MicroRNA-34a Plays a Key Role in Cardiac Repair and Regeneration Following Myocardial Infarction

Yanfei Yang, Hui-Wen Cheng, Yiling Qiu, David Dupee, Madyson Noonan, Yi-Dong Lin, Sudeshna Fisch, Kazumasa Unno, Konstantina-Ioanna Sereti, Ronglih Liao

Rationale: In response to injury, the rodent heart is capable of virtually full regeneration via cardiomyocyte proliferation early in life. This regenerative capacity, however, is diminished as early as 1 week postnatal and remains lost in adulthood. The mechanisms that dictate postinjury cardiomyocyte proliferation early in life remain unclear.

Objective: To delineate the role of miR-34a, a regulator of age-associated physiology, in regulating cardiac regeneration secondary to myocardial infarction (MI) in neonatal and adult mouse hearts.

Methods and Results: Cardiac injury was induced in neonatal and adult hearts through experimental MI via coronary ligation. Adult hearts demonstrated overt cardiac structural and functional remodeling, whereas neonatal hearts maintained full regenerative capacity and cardiomyocyte proliferation and recovered to normal levels within 1-week time. As early as 1 week postnatal, miR-34a expression was found to have increased and was maintained at high levels throughout the lifespan. Intriguingly, 7 days after MI, miR-34a levels further increased in the adult but not neonatal hearts. Delivery of a miR-34a mimic to neonatal hearts prohibited both cardiomyocyte proliferation and subsequent cardiac recovery post MI. Conversely, locked nucleic acid–based anti–miR-34a treatment diminished post-MI miR-34a upregulation in adult hearts and significantly improved post-MI remodeling. In isolated cardiomyocytes, we found that miR-34a directly regulated cell cycle activity and death via modulation of its targets, including Bcl2, Cyclin D1, and Sirt1.

Conclusions: miR-34a is a critical regulator of cardiac repair and regeneration post MI in neonatal hearts.

Modulation of miR-34a may be harnessed for cardiac repair in adult myocardium. (Circ Res. 2015;117:450-459. DOI: 10.1161/CIRCRESAHA.117.305962.)

Key Words: cellular proliferation ■ miR-34a, mouse ■ myocardial infarction ■ myocytes, cardiac ■ regeneration
Yang et al. miR-34a in Neonatal Post-MI

451

the role of miR-34a in regulating endogenous cardiac regeneration in the early postnatal heart remains largely unknown.

In this study, we reveal that cardiac miR-34a levels are low in the early postnatal period, in conjunction with preserved regenerative capacity, and soon rise to adult levels within 1 week after birth. Overexpression of miR-34a in early postnatal mice was found to limit cardiomyocyte proliferation and cardiac regeneration with injury. Conversely, antagonism of miR-34a improved post-MI cardiac function in adult mice in part through modulation of cell cycle and survival proteins, including Bcl2, Cyclin D1, and Sirt1. Collectively, our findings support miR-34a as a key parameter regulating endogenous postinjury cardiac repair and regeneration.

Methods

Animal Models

Eight-week-old male C57BL/6J and pregnant female mice (E16; #027) were purchased from Charles River Laboratories and housed in a temperature-controlled environment with 12 hours light/dark cycles with food and water available ad libitum. MI was generated by ligating the main branch coronary artery in mice 1 day post birth (neonatal) or at 8 weeks (adult) as described previously.21 For surgical MI, adult mice were anesthetized with isoflurane (2%). A rodent ventilator (model 845; Harvard Apparatus Inc) was used to supply oxygen with 2% isoflurane during the surgical procedure. Neonatal mice were anesthetized on ice for 6 minutes and kept on ice during the surgical procedure. Adult and neonatal chests were opened by a horizontal incision through the muscle between the fourth and the fifth intercostal space. The pericardium was then removed in adult mice only. The descending coronary artery was permanently ligated with a suture: 8-0 prolene (BV175-7) for adults and 10-0 nylon (BV100-4) for neonates. After surgery, the thoracic wall and skin were closed with sutures: 5-0 absorbable (CV-23) for adult and 8-0 prolene for neonates. Sham-operated animals underwent an identical surgical operation without occlusion of the coronary artery. To prevent any postoperative discomfort, adult animals received buprenorphine (0.03–0.06 mg/kg). All animal procedures were performed under the guidelines of Harvard Medical School, the Institutional Animal Care and Use Committee, and the National Society for Medical Research.

