CD8⁺ T Cells Regulate Monopoiesis and Circulating Ly6C<sup>high</sup> Monocyte Levels in Atherosclerosis in Mice

Clément Cochain, Miriam Koch, Sweena M. Chaudhari, Martin Busch, Jaroslav Pelisek, Louis Boon, Alma Zernecke

Rationale: Proinflammatory adaptive immune responses are recognized as major drivers of atherosclerotic lesion formation. Although CD8⁺ T cells have recently been proposed as a proatherogenic cell subset, their full scope of actions remains to be elucidated.

Objective: We here addressed the contribution of CD8⁺ T cells to monocyte trafficking in atherosclerosis.

Method and Results: We observed that CD8⁺ T cells express proinflammatory cytokines (interferon-γ, tumor necrosis factor-α, and interleukin-12) within atherosclerotic lesions and spleens of high-fat diet–fed low-density lipoprotein receptor–deficient (Ldlr<sup>−/−</sup>) mice. Antibody-mediated CD8⁺ T-cell depletion in high-fat diet–fed Ldlr<sup>−/−</sup> mice decreased atherosclerotic plaque formation, associated with decreased macrophage accumulation within lesions. Despite a reduction in vascular chemokine (CC-motif) ligand 2 and chemokine (CXC-motif) ligand 1 expression, CD8⁺ T-cell depletion did not directly affect monocyte recruitment to inflamed vessels. However, CD8⁺ T-cell depletion decreased chemokine (CC-motif) ligand serum concentrations and circulating Ly6C<sup>high</sup> monocyte counts. We further evidenced that CD8⁺ T-cell depletion decreased levels of mature monocytes and myeloid granulocyte–monocyte progenitors in the bone marrow and spleen of hypercholesterolemic mice, effects that were partially reproduced by interferon-γ neutralization, showing a role for interferon-γ.

Conclusions: These data suggest that CD8⁺ T cells promote atherosclerosis by controlling monopoiesis and circulating monocyte levels, which ultimately contribute to plaque macrophage burden without affecting direct monocyte recruitment, identifying this cell subset as a critical regulator of proatherogenic innate immune cell responses in atherosclerosis. (Circ Res. 2015;117:244-253. DOI: 10.1161/CIRCRESAHA.117.304611.)

Key Words: atherosclerosis □ inflammation □ lymphocytes □ monocytes

Atherosclerosis is a chronic inflammatory disease of the vascular wall driven by the progressive intimal accumulation of leukocytes, consisting mainly of monocyte-derived macrophages that subsequently give rise to lipid-laden foam cells. However, it is now well recognized that also local and systemic adaptive immune responses play a critical role in atherogenesis. Since the first description of a localization of CD4⁺ and CD8⁺ T cells in human atherosclerotic plaques, pro- and antiatherogenic CD4⁺ T-helper cell subsets have been defined. In particular, CD4⁺ T-helper cells 1 (Th1) and their major cytokine interferon (IFN)-γ promote atherosclerosis, whereas Foxp3⁺CD25⁺ regulatory T cells inhibit vascular inflammation and atherogenesis. Although still under debate, Th2 cells and their signature cytokines (eg, interleukin [IL]-13) may predominantly be protective. The function of Th17 cells and their cytokine IL-17 is still unclear because contradicting reports have been published in past years. Surprisingly, however, the role of CD8⁺ T cells in atherogenesis is still largely unknown.

CD8⁺ T cells have been described to constitute a variable but substantial proportion of the cellular plaque infiltrate in human and murine atherosclerosis. Treatment of apolipoprotein E–deficient (Apoε<sup>−/−</sup>) mice with a CD137 agonist strongly increased vascular CD8⁺ T cell infiltration, associated with exacerbated atherosclerosis. In addition, we and others also demonstrated drastically elevated CD8⁺ T cell levels and atherosclerotic lesion formation in Ldlr<sup>−/−</sup>Pdl<sup>−/−</sup> mice, further suggesting a proatherogenic role of CD8⁺ T cells in atherogenesis. Similarly, CD8⁺ T cell activation by antigen-presenting cells toward an artificial antigen specifically expressed in the

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Nonstandard Abbreviations and Acronyms

<table>
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<th>Abbreviation</th>
<th>Description</th>
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<tr>
<td>CCL</td>
<td>chemokine (CC-motif) ligand</td>
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<tr>
<td>CMP</td>
<td>common myeloid progenitors</td>
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<tr>
<td>CXCL</td>
<td>chemokine (CXC-motif) ligand</td>
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<tr>
<td>DCs</td>
<td>dendritic cells</td>
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<tr>
<td>GM-CSF</td>
<td>granulocyte-macrophage colony-stimulating factor</td>
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<tr>
<td>GMP</td>
<td>granulocyte–monocyte progenitor</td>
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<tr>
<td>HFD</td>
<td>high-fat diet</td>
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<tr>
<td>IFN</td>
<td>interferon</td>
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<tr>
<td>IL</td>
<td>interleukin</td>
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<tr>
<td>Ldfr</td>
<td>low-density lipoprotein receptor</td>
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<tr>
<td>MP</td>
<td>myeloid progenitor</td>
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<td>Th</td>
<td>T helper cell</td>
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vessel wall induced arteritis and increased atherosclerosis, possibly via classical cell toxicity mechanism directed toward vessel wall cells, CD8+ T cells could, in addition, potentiate systemic and vascular inflammation through the secretion of proatherogenic cytokines. For instance, CD8+ T cells were shown to highly secrete IFN-γ in aortic root-draining lymph nodes of hypercholesterolemic mice. Moreover, Kyaw et al recently proposed that CD8+ T cells were able to promote atherogenesis and the development of vulnerable plaques through cytotoxic and proinflammatory effects, mediated by granzyme B, perforin, and tumor necrosis factor (TNF-α) expression. It should be noted, however, that several studies have proposed a nonessential role of CD8+ T cells in atherosclerosis. In Apoe−/− mice, deficiency in CD8 did not significantly affect atherosclerosis, whereas total TCRβ+ T cell or CD4+ T-cell deficiency decreased or increased atherosclerosis, respectively. CD4 deficiency, besides abrogating atheroprotective regulatory T-cell responses, was speculated to lead to defects and proinflammatory properties of CD8+ T cells. Hypercholesterolemic mice deficient in transporter associated with antigen processing (TAP1−/−) mice did not significantly affect atherosclerosis, whereas total TCRβ+ T cell or CD4+ T-cell deficiency decreased or increased atherosclerosis, respectively. These effects, however, may have been masked by a compensatory increase in CD4+ T-cell infiltrates in atherosclerotic lesions of Apoe−/− TAP1−/− mice, leading to total lesional CD3+ T-cell contents comparable with controls. Furthermore, a recent study proposed that a small subset of regulatory CD8+ T cells inhibited proatherogenic activation of follicular helper T cells (Th), thereby limiting lesion development.

