 hours (dilation changes from PEG-catalase inhibitable to L-NAME and c-PTIO inhibitable). Future studies will define the underlying differences between the acute versus chronic effects of telomerase deficiency in vivo and in isolated vessel studies.

We used primary human fibroblasts to provide proof of principle data for translocation of TERT in response to oxidative stress. Because of the fact that endothelial cells are difficult to use for transient transfections, normal human fibroblasts were chosen as a surrogate. In addition, fibroblasts have higher levels of mitochondrial number, enabling us to perform ATP production assays, which are extremely difficult to perform in primary human endothelial cells.

We cannot eliminate a contribution of vascular smooth muscle cells to the observed phenotype. However, we think
this is unlikely because inhibition of telomerase did not alter vascular smooth muscle cells function (Figure 1), whereas activation of telomerase restored physiological endothelial-dependent NOS-mediated (NO) dilation to flow. As telomerase activity is known to increase cellular proliferation, activation of telomerase in vascular smooth muscle cells would be counterproductive to vascular health and would be expected to impair vasoactive function. In mice with systemic deletion of telomerase, a decrease in atherosclerosis-like phenotype has been reported. Similarly published evidence supports that the well-established anti-inflammatory and anti-atherosclerosis properties of PPARγ are at least in part caused by suppression of telomerase activity in the vascular smooth muscle cells.

Conclusions

In summary, we demonstrate direct functional evidence that decreased telomerase activity is sufficient to initiate the transition from NO to H₂O₂ as the primary mediator of FMD in the human resistance vasculature, recapitulating the microvascular properties of telomerase activity in the vascular smooth muscle cells. Moreover, we show that increasing compounds the other authors report no conflicts.

Disclosures

E. Priel filed a patent on AGS 499 and other derivatives as telomerase inhibitors. E. Priel, A.J. Donato, and R.G. Morgan were supported by the National Heart, Lung, and Blood Institute of the National Institutes of Health (NIH) T32HL007792 and American Heart Association (AHA) 14POST18780022. A.J. Donato and R.G. Morgan were supported by a T-32 grant from the National Institute of General Medical Sciences. This work was supported by R01 HL113612 (D.D. Gutterman) and 1P20OD018306 (A.M. Beyer). J.K. Freed was supported by a T-32 grant from the National Institute of General Medical Sciences (high-performance liquid chromatography).

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References

cellular ROS production, improves mitochondrial function, and inhibits p53, and p21(CIP1), but not p16(INK4a).

Telomerase reverse transcriptase binds to and protects mitochondrial DNA. 2001;280:H2087–H2093.

AM J Physiol
Molecular mechanisms of ceramide-mediated telomerase inhibi-


Beyer AM, Durand MJ, Hockenberry J, Gamblin TC, Phillips SA, Gutterman DD. Acute rise in intraluminal pressure shifts the media-
What Is Known?

- Endothelial dysfunction predicts future cardiovascular events.
- Under physiological conditions, the primary mediator of flow-mediated dilation (FMD) is nitric oxide (NO), whereas in pathology, such as coronary artery disease (CAD), the FMD is mediated by the mitochondria-derived, proinflammatory hydrogen peroxide.
- The catalytic subunit of telomerase elongates telomeres in the nucleus to prevent cellular aging and promote proliferation.
- A non-nuclear role for catalytic subunit of telomerase in regulating levels of mitochondrial-derived reactive oxygen species is established in other cell types but its role in vascular cells has not been examined.

What New Information Does This Article Contribute?

- Telomerase activity is decreased in subjects with CAD without measurable telomere shortening.
- Pharmacological inhibition of telomerase activity in arterioles from patients without CAD causes a transition in mechanism of FMD from NO to hydrogen peroxide, similar to the mechanism observed in vessels from patients with CAD.
- Transcriptional activation of catalytic subunit of telomerase restores NO as the dominant mediator of FMD in subjects with CAD.
- Telomerase activity regulates endothelial production of either NO (health) or hydrogen peroxide (disease) in response to endothelial shear stress in the microcirculation.

Endothelial release of NO mediates FMD under physiological conditions and serves to prevent vascular smooth muscle proliferation and inflammation. In subjects with CAD, FMD is mediated by mitochondrial-derived reactive oxygen species, specifically hydrogen peroxide, a proinflammatory and proatherosclerotic dilator. The present study identifies telomerase activity as a regulator of the underlying mechanism of FMD. Telomerase activity represents a novel target to modulate FMD and, thereby, vascular health.
Critical Role for Telomerase in the Mechanism of Flow-Mediated Dilation in the Human Microcirculation
Andreas M. Beyer, Julie K. Freed, Matthew J. Durand, Michael Riedel, Karima Ait-Aissa, Paula Green, Joseph C. Hockenberry, R. Garret Morgan, Anthony J. Donato, Refael Peleg, Mario Gasparrì, Chris K. Rokkas, Janine H. Santos, Esther Priel and David D. Gutterman

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Nitric Oxide production was estimated by following the conversion of DAF2 to DAF2-triazole (DAF2-T) by HPLC as published previously\(^1\). Briefly DAF2-DA (5 µM) was added to cells for 1 hour. Cells were harvested and lysed and supernatants were filtered through a 5Kd cut-off microcon filter. Eluent (80 µl) was injected onto a C18 reverse-phase HPLC column and separated using a mobile phase of sodium phosphate (10 mM, pH 7.5) and acetonitrile (5%) with a flow-rate of 1 ml/mn. DAF2-T was detected by fluorescence (ex 490 nm, em 515 nm) and quantified by area under the curve. Data was normalized to control for any given experimental date. Outliers were identified using Thompson tau method.