miRNA Mimic and Locked Nucleic Acid miRNA Inhibitor Delivery

Immediately after neonatal MI, a miR-34a mimic (miRIDIAN) or control miRNA mimic was injected intramyocardially into each mouse (5 mg/kg). Six hours and 2 days after MI in adults, a miR-34a locked nucleic acid (LNA) inhibitor (miRCURY) or control LNA inhibitor was injected intravenously via tail vein into each mouse (5 mg/kg). Noninvasive transthoracic echocardiography, performed as previously described,22 was used to determine the extent of initial injury 4 hours after MI. Structural and functional alterations were also evaluated through noninvasive transthoracic echocardiography at 1, 3, and 7 days after cardiac injury.

Nonstandard Abbreviations and Acronyms

LNA locked nucleic acid
MI myocardial infarction
miRs microRNAs

Figure 1. Myocardial recovery after myocardial infarction in neonatal hearts. A, Fractional area change (FAC%) in neonatal and adult hearts 4 hours after myocardial infarction (MI) compared with sham controls measured by echocardiographic analysis. *P<0.05 vs sham; #P<0.05 vs neonate. n=9 for neonate sham; n=7 for neonate and adult MI; n=6 for adult sham. B, Terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end labeling (TUNEL) assay (red) in ventricular cross-sections of neonatal and adult hearts 8 hours after MI; samples were counterstained with Hoechst (blue) dye. Representative images of whole ventricular cross-sections (upper), yellow boxes indicate representative infarcted areas shown at higher magnification (lower). C, Quantification of TUNEL-positive nuclei (n=3 for neonate and adult) shown in B. D to G, FAC% measurements of adult (D) and neonatal (F) post-MI hearts by echocardiographic analysis. *P<0.01 vs sham. Left ventricular diameter at end-diastole (LVIDd/mm) measurements of adult (E) and neonatal (G) post-MI hearts by echocardiographic analysis. *P<0.05 vs sham. n=6 for days 1 and 3 sham; n=5 for day 7 sham and MI; n=7 for days 1 and 3 MI. H and I, Postmortem measurements of heart weight (HW; mg) and body weight (BW; g) ratios of adult (H) and neonatal (I) post-MI hearts. *P<0.05 vs sham. n=3 for adult; n=5 for neonate.
Results

Cardiac Regeneration in Neonatal Hearts Is Associated With Increased Cell Cycle Activity

To examine cardiac regenerative capacity in neonatal and adult mouse hearts, MI was induced in 1-day-old neonatal and 8-week-old adult mice by coronary ligation. The degree of cardiac injury was determined by decrements in contractile function using noninvasive transthoracic echocardiography 4 hours post surgery and was comparable with MI in neonatal and adult animals (Figure 1A) at similar heart rates (Online Figure I). Similarly, early post-MI cell death was comparable between neonatal and adult hearts, with similar degrees of terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end labeling (TUNEL) staining (Figure 1B and 1C). After MI, cardiac performance progressively declined in adult hearts relative to sham-operated counterparts, with reduction in cardiac function (Figure 1D) and dilatation of ventricular chambers (Figure 1E). In contrast, although early indices of cardiac performance were reduced in neonatal hearts with injury, cardiac function and chamber dimensions were restored to baseline sham levels within 7 days post MI, consistent with significant cardiac recovery (Figure 1F and 1G). Likewise, whereas adult hearts underwent cardiac remodeling after injury with increased heart weight/body weight ratios (Figure 1H), neonatal heart weight/body weight ratios were comparable with sham-operated controls (Figure II). Consistent with cardiac recovery, neonatal hearts post MI exhibited limited fibrotic tissue replacement (Figure 2A and 2B) and increased cardiomyocyte cell cycle activity (Figure 2C and 2D). In contrast, adult hearts post MI exhibited limited cardiomyocyte proliferation. Note that collagen fibril formation was not mature at 1 day post MI and was therefore not quantified using staining with Masson Trichrome.

miR-34a Expression Is Increased in Adult but Not in Neonatal Mouse Hearts Post MI