Recently, CD8+ T cells were shown to indirectly promote monocyte production in the bone marrow through IFN-γ-mediated activation of bone marrow stromal cells during viral infection. Considering that CD8+ T-cell depletion leads to a reduced macrophage accumulation in atherosclerotic lesions, we here re-evaluated the function of CD8+ T cells during atherogenesis. Using a model of antibody-mediated depletion of CD8+ T cells in atherosclerotic low-density lipoprotein receptor–deficient (Ldfr−/−) mice, we here show that CD8+ T cells control Ly6C<sup>high</sup> (Ly6C<sup>hi</sup>) monocyte levels in circulating blood, spleen, and bone marrow, ultimately leading to reduced macrophage accumulation in plaques and decreased lesion formation without directly affecting monocyte recruitment. Furthermore, we show that CD8+ T cells promote medullar monopoiesis during atherogenesis, uncovering a previously unrecognized mechanism of CD8+ T-cell–driven atherogenesis.

Methods

Animals

Ldlr<sup>−/−</sup> mice (C57BL/6J background, obtained from the Jackson Laboratory) were bred and housed under specific pathogen free conditions. Eight- to 10-week-old male mice were placed on an atherogenic diet (15% milk fat, 1.25% cholesterol; Altromin) for 6 weeks, and treated with weekly intraperitoneal injections of 250 μg of rat antimouse-CD8<sub>α</sub> (clone YTS-169, referred to as anti-CD8<sub>α</sub> or a-CD8<sub>α</sub>), rat antimouse-CD8<sub>β</sub> (clone YTS-156.7.7, referred to as anti-CD8<sub>β</sub> or a-CD8<sub>β</sub>), or isotype-matched irrelevant rat-anti-Phy1 (clone APRC-MAC51, referred to as control). To analyze monocyte recruitment to the vascular wall, mice fed an atherogenic diet and treated with control or anti-CD8<sub>α</sub> for 5 weeks received intravenous injections of 10×10<sup>6</sup> bone marrow mononuclear cells (prepared by gradient density centrifugation on Histopaque 1083) from mice expressing the CD45.1 haplotype. Forty-eight hours after cell transfer, aortae and aortic roots were collected and prepared for flow cytometry. For in vivo IFN-γ-neutralization, mice received intraperitoneal injections of 250 μg of rat antimouse IFN-γ antibody (clone XMG1.2) or irrelevant isotype-matched rat anti–β-galactosidase antibody (clone GL113) every 48 hours for 3 weeks. All animal experiments were approved by local authorities (Regierung von Unterfranken, Würzburg, Germany) to comply with German animal protection law.

Immunohistochemistry on Human Tissues

Immunohistochemistry was performed on representative sections of human carotid artery tissue samples from patients with high-grade carotid artery stenosis (>70%) who underwent carotid endarterectomy in the Department of Vascular and Endovascular Surgery (Klinikum Rechts der Isar, Technical University Munich). The carotid plaque was fixed in formalin and embedded in paraffin. For analysis of CD8 expression, tissue sections were incubated overnight at 4°C with monoclonal mouse antihuman CD8 antibody visualized using peroxidase/DAB.

Enzymatic Tissue Digestion and Flow Cytometry

Blood counts were analyzed using an automated hematology analyser (Sysmex). For fluorescence activated cell sorting analyses, single-cell suspensions from blood, whole aortae, and aortic root after enzymatic dissociation using Liberase Blendzyme TL solution (Roche) or bone marrow were resuspended in HBSS, and cells were stained for 30 minutes at 4°C using specific antibodies from Becton Dickinson Biosciences or eBioscience. Intracellular labeling of TNF-α, IL-12, IL-17A, and IFN-γ was performed after PMA (phorbol-12-myristat-13-acetate)/ionomycine/brefeldin stimulation for 4 hours using the Becton Dickinson Cytofix/Cytoperm Kit (Becton Dickinson Biosciences). Bone marrow cells were incubated for 4 hours with brefeldin before intracellular staining for granulocyte-macrophage colony-stimulating factor (GM-CSF) and IL-6. Probes were analyzed using a FACSCanto II (Becton Dickinson) and FlowJo 7.6 software (Treestar Inc).

Analysis of Atherosclerotic Lesion Development

Arteries were perfusion-fixed in situ with phosphate buffered saline (PBS) followed by 4% buffered paraformaldehyde in PBS. The aortic root was cut in 5-μm transverse cryosections. Aortic root sections and the en face prepared aortae were assessed for atherosclerotic plaque size after staining with Oil-Red-O. Plaque size was quantified by computerized image analysis (Diskus Software).

Quantitative Real-Time Polymerase Chain Reaction

Total RNA was extracted from tissues using a Nucleo Spin RNAII extraction Kit (Macherey-Nagel, Düren, Germany) in accordance with the manufacturer’s instructions. Equal amounts of template
RNA were used for cDNA synthesis; RNA was reverse transcribed using random hexamer primers. Quantitative SYBR-green real-time polymerase chain reaction was performed on duplicate samples of template cDNA on an Applied Biosystems 7900HT fast real-time polymerase chain reaction system using specific primer pairs. Quantitative measurements were determined using the ΔΔCt method, with Hprt as the housekeeping gene.

**In Vitro Coculture Assays**

Bone marrow–derived macrophages were obtained by culturing total bone marrow cells from Ldlr−/− mice in Roswell Park Memorial Institute (RPMI) medium containing 10% heat-inactivated FCS, antibiotics, and 15% L929 cell-conditioned medium as a source of macrophage colony-stimulating factor (M-CSF). CD8+ T cells were isolated from the spleen of Ldlr−/− mice using antimouse CD8α microbeads and magnetic cell sorting according to the manufacturer’s instructions (Miltenyi). Bone marrow–derived macrophages or SV40-transformed mouse endothelial cells (SVEC) were serum starved and cultured alone or with isolated CD8+ T cells at a 1:1 ratio in RPMI containing 0.5% FCS, 5 μg/mL anti-CD3, and 1 μg anti-CD28 antibody, with the addition of 10 μg/mL anti–IFN-γ-neutralizing antibody or 10 μg/mL of isotype control antibody. After 24 hours of incubation, conditioned medium was collected and chemokine expression assessed by ELISA.

**ELISA**

Chemokine (CC-motif) ligand 2 (CCL2), chemokine (CXC-motif) ligand 1 (CXCL1), and CCL5 levels in serum and cell culture supernatants were measured using the corresponding ELISA Development Kits according to the manufacturer’s instructions (Peprotech).