Protein expression was evaluated using standard Western blot techniques. Cells were lysed centrifuged and separated from debris in cold lysis buffer supplemented with a protease inhibitor cocktail (Roche)\(^2-4\). Isolated protein was quantified using a Bradford micro assay. Protein (10-50 µg) was subjected to 10% SDS-PAGE and transferred to PVDF membranes prior to blotting with polyclonal antibodies against p-eNOS/ eNOS (1:1000 BD Biosciences) or GABDH.

Immunohistochemistry: was performed at Children’s Hospital of Wisconsin pathology core. In brief, optimal immunostaining protocol was developed with the Leica Bond Max Immunostainer platform. Tissues were fixed in 10% neutral-buffered formalin, processed and embedded into paraffin blocks. Sections were cut at 4µm, mounted on positively charged glass slides and incubated at overnight at 45°C. All paraffin sections were de-waxed prior to staining on the instrument. The TERT antibody required antigen retrieval using Leica H1 antigen retrieval reagent for 10 minutes. The antibody was detected and visualized using Bond Polymer Refine Detection System (DS9800) with the addition of a DAB enhancer (AR9432), using the MOD F protocol/software installed by Leica field service technicians. An optimal concentration of 1:100 was determined for TERT. All slides were counter-stained with hematoxylin and cover slipped using a synthetic mounting media. Omission of the primary antibody served as negative control.

Supplemental Study limitations

Influence of Clinical Parameters and Tissue Origin

We used adipose tissue from several body sites, both visceral and subcutaneous, potentially increasing the variability of our data since vessels from these sites can demonstrate unique responses to selective vasodilators\(^5\). However our experience indicates that in responses to flow and mechanisms of FMD in health and disease are similar in arterioles across fat depots, and similar to responses observed in human coronary arterioles. Thus the same change in dilator mechanism to flow that we observe in adipose vessels exposed to an inhibitor of telomerase, occurs in coronary arterioles in response to an elevation in arterial pressure \(^6\) or in vessels taken from patients with CAD \(^7\). Thus the switch from NO to H\(_2\)O\(_2\) as a mediator of FMD is a common
response to acute or chronic vascular stressors and appears conserved across a limited number of tissues.

Since discarded de-identified specimens were used, it was not possible to control for all clinical variables such as duration and extent of CAD, effect of medications, and presence of risk factors such as hypertension, diabetes, obesity or high cholesterol. No heart tissue and only limited amounts of adipose tissue from truly healthy subjects were included in this study since all underwent surgery, limiting our conclusions to pathophysiological conditions. Diseases such as obesity, hypertension, and hyperlipidemia may be confounders in contributing to mitochondrial ROS production. The high prevalence of diabetes in our cohort makes it difficult to exclude an effect of diabetes on our observed phenotypes. Importantly when stratified by presence or absence of CAD, each group had similarly elevated overall BMI (Table 1). Heterogeneity among subjects enhances the strength of our findings that decreased telomerase activity contributes to the development of CAD by enabling a rise in mitochondrial free radical production.

While we cannot completely exclude effects of medications on the observed changes we believe it is not likely that there were residual drug effects since vessels were incubated and repetitively washed in buffer and/or media for several hours prior to experimentation. We have previously shown that this is sufficient for washout of some medications with long half-lives. In addition, the effects of applied pharmacological dilator agents are reversed by washing with fresh buffer, suggesting that any residual influence is minimized. Future studies will need to address the influence of specific cardiovascular risk factors and other phenotypical characteristics.

Due to the limitation in tissue volume (each microvessel weighs less than 10 µg wet weight) we cannot reliably evaluate telomerase protein levels in the microvessels and instead presented data from whole heart tissue. We did measure telomere length in isolated arterioles and found it to be significantly shorter in heart tissue compared to microvascular samples, with no difference between CAD and non CAD in either tissue. Both measurements are consistent with the idea that the effects of TERT are telomere-independent and can be normalized by stimulating TERT transcription (AGS 499) in CAD tissue.

**Online Figure I: Immunohistochemistry of isolated human coronary vessels from subjects with and without CAD.** TERT expression was observed in both SMC and vascular endothelial cells. No quantification was performed. Slides were scanned and imaged using Hamamatsu digital imaging software NDPview2. Indicated Scale bar = 100µM for each individual image.

**Online Figure II: Telomerase Activation Increased NOS levels and NO Production in Cultured Endothelial Cells.** Cultured HUVECs were treated with the TERT activator or inhibitor of telomerase activity (AGS 499 or BIBR 1532 respectively). All values displayed as fold change to Vehicle. A) Telomerase activation resulted in transcriptional increase eNOS mRNA while inhibition of telomerase activity had no effect; N≥5. B) Telomerase activation increased p-eNOS S1117 without effecting total protein (both normalized to GABDH) N≥5, C) Telomerase activation increased NO producing while telomerase inhibition had no effect. No measured via DAF-2A/HPLC. * p<0.05 vs. Vehicle t-Test Tukey post hoc analysis.
On line References


**Online Figure II**

(A) **eNOS mRNA** (fold change from vehicle)

- Vehicle
- AGS 499
- BIBR 1532

(B) **Protein** (fold change from Vehicle)

- T-eNOS
- GAPDH
- p-eNOS
- GAPDH

(C) **DAF-2A** (fold change from Vehicle)

- Vehicle
- AGS 499
- BIBR 1532