Expression of miR-34 family members (miR-34a, miR-34b, and miR-34c) was examined in the mouse left ventricle, intestine, skeletal muscle, lung and liver tissues, with miR-34a found to be most highly expressed in heart tissue (Figure 3A–3C). Using in situ hybridization, we found that miR-34a is expressed in both cardiomyocytes and noncardiomyocytes within the heart (Figure 3D; U6 and scrambled miRNA control in Online Figure II). Real-time polymerase chain reaction revealed a similar degree of miR-34a expression between neonatal rat cardiomyocytes and neonatal rat cardiac fibroblasts (Figure 3E),
Yang et al

miR-34a in Neonatal Post-MI

453

with increasing expression during the first week of life, reaching adult levels at 1 week after birth (Figure 3F). With cardiac injury, miR-34a levels remained low in early postnatal hearts (Figure 3G), whereas in adult hearts, miR-34a levels initially declined within 24 hours post cardiac injury and then increased progressively within 1 week post MI (Figure 3H).

miR-34a Overexpression Prevents Post-MI Recovery in Neonatal Hearts

To determine whether miR-34a modulates post-MI cardiac regeneration in early postnatal hearts, miR-34a levels were augmented by delivery of a miR-34a mimic to the myocardium at the time of MI with confirmed miR-34a sustained expression at 7 days post injection (Figure 4A). Increased miR-34a levels inhibited functional post-MI recovery in neonatal mouse hearts (Figure 4B), whereas no effect was observed with delivery of a control miR mimic (Figure 4C). miR-34a overexpression reduced post-MI cardiomyocyte proliferation (Figure 4D and 4E), increased programmed cell death (Figure 4F; Online Figure III) and resulted in greater tissue fibrosis (Figure 4G and 4H). Collectively, these results suggest that low miR-34a levels in the early postnatal heart enable cardiac regeneration.

Inhibition of miR-34a Improves Cardiac Function in Adult Hearts Post MI

To determine whether antagonizing miR-34a enhances cardiac regeneration in adult hearts, LNA-based anti–miR-34a (LNA-34a) or scrambled negative control was delivered intravenously at 6 hours and 2 days after MI in adults (Online Figure IV A). LNA-34a effectively inhibited miR-34a expression levels (Figure 5A), with subsequent improvement in post-MI cardiac function (Figure 5B), diminished cardiac remodeling (Figure 5C), and reduced fibrosis (Figure 5D and 5E) in adult hearts. Inhibition of miR-34a levels also increased cardiomyocyte cell cycle activity (Figure 5F and 5G) and prevented cell death in adult hearts post MI (Figure 5H; Online Figure IVB),
suggesting that upregulation of miR-34a may contribute to the loss of endogenous regeneration in the adult heart.

**miR-34a Overexpression Inhibits Neonatal Cardiomyocyte Cell Cycle Progression and Survival**

To determine whether miR-34a regulates cardiac regeneration in a cell autonomous manner, neonatal cardiomyocytes were directly transfected with a miR-34a mimic in vitro. Immunostaining of Dy547-labeled miRNA mimic suggested efficient entry into neonatal cells (Figure 6A) with a transfection yield of 97.4±1.6% and specific upregulation of miR-34a expression (Figure 6B) without change in the expression of other miR-34 family members such as miR-34c (Figure 6C).

Increasing expression levels of miR-34a reduced incorporation of the thymidine analogue, 5-ethynyl-2′-deoxyuridine, in neonatal cardiomyocytes suggesting inhibition of entry into S phase of the cell cycle (Figure 6D and 6E). Moreover, direct overexpression of miR-34a induced cell death in neonatal cardiomyocytes (Figure 6F and 6G). These data indicate that miR-34a directly modulates cardiomyocyte proliferation and survival in a cell autonomous manner.