**Statistical Analysis**

Results are expressed as mean±SEM and were analyzed by Student t test or 1-way ANOVA followed by Bonferroni post-testing for multiple comparisons using Prism GraphPad 6.0 software. Differences with P<0.05 were considered to be statistically significant.

**Results**

**CD8+ T Cells Are Present in Human and Murine Atherosclerotic Plaques**

Given findings of CD8+ T-cell infiltrates within atherosclerotic plaques,8 we first assessed the presence of this cell population in human and murine atherosclerotic vessels. Consistent with previous reports, immunostaining of human carotid artery atherosclerotic plaque tissue revealed abundant CD8+ T-cell infiltrates at various locations within lesions, including the plaque shoulder (Figure 1A) and around necrotic cores (Figure 1B). CD8+ T-cell levels were also detected by fluorescence activated cell sorting analysis in the enzymatically digested aortic root (Figure 1C) and aorta (Online Figure I) in Ldlr−/− mice after 6 weeks of high-fat diet (HFD) feeding, and CD8+ T-cell levels were increased at these sites when compared with chow-fed mice without alterations within the distribution of CD4+ versus CD8+ T-cell subsets among total CD3+TCRβ+ T cells (Online Figure IA–IC). Analysis of the cytokine production profile of aortic root CD8+ T cells in atherosclerotic mice...
revealed large proportions of CD8+ T cells to produce TNF-α (43.0% of CD8+ T cells) and IFN-γ (17.9%), whereas only few cells expressed IL-12 (1.0% of CD8+) and no staining could be detected for IL-17A (Figure 1D). Systemically, splenic CD8+ T cells expressed TNF-α (82.1% of CD8+ T cells), IFN-γ (22.2%), and IL-12 (19.8%; Figure 1E). These data show that CD8+ T cells infiltrate human and murine lesions, and in addition produce proinflammatory cytokines both in inflamed vessels and systemically in atherosclerotic mice.

**CD8+ T-Cell Depletion Reduces Atherosclerosis and Plaque Macrophage Accumulation**

To evaluate the functional role of CD8+ T cells in atherosclerosis, this cell subset was continuously depleted in Ldlr−/− mice. Repeated injections with a monoclonal-depleting anti-CD8α antibody reduced CD8+ T-cell infiltrates in the aortic root and aorta (Figure 2A and 2B; Online Figure ID and IE) and systemically in circulating blood and spleen without affecting CD4+ T-cell numbers (Figure 2A and 2B; Online Figure II) or their activation and polarization toward Th1 or regulatory T cell lineages (Online Figure III) compared with mice receiving a control antibody. As a subset of dendritic cells (DCs) in lymphoid organs expresses CD8α, we furthermore analyzed the DC compartment in treated mice. Frequencies of CD8α+ and CD11b+ DCs in the spleen, as well as total numbers of CD11c+MHCI+ DCs in atherosclerotic vessels, were not affected in anti–CD8α-treated mice (Online Figure IVA–IVC). In addition, isolated splenic CD11c+ DCs from anti–CD8α-treated mice loaded with OVA323-339 peptide did not show any alterations in their capacity to trigger antigen-dependent proliferation of naïve OTII CD4+ T cells, or regulatory T cell and Th1 polarization compared with control-treated mice (Online Figure IVD–IVF), further confirming the specificity of our CD8+ T-cell depletion strategy.

CD8+ T-cell depletion entailed a significant 33% reduction in atherosclerotic lesion size in the aortic root (Figure 2C and 2D) and a 15% decrease in lesion formation in the aortic arch (Figure 2E) compared with mice injected with control...
antibody after 6 weeks of HFD feeding. No differences in serum cholesterol (control antibody, 7728±521 µg/mL; anti-CD8, 7214±31 µg/mL) or triglyceride levels (control antibody, 5.5±0.3 mmol/L; α-CD8, 5.5±0.2 mmol/L) were observed between groups. Histological analyses of the plaque furthermore revealed an unaltered plaque collagen content, but a 2.5-fold reduction in the relative necrotic core area and a 1.2-fold reduction in macrophage content (Figure 2I–2H).

CD8⁺ T cells express IFN-γ, TNF-α, and IL-12, and we furthermore assessed local mRNA expression of these cytokines. However, only Ifng but not Tnfa or Il12 transcripts were reduced in the aortic root of anti–CD8α-treated Ldlr⁻/⁻ mice (Figure 2J–2L).

As lesion-infiltrating CD8⁺ T cells express IFN-γ, TNF-α, and IL-12, we furthermore analyzed the effects of CD8⁺ T cells on macrophage polarization. In vitro, coculture of bone marrow–derived macrophages with activated CD8⁺ T cells promoted M1 polarization, as reflected by increased inducible nitric oxide synthase (Nos2) expression, an effect that was largely but not completely prevented in the presence of a neutralizing anti–IFN-γ antibody (Online Figure V). In vivo, however, mRNA expression of Nos2 and of the M2 markers Peroxisome proliferator-activated receptor gamma (Pparγ), Arginase-1 (Arg1), and mannose receptor, C type 1 (Mrc1), and Pparg and of the M2 markers Peroxisome prolif-...
test this, Ldlr<sup>−/−</sup> mice (which express the CD45.2 haplotype) treated with control or anti-CD8<sub>α</sub> antibody for 5 weeks received intravenous injections of mononuclear cells extracted from the bone marrow of mice carrying the CD45.1 haplotype. After 48 hours, flow cytometry of atherosclerotic vessels revealed similar levels of adoptively transferred CD45.1<sup>+</sup>CD45.2<sup>−</sup>Ly6G<sup>−</sup> monocytes in anti–CD8<sub>α</sub>- and control-treated mice (Figure 4A and 4B), indicating that CD8<sup>+</sup> T-cell depletion did not directly affect monocyte recruitment from the blood to the inflamed vessel wall.

Macrophage influx into atherosclerotic lesions is also highly dependent on blood monocyte counts. Circulating levels of inflammatory Ly6<sub>C</sub>hi monocytes were decreased in anti–CD8<sub>α</sub>-treated Ldlr<sup>−/−</sup> mice at 6 weeks of HFD feeding, whereas numbers of Ly6C<sub>low</sub> (Ly6C<sup>+</sup>) monocytes remained unaffected (Figure 5A–5C). Chow-fed Ldlr<sup>−/−</sup> mice submitted to the same regimen of CD8<sup>+</sup> T-cell depletion did not show any alterations in blood monocyte counts after 6 weeks, or changes in CCL2 serum levels (Online Figure VII), showing that CD8<sup>+</sup> T cells contribute to monocytosis under conditions of hypercholesterolemia.

In inflammatory settings, monocytes can be mobilized to the blood from different compartments, namely the spleen and the bone marrow. Therefore, we analyzed levels of mature Ly6C<sup>+</sup> monocytes in these organs. Anti-CD8<sub>α</sub> treatment reduced levels of Ly6C<sup>+</sup> monocytes in the bone marrow (Figure 5D–5F) and spleen (Figure 5G) of Ldlr<sup>−/−</sup> mice at 6 weeks of HFD feeding, demonstrating that CD8<sup>+</sup> T cells control the size of the medullar and splenic monocyte pool in hypercholesterolemia.

**CD8<sup>+</sup> T-Cell Depletion Affects Monopoiesis During Hypercholesterolemia**

To investigate whether CD8<sup>+</sup> T cells may control monopoiesis, we assessed levels of total Lin<sup>−</sup>ckit<sup>−</sup>sca1<sup>+</sup> hematopoietic stem cells, total Lin<sup>−</sup>ckit<sup>−</sup>sca1<sup>−</sup> myeloid progenitors (MPs), Lin<sup>−</sup>ckit<sup>−</sup>sca1<sup>−</sup>CD34<sup>+</sup>CD16<sup>−</sup>/32<sub>high</sub> common MPs (CMPs), and Lin<sup>−</sup>ckit<sup>−</sup>sca1<sup>−</sup>CD34<sup>+</sup>CD16<sup>−</sup>/32<sub>low</sub> granulocyte-MPs (GMPs) in the bone marrow of anti-CD8<sub>α</sub> and control antibody-treated mice after 6 weeks of HFD feeding. Levels of total hematopoietic stem cells and MPs were unaltered, both in frequencies among total bone marrow cells and in absolute cell counts (Figure 6A–6D). In contrast, frequencies of GMPs among MPs (Figure 6E–6G) and absolute cell counts of GMPs per femur (Figure 6H and 6I) were strongly reduced in the bone marrow of anti–CD8<sub>α</sub>-treated mice, whereas CMP numbers were unaltered but showed a relative increase in their frequencies (Figure 6E–6I). Similarly, a significant reduction in GMP cell counts was obtained in the spleen of these mice (Online Figure VIII). These data suggest that CD8<sup>+</sup> T cells promote monocyte expansion by promoting the differentiation of common myeloid to GMPs or through promotion of GMP proliferation.

As GMPs also give rise to neutrophils, we furthermore analyzed their distributions in different organs. Neutrophil counts were significantly diminished in the bone marrow, but not in the blood, spleen, or in atherosclerotic vessels (Online Figure IX).

Absolute cell counts of MPs, CMPs, and GMPs were unaffected in bone marrow of Ldlr<sup>−/−</sup> mice treated with anti-CD8<sub>α</sub> for 6 weeks and fed a normal chow although a minute 3.9% increase in CMP frequencies reached significance (Online Figure X). Likewise, MP, CMP, and GMP levels were not affected in the spleen of chow-fed mice (Online Figure XI). These findings show that CD8<sup>+</sup> T cells promote monopoiesis specifically under conditions of hypercholesterolemia. In line, anti-CD8<sub>α</sub> treatment did not affect Ly6C<sup>+</sup> monocyte levels in the bone marrow of Ldlr<sup>−/−</sup> mice fed a normal chow (Online Figure XIIA). However, low counts of inflammatory monocytes in spleen were further diminished by anti-CD8<sub>α</sub> treatment in Ldlr<sup>−/−</sup> mice fed a normal chow (Online Figure XIIIB), suggesting that CD8<sup>+</sup> T cells control monocyte accumulation in the spleen both under normal and hypercholesterolemic conditions by mechanisms independent of direct effects on monopoiesis.

**CD8<sup>+</sup> T-Cell Depletion Using Anti-CD8β Antibodies Reduces Ly6C<sup>+</sup> Monocyte Levels and Monopoiesis**

Even though our data indicate that CD8<sup>+</sup> T-cell depletion using anti-CD8<sub>α</sub> antibodies had no effect on the DC compartment or on CD4<sup>+</sup> T-cell levels or activation, we wanted to further validate our findings using an alternative monoclonal...
antibody targeting CD8β. After 6 weeks of HFD, Ldlr−/− mice treated weekly with 250 μg of anti-CD8β antibody showed an efficient depletion of CD8+ T cells in blood and spleen (Online Figure XIIIA and XIIIB). Atherosclerotic lesion development was reduced by 18.4% in the aortic arch of anti-CD8β compared to control-treated treated mice although it remained borderline significant (P=0.06; Online Figure XIIIC). In the aortic root, lesion size also tended to be reduced although this was not statistically significant (P=0.16; Online Figure XIIID). Plaque macrophage content was significantly reduced.

Figure 5. CD8+ T cells control systemic monocyte homeostasis. A, Representative fluorescence activated cell sorting (FACS) plots of CD115 vs Ly6C expression on total blood cells; absolute counts of (B) Ly6C+ and (C) Ly6C− monocytes in circulating blood in Ldlr−/− mice treated or not with a-CD8α after 6 weeks of high-fat diet (HFD). D, Representative FACS plots of CD115 vs Ly6C expression in total bone marrow cells allow identification of CD115+Ly6C− monocytes in control and a-CD8α–treated Ldlr−/− mice after 6 weeks of HFD. Quantification of (E) percentages of CD115+Ly6C− monocytes in total bone marrow cells and (F) absolute counts of CD115+Ly6C− monocytes in bone marrow, and (G) Ly6C+ monocytes in the spleen from control and a-CD8α–treated Ldlr−/− mice after 6 weeks of HFD (n=5–8 per group). *P<0.05, ***P<0.001.

Figure 6. a-CD8α treatment reduces medullar granulocyte–monocyte progenitor (GMP) levels in high-fat diet (HFD) fed Ldlr−/− mice. Levels of hematopoietic stem cell (HSC) in (A) percentage of total cells and (B) absolute cell numbers; levels of myeloid progenitors (MP) in (C) percentage of total cells and (D) absolute cell numbers; (E) representative fluorescence activated cell sorting (FACS) plots of CD34 vs CD16/32 expression on gated Lin−ckit+sca1− MP allows identification of CD34+CD16/32− GMP and CD34+CD16/32+ common myeloid progenitors (CMP). Proportions of (F) CMP and (G) GMP among gated MP; absolute cell counts of (H) CMP and (I) GMP in the bone marrow from control and a-CD8α–treated Ldlr−/− mice after 6 weeks of HFD (n=7–8 per group).
in anti–CD8β-treated mice (Online Figure XIIIIE), indicating a reduced lesional macrophage accumulation.