**miR-34a Regulates Cardiomyocyte Proliferation and Survival Through Expression of Bcl2, Cyclin D1, and Sirt1**

To identify potential miR-34a targets responsible for modulating proliferation and survival of cardiomyocytes post MI, we examined genome-wide mRNA expression and focused on those genes selectively downregulated in adult hearts relative to neonatal counterparts, with 7 genes predicted as miR-34a targets. We further prioritized follow-up investigations of 4 genes, Sirt1, Cyclin D1, Bcl2, and Jag1, previously implicated in cellular aging, cell cycle activity, survival, and the Notch pathway, all potentially important components of cardiac regeneration. Among these 4 genes, we found that miR-34a did not inhibit Jag1 3′ untranslated region luciferase activity (Online Figure VA) or protein levels (Online Figure VB and VC) indicating that Jag1 is not a target of miR-34a in our experimental setting. miR-34a, however, effectively reduced protein levels for Sirt1, Cyclin D1, and Bcl2 in both cardiomyocytes in vitro and whole hearts in vivo (Figure 7A–7C; Online Figure VIA and VIB). Conversely, inhibition of miR-34a by LNA in adult post-MI hearts increased protein levels of all 3 target genes (Figure 7D and 7E). The 3′ untranslated
region luciferase assays confirmed miR-34a modulation of Bcl2, Cyclin D1, and Sirt1 expression, with loss of miR-34a dependency with selective mutation of the miR-34a binding site (Figure 7F). miR-34a-mediated downregulation of target genes was rescued via adenoviral-mediated overexpression of Bcl2, Cyclin D1, and Sirt1 (Figure 7G–7I). Maintenance of Bcl2 and Cyclin D1 protein levels in neonatal cardiomyocytes preserved cardiomyocyte proliferation and cell cycle capacity as well as protected against cell death mediated by the miR-34a expression (Figure 7J and 7K; Online Figure VIIA and VIIB). Of note, Sirt1 was effective only in protecting against cell death but not loss of cell proliferation (Figure 7J and 7K). Collectively, these results suggest that the downregulation of Bcl2, Cyclin D1, and Sirt1 may play important roles in mediating the effects of miR-34a on MI cardiomyocyte cell death and proliferation.

Discussion
Recent studies have revealed that neonatal rodent hearts maintain a significant regenerative capacity after cardiac injury, particularly if such injury is imposed within the first few days after birth.3,4,21,28–30 The processes mediating the regenerative capacity of neonatal hearts and the mechanisms by which this regenerative capacity is lost early after birth still remain unclear. Consistent with prior reports, we found that <2-day-old neonatal hearts recovered almost fully from significant MI injury without loss of contractile function and with limited evidence of scarring/remodeling.4,31,32 Prior literature has identified miRNAs as critical regulators of cellular processes including senescence, proliferation, and survival. miR-34a levels have been shown to increase in response to the aging process in several tissues, including the heart.16,17 Our results demonstrate that the expression of miR-34a early after birth...
miR-34a overexpression inhibits neonatal cardiomyocyte cell cycle progression and survival. A, Neonatal rat ventricular myocytes (NRVMs) transfected with 20 nmol/L Dy547-labeled miRNA mimic were fixed 48 hours post transfection and immunostained against cardiac troponin T (cTnT, green). DAPI (blue) was used for nuclear staining. B and C, Real-time polymerase chain reaction analysis of miR-34a (B) and miR-34c (C) levels in 20 nmol/L negative control miRNA (NC) or miR-34a mimic transfected NRVMs. Sno202 RNA was used as an internal normalizing control for small RNAs. *P<0.05 vs NC. n=3 for all groups. D, NRVMs transfected with 20 nmol/L NC or miR-34a mimic were treated with a thymidine analogue (5 μmol/L 5-ethyl-2′-deoxyuridine [EdU]) for 48 hours and stained with an EdU assay kit (red). E, Quantification of EdU-positive cardiomyocytes shown in D. *P<0.05 vs NC. n=4 for NC; n=5 for miR-34a. F, NRVMs transfected with 20 nmol/L NC or miR-34a mimic for 48 hours were stained with a terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end labeling (TUNEL) assay kit (red). G, Quantification of TUNEL-positive cardiomyocytes shown in F. *P<0.05 vs NC. n=5 for all groups.

Figure 6. miR-34a overexpression inhibits neonatal cardiomyocyte cell cycle progression and survival. A, Neonatal rat ventricular myocytes (NRVMs) transfected with 20 nmol/L Dy547-labeled miRNA mimic were fixed 48 hours post transfection and immunostained against cardiac troponin T (cTnT, green). DAPI (blue) was used for nuclear staining. B and C, Real-time polymerase chain reaction analysis of miR-34a (B) and miR-34c (C) levels in 20 nmol/L negative control miRNA (NC) or miR-34a mimic transfected NRVMs. Sno202 RNA was used as an internal normalizing control for small RNAs. *P<0.05 vs NC. n=3 for all groups. D, NRVMs transfected with 20 nmol/L NC or miR-34a mimic were treated with a thymidine analogue (5 μmol/L 5-ethyl-2′-deoxyuridine [EdU]) for 48 hours and stained with an EdU assay kit (red). E, Quantification of EdU-positive cardiomyocytes shown in D. *P<0.05 vs NC. n=4 for NC; n=5 for miR-34a. F, NRVMs transfected with 20 nmol/L NC or miR-34a mimic for 48 hours were stained with a terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end labeling (TUNEL) assay kit (red). G, Quantification of TUNEL-positive cardiomyocytes shown in F. *P<0.05 vs NC. n=5 for all groups.

is closely associated with the loss of regenerative potential. Consistent with this observation, antagonizing miR-34a expression in adult mouse hearts markedly improved cardiac repair and post-MI remodeling, whereas increasing miR-34a expression in early postnatal hearts impaired regeneration. Together, our results suggest that miR-34a may represent a critical key regulator of repair/regeneration capacity in the heart and may be harnessed for enhancing endogenous repair mechanisms in the adult heart post MI.

The therapeutic effects of miR-34a have been previously examined in the heart with mixed results. Consistent with our findings, miR-34a has been found to be highly expressed in cardiac tissue, and loss of miR-34a improves cardiac function and reduces cell death in aging hearts. In addition, acute inhibition of miR-34a using LNA or an antagonist delivered intravenously within several hours post MI also significantly improved cardiac function at 2 weeks. Other studies, however, have suggested that later delivery of LNA subcutaneously post MI was only effective when targeting multiple miR-34 family members, but not miR-34a alone, at even a much higher concentration. Our findings are consistent with these reports, with early intravenous LNA targeting miR-34a found to improve post-MI cardiac function. It is likely that the mode of delivery and subsequent kinetics of miR-34a inhibition, as well as the timing of miR antagonism, dictate the complex post-MI regenerative response. miR-34a was noted in our studies to be expressed in both cardiomyocytes and noncardiomyocytes, and potential discordant effects in these cells with miR-34a inhibition may further be critical in dictating overall response. In settings of established cardiac injury, it is likely that antagonism of miR-34a alone is insufficient to promote cardiac regeneration, and a more broad inhibition of miR-34 family members is required.

Several molecular mediators and signaling pathways have been previously implicated as regulators of neonatal cardiomyocyte regeneration, including miR-15 family members and Yap. Cardiac-specific overexpression of miR-195 (a miR-15 family member) impaired neonatal post-MI regeneration with a reduced proportion of mitotic cardiomyocytes. Yap belongs to the evolutionarily conserved Hippo pathway that controls organ size and has been implicated as a regulator of cardiomyocyte proliferation in both embryonic hearts and adult post-MI hearts. miR-34a shares several similarities with these mediators of neonatal cardiac regeneration. Both miR-34 and miR-15 family members have been identified as tumor suppressors, with coregulation in cancer cells to control cell cycle progression in a synergistic and Rb-dependent fashion. Yap inhibits miR-34a processing by sequestering p72 from the microprocessor. Moreover, our data suggest that miR-34a levels are higher in adult than in neonatal hearts and are further upregulated in adult post-MI hearts, with a pattern consistent with other negative regulators of cardiac regeneration, such as miR-15 family members.

To further assess the mechanism by which miR-34a regulates cardiomyocyte proliferation and cardiac regeneration, we specifically examined the expression of 3 miR-34a targets which have previously been linked to cellular proliferation and survival, including Sirt1, Cyclin D1, and Bcl2. Sirt1, a class III histone deacetylase and an age-related protein, has been shown to prevent neonatal cells from death in vitro and to slow aging-associated apoptosis and senescence. Cyclin D1 plays important roles in regulating cell proliferation in the developing heart. Overexpression of Cyclin D1 in the adult heart improves cardiac function after ischemic injury via promoting cardiomyocyte proliferation and survival. Bcl2, an antiapoptotic protein, promotes cell survival in many organs, including the heart. More importantly, Bcl2 overexpression has been shown to promote cardiomyocyte cell cycle activity in postnatal mouse hearts. Our data suggest that the prosurvival functions of Bcl2, Cyclin D1, and Sirt1, as well as proproliferative nature of Bcl2 and Cyclin D1, are critical to miR-34a effects, and overexpression of these miR-34a target genes rescues cardiomyocyte cell cycle activity in miR-34a–expressing cells.
value of miR-34a is further complicated by the expression of miR-34a and potential differing effects in a number of cell types. For instance, given miR-34a’s tumor suppressive activity, intravenously delivered liposomal-based miR-34a mimics have entered into phase I clinical trials as emerging cancer therapeutics. It remains to be seen whether chronic exposure to miR-34a may affect cardiac function in an adverse manner, as has been observed with other anticancer therapies.41

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Disclosures

None.