Anti–CD8β-treated mice displayed reduced blood Ly6C+ monocyte counts, whereas Ly6C+ monocyte and neutrophil numbers were similar to controls (Online Figure XIIVA–XIVC). Bone marrow and spleen Ly6C+ monocyte levels were also significantly reduced (Online Figure XIVD and XIVF). Moreover, a reduction in the number of GMPs but no alterations in CMPs was noted in bone marrow of anti–CD8β-treated mice (Online Figure XIVE and XIVF). These data further corroborate our findings that during atherogenesis, CD8+ T cells control monopoiesis, circulating and splenic monocyte levels, and ultimately macrophage accumulation within plaques.

**CD8+ T-Cell Depletion Reduces GM-CSF and IL-6 Expression in Bone Marrow Cells**

It was recently proposed that CD8+ T cells promote monopoiesis during acute viral infection through induction of IL-6 expression by bone marrow mesenchymal stromal cells. GM-CSF has also been proposed as a regulator of the myeloid lineage during inflammation. In Ldlr−/− mice fed a HFD for 6 weeks, expression of GM-CSF (Online Figure XVA and XV); and IL-6 (Online Figure XV and XVE) was significantly reduced in total bone marrow cells by 28.3% and 16.5%, respectively, in anti–CD8β-compared to control-treated mice. Expression of both cytokines was low in CD8+ T cells, representing 0.6±0.1% and 0.8±0.2% of GM-CSF and IL-6 expressing cells, respectively. Although most GM-CSF and IL-6 expression was found in CD45+ cells, we in addition detected a significant 41.8% reduction in IL-6 expression in CD45− cells; low frequencies of GM-CSF+ cells among CD45− cells were comparable between groups (Online Figure XVC and XVF).

**IFN Neutralization Partially Recapitulates the Effects of CD8+ T-Cell Depletion**

IFN-γ has been proposed as the main mediator of CD8+ T-cell–induced monopoiesis during viral infection. CD8+ T cells were the major IFN-γ-expressing cell population in the bone marrow, and anti-CD8αtreatment significantly reduced frequencies of total medullar IFN-γ+ cells (Online Figure XVI). Notably, short-term treatment of HFD-fed Ldlr−/− mice with an IFN-γ-neutralizing antibody for 3 weeks induced a similar shift in medullar MP subpopulations and increased CMP but decreased GMP frequencies (Online Figure XVIIA–XVIIIC), accompanied by a trend towards decreased Ly6C+ and reduced Ly6C+ monocytes in blood (Online Figure XVIIID and XVIIIE). In contrast, however, IFN-γ neutralization was associated with a significant expansion of mature monocytes in the bone marrow (Online Figure XVIIIF). Although IFN-γ may thus, in part, be responsible for effects of CD8+ T cells on bone marrow monopoiesis during hypercholesterolemia, these findings also suggest that short-term IFN-γ blockade interferes with monocyte mobilization.

**Discussion**

In the present study, we demonstrate a novel mechanism by which CD8+ T cells contribute to atherogenesis through modulation of medullar monopoiesis and circulating Ly6C+ monocyte levels, thereby indirectly controlling macrophage accumulation within lesions.

Previous studies addressing the role of CD8+ T cells in atherogenesis have mostly used genetically engineered mouse models of CD8+ T-cell deficiency with contradictory results, which may be because of compensatory mechanisms in these mice. We circumvented this hurdle by treating Ldlr−/− mice with an anti-CD8α monoclonal antibody, which efficiently depleted CD8+ T cells while not altering DCs levels, including CD8α+ DCs, and functionality of splenic CD11c+ DCs, as well as leaving CD4+ T cell numbers, activation and polarization untouched, thus confirming the specificity of our depletion strategy.

CD8+ T-cell depletion with the anti-CD8α antibody entailed a significant decrease in atherosclerotic lesion formation in the aortic root and the aortic arch, in line with a recent study using a similar approach in Apoe−/−mice, corroborating a proatherogenic role of CD8+ T cells. Lesions of anti–CD8α-treated mice had smaller necrotic cores, which is consistent with cytotoxic properties of CD8+ T cells in plaques that may depend on perforin and granzyme B expression. Also coherent with previous findings, this reduction in lesion formation was associated with a reduction in lesional macrophage accumulation. Furthermore, we here observed diminished expression of CCL2 and CXCL1 in atherosclerotic tissue, chemokines that are known to attract monocytes. Reduced chemokine expression in plaque tissue may reflect changes in lesion cellular composition (ie, reduced macrophage content) or reduced chemokine expression by individual plaque cells. In vitro, activated CD8+ T cells promoted expression of CCL2, CXCL1, and CCL5 in macrophages and endothelial cells, and IFN-γ-dependent induced CCL2 in macrophages, suggesting that CD8+ T cells may also promote chemokine expression by neighboring cells within plaques. These results also raised the possibility that CD8+ T cell may indirectly promote monocyte recruitment to the plaque. The recruitment of adoptively transferred monocytes, however, was unaffected in anti–CD8α-treated atherosclerotic mice, ruling out this hypothesis.

Circulating Ly6C+ monocytes are known to give rise to macrophages in atherosclerotic lesions, and their circulating levels dictate the extent of macrophage accumulation within plaques. Anti–CD8αtreated mice displayed reduced circulating Ly6C+ monocyte counts. Considering that direct monocyte recruitment to plaques was unaffected by CD8+ T-cell depletion, this suggests that reduced circulating Ly6C+ monocyte levels in CD8+ T-cell depleted mice caused the reduction in macrophage accumulation within plaques. A recent study suggested that lesional proliferation of macrophages is responsible for their accumulation during late phases of plaque development, and one could speculate that CD8+ T cells may interfere with macrophage proliferation at this stage. Our model of atherogenesis after 6 weeks of HFD feeding, however, is rather representative of the early steps of atherogenesis, when monocyte recruitment prevails.

Significantly decreased serum CCL2 concentrations in anti–CD8αtreated mice may indicate a possible impairment
of CCL2-mediated monocyte mobilization from the bone marrow or spleen. Contrary to this notion, however, levels of mature monocytes were similarly reduced in bone marrow and spleen of anti–CD8α-treated hypercholesterolemic mice, suggesting that CD8+ T cells control myelopoiesis. Spleen but not bone marrow monocyte levels were also reduced in normocholesterolemic mice, suggesting that CD8+ T cells control the size of the splenic monocyte pool also under homeostatic conditions; the mechanisms underlying this phenomenon require further investigations.