References


**Novelty and Significance**

**What Is Known?**

- After myocardial infarction, the regenerative capacity of adult mouse hearts is limited.
- Neonatal mouse hearts, however, can mount a robust regenerative response to myocardial infarction injury leading to cardiac recovery.
- The mechanisms underlying robust cardiac regeneration in neonatal mouse hearts remain unknown.

**What New Information Does This Article Contribute?**

- microRNA-34a (miR-34a) is highly expressed in adult hearts and is associated with the loss of postinjury regenerative capacity.
- Overexpression of miR-34a in neonatal hearts inhibits cardiac regeneration.
- Suppression of miR-34a in adult hearts increases post–myocardial infarction cardiac regeneration and function.
- miR-34a modulates cardiac regeneration through targeting of Bcl2, Cyclin D1, and Sirt1 in cardiomyocytes.

The capacity for robust cardiac regeneration in response to injury in neonatal hearts remains among the most promising avenues for understanding endogenous regenerative mechanisms. Although microRNAs have been implicated in several cellular processes, their role in mediating cardiac regeneration in neonatal and adult hearts remains unknown. In this study, we find that miR-34a is increased in the myocardium soon after birth and is associated with the loss of regenerative capacity. Exogenous expression of miR-34a in neonatal hearts abrogates regeneration, and suppression of miR-34a in adult hearts improves postinjury regeneration and cardiac function. Finally, we find that miR-34a inhibits cardiomyocyte proliferation and induces cardiomyocyte cell death through targeting Bcl2, Cyclin D1, and Sirt1. These results suggest that inhibition of miR-34a may present a therapeutic option to enhance endogenous repair in adult post–myocardial infarction hearts.
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SUPPLEMENTAL MATERIAL

Supplemental Methods

Materials
Laboratory reagents were purchased from Sigma. The EdU staining kit and TUNEL staining kits were acquired from Life Science Technologies and Roche, respectively. miRIDIAN microRNA mimic negative control (cel-miR-67) and miR-34a were purchased from Dharmacon. miRCURY LNA™ microRNA power inhibitors were used for suppression of miR-34a. microRNA ISH optimization kit 1 and LNA-modified miR-34a (5’- and 3’-DIG-labeled) probes were obtained from Exiqon. Antibodies were obtained from Sigma (anti-α-tubulin and anti-β-actin), Thermo Scientific (anti-cardiac Troponin T), Cell Signaling (anti-Cyclin D1), Millipore (anti-Sirt1), BD Pharmingen (anti-Bcl2), Life Technologies (anti-Digoxigenin) and Abcam (anti-Histone H3, phospho S10, pH3). Sirt1 adenovirus was a kind gift from Dr. J. Sadoshima (Rutgers New Jersey Medical School). Bcl2 and Cyclin D1 adenoviruses were obtained from Vector Biolabs.

Isolation and Culture of Neonatal Rat Ventricular Myocytes
Primary cultures of ventricular myocytes were obtained from 1-day-old Wistar rats (cat#003, Charles River Laboratories) and prepared using a Neonatal Heart Dissociation Kit (Miltenyi Biotec Inc.) according to the manufacturer instructions. Briefly, after enzymatic digestion, ventricles were subjected to mechanical dissociation using a gentleMACS™ Dissociator. Cell suspensions were applied to a discontinuous Percoll gradient and myocyte layers were harvested and cultured as described previously.1

RNA Isolation and Quantitative Reverse Transcription–Polymerase Chain Reaction (qRT-PCR)
RNA was extracted from hearts or cultured cells in accordance with the manufacturer protocol for the miRNeasy Mini Kit (Qiagen), from which qRT-PCR TaqMan® MicroRNA Assays of miR-34a, miR-34b, miR-34c and snoRNA202 from Life Technologies were performed.

Immuno-cytochemistry
Cells were fixed with PBS containing 4% paraformaldehyde, permeabilized with 0.2% Triton X-100 PBS, blocked with PBS containing 3% BSA and incubated with the desired primary antibodies, including anti-cardiac troponin T (dilution 1:400) and anti-pH3 (dilution 1:200). Alexa Fluor 488 or Alexa Fluor 555 Dye-conjugated secondary antibodies (Invitrogen) were used for fluorescent confocal analysis (dilution of 1:200).

Heart Fixation and Histology
Harvested hearts were arrested in diastole with 3M KCl and fixed with 10% buffered formalin solution. Hearts remained overnight in 10% buffered formalin solution at 4°C and were then imbedded in paraffin for analysis by histology. Samples were sectioned at the Histology Core Facility at Beth Israel Deaconess Medical Center.

Immuno-histochemistry
Formalin fixed tissue slides were de-paraffinized via incubation at 65°C for 30 minutes, followed by washing twice with xylene, and re-hydrating through sequential three-minute incubations in ethanol (100%, 100%, 90%, 80%) and water. Antigen retrieval was performed by microwaving in citrate buffer (0.1 M, pH 6.0) for 10 minutes. Slides were incubated with primary antibodies against pH3 (Abcam) and cardiac-troponin T (Thermo Scientific) for 1 hour at 37°C, followed by incubation with appropriate secondary antibodies under the same conditions. Slides were subsequently washed and mounted with DAPI-containing medium (Vector Labs). Infarct border zones were scanned for the presence of pH3 positive cardiomyocytes (CM) through examination of nine randomly chosen fields within each section with a LSM 700 (Zeiss) confocal microscope. Image acquisition and analysis were performed in a blinded
fashion. Myocyte borders were defined by staining of cardiac tissue with Alexa Fluor 555-conjugated wheat germ agglutinin (WGA) (Invitrogen). Apoptotic cell death was determined by TUNEL staining (Roche) according to manufacturer instructions.

**miRNA in situ Hybridization (ISH)**
LNA-modified probes for miR-34a (5’- and 3’-DIG-labeled), scrambled microRNA control (5’- and 3’-DIG-labeled), U6 snRNA control (5'-DIG-labeled), microRNA ISH buffer and Proteinase K were purchased from Exiqon. Formalin-fixed, paraffin-embedded tissue sections were subjected to *in situ* hybridization at 55°C, according to manufacturer instructions. Sections were then immunostained with anti-DIG primary antibody and Alexa Fluor 555 Dye-conjugated secondary antibody (Life technologies).

**Transfection and Luciferase Assay**
Cells were transfected with either a miRIDIAN miR-34a mimic or a negative control, cel-miR-67 (Dharmacon), using X-tremeGENE HP transfection reagent (Roche) according to the manufacturer’s instructions. 48 hours after transfection, cells were lysed with Passive Lysis Buffer (Promega) and luciferase activity was measured using the luciferase assay system (Promega) with a SpectraMax M5 Plate Reader (Molecular Devices). For the 3’UTR luciferase assay, the Bcl2, Cyclin D1 and Sirt1 3’UTR miR-34a target site (sequence with miR-34a binding sites) or a control site (sequence with mutated miR-34a binding sites) was cloned into the pMIR-REPORT vector (Ambion).

**Echocardiography**
Echocardiograms were obtained at 4 hours, 1 day, 3 days and 7 days following MI with a Vevo 2100 Imaging digital ultrasound system (VisualSonics) using a 22-55 MHz (MS550D) for adult and 30-70MHz (MS700) transducer for neonatal mice, as previously described.2 Prior to echocardiography, each adult mouse chest wall was shaved. Adult and neonatal mice were then placed on a heated platform in the supine position. Echocardiographic analysis was performed on mice anesthetized with oxygen mixed with isofluorane (2%) at a heart rate of >400 beats/minute. The only exception was LNA inhibitor echocardiographic studies, wherein mice received oxygen without isofluorane. Data acquisition was initiated with the parasternal cardiac long axis view followed by transition to a short axis view, at the level of mid-papillary muscles. Echocardiographic measurements were obtained from short axis B-mode images. All data were acquired and analyzed in a blinded fashion.

**Statistics**
Statistical analyses were performed using one-way ANOVA analysis and Student unpaired *t* test using GraphPad Prism. All values are expressed as mean ± SEM. *P*<0.05 was considered statistically significant.