Recent evidence indicates that hypercholesterolemia primes hematopoietic stem and progenitor cells toward myelopoiesis, including monopoiesis, and that CD8+ T cells can promote emergency monopoiesis during viral infection. We here evidenced that CD8+ T-cell depletion led to decreased levels of GMPs, which represent a late stage of myeloid cell differentiation, both in the bone marrow and the spleen, an effect that occurred specifically in hypercholesterolemic mice. Consistent with lower GMP levels, bone marrow neutrophil levels were also decreased, but their counts at other sites (blood, spleen, and atherosclerotic vessels) were unchanged, suggesting that the size of the medullar neutrophil pool remained sufficient to provide for unaltered neutrophil counts at peripheral sites.

Our data thus suggest that CD8+ T-cell depletion limits the medullar production of mature monocytes, which are then mobilized to the bloodstream at lower numbers. Reduced CCL2 blood levels may contribute to lower circulating Ly6C+ monocyte counts in CD8+ T-cell depleted mice. Reduced blood Ly6C+ monocyte numbers then, in turn, lead to diminished macrophage accumulation within plaques. Corroborating these findings, similar evidence was obtained using a different depleting antibody targeting CD8β.

In a recent study, Schürch et al proposed that CD8+ T cells promote medullar monopoiesis through an IFN-γ–dependent induction of IL-6 in bone marrow mesenchymal stromal cells in acute viral infection. In our study, CD8+ T-cell–depleted mice displayed reduced IL-6 expression in bone marrow cells, including CD45+ bone marrow cells. CD8+ T cells themselves displayed minimal IL-6 expression in bone marrow, indicating that CD8+ T cells control IL-6 expression in other bone marrow cells and in turn may promote monopoiesis also in hyperlipidemia-associated chronic inflammation. Short-term IFN-γ neutralization in atherosclerotic mice reduced medullar GMP and circulating monocyte levels, similar to findings in CD8+ T-cell–depleted mice. In contrast, it also entailed an increased accumulation of monocytes in the bone marrow. Although IFN-γ secretion by CD8+ T cells may, thus in part, contribute to monopoiesis and Ly6C+ monocytosis during athrogenesis, this suggests that other mechanisms are involved that remain to be elucidated. Of note, Kyaw et al proposed that proatherogenic effects of CD8+ T cells were IFN-γ independent. Expression of GM-CSF was also reduced in bone marrow cells in CD8+ T-cell–depleted mice, which may further contribute to the observed phenotype.

Recently, Clement et al showed that a small subset of regulatory CD8+ T cells limited atherosclerosis through inhibition of Th1 cells. Our results and those of Kyaw et al demonstrate that depletion of total CD8+ T cells leads to atheroprotective effects, indicating that when considering the CD8+ T-cell compartment in its entirety, proatherogenic effects prevail.

In conclusion, we here show that CD8+ T cells promote the expansion of the monocyte compartment in bone marrow, spleen, and blood and lead to an increased accumulation of macrophages in the arterial intima promoting atherosclerotic lesion formation. Our work identifies CD8+ T cells as a potential therapeutic target for immune modulation of atherosclerosis.

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Disclosures

None.

References

Novelty and Significance

**What Is Known?**

- Macrophage accumulation within atherosclerotic lesions is dependent on blood monocyte counts and determines atherosclerotic lesion growth.
- CD8+ T cells are present in atherosclerotic lesions in humans and mice and can promote atherosclerosis by cytotoxic and proinflammatory mechanisms.

**What New Information Does This Article Contribute?**

- We revealed that CD8+ T cells control monopoiesis in bone marrow and spleen and systemic levels of inflammatory Ly6C(++) monocytes in hypercholesterolemic mice.
- By controlling circulating monocyte counts, CD8+ T cells promote lesional macrophage accumulation and atherosclerosis lesion growth.

Atherosclerotic lesion development is driven by the progressive intimal accumulation of monocyte-derived macrophages, which is likely to be highly dependent on blood monocyte counts.
CD8⁺ T Cells Regulate Monopoiesis and Circulating Ly6C<sup>high</sup> Monocyte Levels in Atherosclerosis in Mice

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Material and methods

Animals

Ldlr-/- mice (C57BL/6J background, obtained from the Jackson Laboratory) were bred and housed in specific pathogen free conditions. 8 to 10 week old males were put on an atherogenic diet (15% milk fat, 1.25% cholesterol, Altromin, Lage, Germany) for 6 weeks, and treated with weekly intraperitoneal (i.p.) injections of 250µg of 250 µg of rat anti-mouse-CD8α (clone YTS-169, referred to as anti-CD8α or a-CD8α), rat anti-mouse-CD8β (clone YTS-156.7.7, referred to as anti-CD8β or a-CD8β) or isotype-matched irrelevant rat anti-Phyt1 (clone AFRC-MAC51, referred to as Ctrl). At the end of the treatment, mouse anti-rat IgG antibodies could be detected at similar levels in the serum of mice treated with anti-CD8α, anti-CD8β or control antibody (not shown), indicating that anti-rat antibody production does not constitute an invalidating bias for this experimental design. For analysis of monocyte recruitment to the vascular wall, mice fed an atherogenic diet and treated with Ctrl or anti-CD8α for 5 weeks received intravenous (i.v.) injections of 10x10⁶ bone marrow mononuclear cells (prepared by gradient density centrifugation on Histopaque 1083) from mice expressing the CD45.1 haplotype. 48 hours after cell transfer, aortae and aortic roots were collected together and prepared for flow cytometry. For in vivo IFNγ neutralization, mice received i.p. injections of 250µg of rat anti-mouse IFNγ antibody (clone XMG1.2) or irrelevant isotype matched rat anti-β-Galactosidase (clone GL113) every 48 hours for 3 weeks. All animal experiments were approved by local authorities (Regierung von Unterfranken, Würzburg, Germany) to comply with German animal protection law.
**Immunohistochemistry on human tissues**

Immunohistochemistry was performed on representative sections of human carotid artery tissue samples from patients with high-grade carotid artery stenosis (> 70%) that underwent carotid endarterectomy (CEA) in the Department of Vascular and Endovascular Surgery (Klinikum rechts der Isar, Technical University Munich). The carotid plaque was fixed in formalin and embedded in paraffin. For analysis of CD8 expression, tissue sections were incubated overnight at 4 °C with monoclonal mouse anti-human CD8 antibody (1/1000, Clone C8144B, DAKO) and visualized using peroxidase/DAB ChemMate Detection Kit (Dako) according to the manufacturer’s instructions. Images were recorded using a Leica DM 4000B microscope and JVC KY-F75U camera.

**Organ collection, enzymatic tissue digestion and flow cytometry**

Blood counts were analysed using an automated hematology analyser (KX-21N, Sysmex, Germany). For FACS analyses, tissues were passed through a 70 µm filter (BD Biosciences, Germany) to obtain single-cell suspensions. Whole blood was combined with a red blood cell lysis buffer (155 mM NH₄Cl, 10 mM KHCO₃, 0.1 mM EDTA) to allow the isolation of leukocytes. After flushing mice with 5-10 ml of phosphate buffer saline, organ collection was performed under a stereomicroscope. The aorta was carefully separated from surrounding adipose and connective tissue. The plaque-containing aortic root was visualized at the origin of the aortic arch on top of the left ventricle as a tissue with a characteristic creamy-white color and excised with minimal surrounding myocardial tissue. Aortas and aortic roots were enzymatically dissociated using Liberase Blendzyme TL (Roche, Mannheim, Germany) solution for 30 minutes at 37°C. Bone marrow was flushed from one femur with Hanks Buffered Saline Solution (HBSS). The resulting single cell suspensions were resuspended in HBSS and cells were stained for 30 minutes at 4°C. Antibodies used for FACS were purchased from eBioscience (Frankfurt a. Main, Germany) or BD (Heidelberg, Germany). The
following antibodies were used: CD45 (clone 30-F11), CD45.1 (clone A20), CD45.2 (clone 104), CD3 (clone 500A2 or 17A2), TCRβ (H57-597), CD8a (clone 53-6.7), Ly6C (clone AL21), Ly6G (Clone 1A8), NK1.1 (PK136), IFNγ (clone XMG1.2), CD4 (clone RM4-5 or GK1.5), CD11b (clone M1/70), CD11c (clone N418), IL-17a (clone eBio17B7), F4/80 (BM-8), CD25 (PC61.5), Foxp3 (FJK-16S), CD44 (IM7), CD62L (MEL-14), TNFα (clone MP6-XT22), IL12 (clone C15.6), CD16/32 (clone 93), c-kit (clone 2B8), Sca1 (clone D7), CD34 (clone RAM34), CD90 (clone 53-2.1), B220 (clone RA3-6B2), CD49b (clone DX5), Ter119 (clone Ter119), Gr1 (clone RB6-8C5), IL7Rα (clone A7R34), IL-6 (clone MP5-20F3), GM-CSF (clone MP1-22E9). For analysis of bone marrow progenitor populations, lineage antibody cocktail contained antibodies against CD90, B220, CD19, CD49b, Ter119, Gr1, NK1.1 and IL7Rα. Intracellular labelling of TNFα, IL12, IL17A and IFNγ was performed after PMA/Ionomycine/Brefeldin stimulation for 4 hours using the BD Cytofix/Cytoperm™ Kit (BD Biosciences). Bone marrow cells were incubated for 4 hours with Brefeldin before GM-CSF and IL-6 intracellular stainings. Probes were analyzed using a FACSCanto II (Becton Dickson, USA) and FlowJo 7.6 software (Treestar Inc., USA).

Analysis of atherosclerotic lesion development

Arteries were perfusion-fixed in situ with phosphate buffered saline (PBS) followed by 4% buffered formaldehyde paraformaldehyde in PBS (PFA; Sigma Aldrich, USA). The heart and whole aorta were removed and carefully cleaned of extraneous fat before being post-fixed in 4% PFA. The aortic root was then cut 5-μm transverse cryosections. Aortic root sections were assessed for atherosclerotic plaque size after staining with Oil-Red-O. All images were recorded with a Leica DM 4000B fluorescence microscope and JVC KY-F75U camera. Plaque size was quantified by computerized image analysis (Diskus Software, Hilgers, Germany).
Quantitative Real-Time PCR

Total RNA was extracted from tissues using a Nucleo Spin RNAII extraction Kit (Macherey-Nagel, Düren, Germany) in accordance with the manufacturer’s instructions. Equal amounts of template RNA (1 µg) were used for cDNA synthesis; RNA was reverse transcribed using random hexamer primers (Thermo Scientific, St. Leon-Rot, Germany). Quantitative SYBR-green real-time PCR was performed on duplicate samples of template cDNA on an Applied Biosystems 7900HT fast real-time PCR system using specific primer pairs (Eurofins MWG Operon, Germany): *Ifng* forward 5’-GCTGTTTCTGGCTGTTACTGC-3’, reverse 5’-TCACCATCCTTTTGCCAGTTCC-3’; *Tnfa* forward 5’-CTGTAGCCCACGTCGTCAGC-3’, reverse 5’-GGTTGTCTTTGAGATCCATGC-3’; *Il12* quantitec primer assay QT01048334 (Qiagen, Hilden, Germany); *Ccl2* forward 5’-CCCACTCACCCTGCTA-3’, reverse 5’-TTACGGGTCAACTTCACATC-3’; *Ccl5* forward 5’-GCTGCCCTACCACCATCCTCCT-3’, reverse 5’-GGCACACACTGGCGGTTCCCTTC-3’; *Vcam1* forward 5’-TGATTGGGAGAGACAAAGCA-3’, reverse 5’-AACAACCAGATCCCACACTTT-3’; *Icam1* forward 5’-TGCCCTGGGGATGCACACT-3’, reverse 5’-GGCTGTAGGTGGGTCCGGGT-3’; *Pselectin* forward 5’-CTGCCCCATGCTCTGTGGGC-3’, reverse 5’-CCCCAACCACCTGCTCGGTCT-3’. *Nos2* forward 5’-GTTCTCAGCCCATGCTCTGTGGGC-3’, reverse 5’-GTTCTCAGCCCATGCTCTGTGGGC-3’. *Pparg* forward 5’-TTATAGCTGTCATTATTCTCACTGAG-3’, reverse 5’-GACTCTGGGTTGGGAGAAGGCTGAC-3’. *Arg1* forward 5’-GGAAAGCCAATGAAGAGCTG-3’, reverse 5’-GATGCTTTCCACAGGACACAG-3’. *Mrc1* forward 5’-GATGCTTTCCACAGGACACAG-3’.
CACTCATCCATTACAACCAAAGC-3’, reverse 5’- CAGGAGGACCACGGTGAC-3’.
After an initial denaturation at 95°C for 10 minutes, 40 cycles of denaturation at 95°C for 15 seconds and annealing at 60°C for 60 seconds was carried out. After PCR amplification, dissociation curves were constructed to confirm formation of the intended PCR products. Quantitative measurements were determined using the ΔCt method, with Hprt as the housekeeping gene.