**Supplemental References**
Online Figure I. Heart Rate of neonate and adult post myocardial infarction (MI).
Heart rate of neonatal and adult mouse 4 hours post-MI hearts compared to Sham controls measured by echocardiography. N=9 for Neonate Sham; N=7 for Neonate MI and Adult MI; N=6 for Adult Sham.
Online Figure II. In situ hybridization of U6 and Scrambled miRNA.
Postnatal Day 8 mouse ventricular cross sections were subjected to in situ hybridization using DIG-labeled U6 and Scrambled miRNA probes, and stained with anti-DIG (red) and anti-cardiac Troponin T (cTnT, green) antibodies as well as with wheat germ agglutinin (WGA, white) and Hoechst dye (blue). Representative images of sections at low magnification (Upper panels) followed by high magnification images (Lower panels) of regions indicated by yellow boxes.
Online Figure III. miR-34a overexpression increases TUNEL positive cells in neonatal post myocardial infarction (MI).
Ventricular cross sections of neonatal hearts border region at 7 days post-MI were stained with a TUNEL assay kit (red), and immunostained with anti-cardiac Troponin T antibody (cTnT, green) and Hoechst dye (blue). TUNEL positive cells are indicated by white arrows.
Online Figure IV. miR-34a inhibition in adult myocardial infarction (MI).
A, Time schedule of adult MI and LNA inhibitor injection. B, Ventricular cross sections of adult border region at 7 days post-MI were stained with a TUNEL assay (red) kit, and immunostained with anti-Troponin T antibody (cTnT, green) and Hoechst dye (blue). TUNEL positive cells are indicated by white arrows.
Online Figure V. Jag1 is not a target of miR-34a in NRVMs.
A, Luciferase assay in HEK-293 cells were co-transfected with luciferase constructs harboring 3′UTR sequences from either wild-type (WT) or mutated (Mut) Jag1 with or without miR-34a mimic for 48 hours. N=3. B to C, NRVMs either infected with adenovirus overexpressing LacZ or Jag1 or transfected with NC or miR-34a mimic for 48 hours were harvested for immunoblot analyses using anti-Jag1 and anti-β-Actin antibodies (B); results were quantified by densitometry (C). N=4 for all groups.
Online Figure VI. Inhibition of miR-34a targets in vivo.
A. Immunoblot analysis in neonatal 7 days post-MI hearts using anti-Bcl2, anti-Cyclin D1, anti-Sirt1 and anti-β-Actin antibodies. B. Densitometry quantification of results shown in (A). *P<0.05 vs. NC. N=7 for Bcl2 and Cyclin D1; N=3 for Sirt1.
Online Figure VII. Bcl2, Cyclin D1 and Sirt1 mediate miR-34a’s function.

A, 12 to 24 hours after transfection with NC or miR-34a mimic and transduction with Ad-LacZ, Ad-Bcl2, Ad-Cyclin D1, or Ad-Sirt1, neonatal rat ventricular myocytes (NRVMs) were treated with 50μM EdU for 48 hours. Cells were stained with an EdU assay (red) kit, and immunostained with anti-cardiac Troponin T antibody (cTnT, green) and Hoechst dye (blue). B, 12 to 24 hours after transfection with NC or miR-34a mimic, NRVMs were transduced with adenoviruses: Ad-LacZ, Ad-Bcl2, Ad-Cyclin D1, or Ad-Sirt1 for 48 hours. Cells were stained with a TUNEL assay (red) kit, and immunostained with anti-cardiac Troponin T antibody (cTnT, green) and Hoechst dye (blue).
Online Figure VIII. Full blots for Figure 7.
Online Figure VIII. Full blots for Figure 7.

7A, Membrane was cut between 50kD and 37kD. The upper half of membrane was incubated with anti-α-Tubulin antibody and imaged at 700nm. The lower half of membrane was incubated with anti-Bcl2 antibody and imaged at 800nm.

7B, Upper panel: WB using anti-Cyclin D1 antibody was imaged at 800nm. Bottom panel: WB using anti-α-Tubulin antibody and imaged at 700nm.

7C, Upper panel: WB using anti-Sirt1 antibody was imaged at 800nm. Bottom panel: WB using anti-α-Tubulin antibody and imaged at 700nm.

7D, WB using anti-Bcl2, anti-Cyclin D1 and anti-Sirt1 antibodies and were imaged at 800nm, 700nm and 800nm respectively. Membrane was striped, incubated with anti-β-Actin antibody and then imaged at 800nm.

7F, Membrane was cut at 37kD. The upper half of membrane was incubated with anti-β-Actin antibody and imaged at 800nm; the lower half of membrane was incubated with anti-Bcl2 antibody and imaged at 800nm.

7G, Membrane was cut between 50kD and 37kD. The upper half of membrane was incubated with anti-α-Tubulin antibody; the lower half of membrane was incubated with anti-Cyclin D1 antibody and imaged at 700nm.

7H, Left panel: WB using anti-Sirt1 antibody was imaged at 800nm. Right panel: Membrane was striped, incubated with anti-β-Actin antibody and then imaged at 800nm.