In vitro co-culture assays
Bone marrow derived macrophages were obtained by culturing total bone marrow cells from Ldlr−/− mice in RPMI medium containing 10% heat inactivated FCS, antibiotics and 15% L929 Cells Conditioned Medium as a source of M-CSF. CD8+ T cells were isolated from the spleen of Ldlr−/− mice using anti-mouse CD8a microbeads and magnetic cell sorting according to the manufacturer’s instructions (Miltenyi, Bergisch Gladbach, Germany). Bone marrow derived macrophages or SVEC endothelial cells were serum starved and cultured alone or with isolated CD8+ T cells at a 1:1 ratio in RPMI containing 0.5% FCS, 5 µg/ml anti-CD3 and 1 µg anti-CD28 antibody, with addition of 10 µg/ml anti-IFNγ neutralizing antibody or 10 µg/ml of isotype control antibody. After a 24h incubation, conditioned medium were collected and chemokine expression assayed by ELISA. Dendritic cells (DCs) and naïve OTII CD4+ T cells were isolated from spleens using magnetic selection kits (CD11c and Naïve T cell Isolation Kit) from Miltenyi, according to the manufacturer’s instructions. DCs were loaded with 250 ng/ml OVA323-339 peptide and naïve OTII CD4+ T cells labeled with 5 µM CFSE, and cultured at a 1:3 ratio for 3 days. Proliferation was assessed by CFSE dilution while intracellular staining for IFNγ (as described above) and Foxp3 were used for Th1 and Treg polarization analysis.
ELISA

Blood was extracted by retro-orbital puncture and placed in a Serum Collection Tube (Sarstedt, Nümbrecht, Germany). Serum was then collected according to the manufacturer’s instruction. CCL2, CXCL1 and CCL5 levels in serum and cell culture supernatants were measured using the corresponding ELISA development kits according to the manufacturer’s instructions (Peprotech, Hamburg, Germany).

Statistical Analysis

Results are expressed as mean ± SEM and were analyzed by a Student’s t-test or 1-way analysis of variance (ANOVA) followed by Bonferonni post-testing for multiple comparisons using Prism GraphPad 6.0 software. Differences with p values < 0.05 were considered to be statistically significant.
Figure S I: CD8+ T cells in atherosclerotic vessels and macrophage levels in aortae of anti-CD8α treated hypercholesterolemic mice

A) Quantitative analysis of absolute CD8+ T cell counts in single cell suspensions obtained from the aorta of Ldlr-/- mice on normal chow and after 6 weeks of HFD feeding. B) Distribution of CD4+ and CD8+ subsets in total T cells in the aortic root and aorta of chow and HFD-fed Ldlr-/- mice and C) representative FACS plot of T cell subsets in the aorta of chow or HFD fed Ldlr-/- mice. D) Representative CD8+ vs. CD4+ T cell gating in pre-gated CD45+CD3+TCRβ+ T cells and E) quantitative analysis of absolute CD8+ and CD4+ T cell counts in the aorta of Ldlr-/- mice fed a HFD and treated with Control (Ctrl) or a-CD8α antibody for 6 weeks (n=5-10/group). F) Absolute counts of CD45+CD11b+F4/80+ macrophages in single cell suspensions obtained from the aorta of Ldlr-/- mice fed a HFD and treated with Control (Ctrl) or a-CD8α antibody for 6 weeks (n=5-10/group). **p<0.01, ***p<0.001.
Figure S II. CD8⁺ T cell depletion.
Circulating counts of A) CD8⁺ and C) CD4⁺ T cells; C) CD8⁺ and D) CD4⁺ T cells counts in the spleen. All data were obtained from Ldlr⁻/⁻ mice fed HFD and treated with Control (Ctrl) or a-CD8α antibody for 6 weeks (n=5 - 8/group). **p<0.01, ***p<0.001.
Figure S III: CD4⁺ T cell activation in anti-CD8α treated hypercholesterolemic mice

Percentages of activated (CD44⁺CD62L⁻) cells in the A) spleen and B) blood, CD25⁺Foxp3⁺ regulatory T cells in the C) spleen and D) blood and IFNγ⁺ cells in the E) spleen and F) blood among CD4⁺ T cells in Ldlr⁻/⁻ mice fed a HFD for 6 weeks and treated with Control (Ctrl) or a-CD8α antibody (n=5-8/ group).
Figure S IV: Effect of anti-CD8α treatment on the DCs.
Percentages of A) CD8α+ and b) CD11b+ DCs among CD11c+MHCIi+ dendritic cells in the spleen and C) absolute counts of CD11c+MHCIi+ DCs in total aorta and aortic roots of Ldlr−/− mice fed a HFD for 6 weeks and treated with control or anti-CD8α antibodies. (D-F) CD11c+ DCs were isolated from the spleens of Ldlr−/− mice fed a HFD for 6 weeks and treated with control or anti-CD8α antibodies with anti-CD11c magnetic beads, loaded with OVA323-339 peptide and assayed for their ability to induce D) proliferation of naive OTII splenic CD4+ T cells, and E) IFNγ+ Th1 or F) Foxp3+ Treg polarization.
Figure S V: Role of CD8+ T cells in macrophage polarization.

A) Expression of iNOS (Nos2) mRNA relative to Hprt in macrophages cultured alone (Ctrl), with activated CD8+ T cells or with activated CD8+ T cells in presence of an a-IFNγ neutralizing antibody. Expression of B) iNOS (Nos2), C) Arg1, D) Mrc1 and E) Pparg mRNA relative to Hprt in the aortic sinus of Control (Ctrl) or anti-CD8α-treated Ldlr<sup>-/-</sup> mice fed a HFD for 6 weeks (n=6/group).
Figure S VI: Adhesion molecule expression in the aortic sinus
Expression of A) Vcam1, B) Icam1, C) Pecam1 and D) Pselectin mRNA relative to Hprt in the aortic sinus of Control (Ctrl) or anti-CD8α Ldlr<sup>−/−</sup> mice fed a HFD for 6 weeks (n=7-9/group).
Figure S VII: Blood monocytes and CCL2 serum levels in chow fed mice
A) Circulating levels of Ly6Chhi monocytes and B) serum levels of CCL2 in normcholesterolemic mice treated with Control (Ctrl) or anti-CD8α antibody for 6 weeks (n=3-5/group).
Figure S VIII: Myeloid progenitors in spleens of HFD-fed mice.
A) Absolute numbers of MP; B) frequencies of CMP among MP and C) absolute cell numbers; D) frequencies of GMP among MP and E) absolute cell numbers in spleens of Control (Ctrl) or anti-CD8α-treated Ldlr−/− mice fed a HFD for 6 weeks (n=7-8/group). *p<0.05.
Figure S IX: Effect of CD8+ T cells depletion on neutrophil levels.
Absolute numbers of neutrophils in A) bone marrow, B) blood, C) spleen, D) aortic root and E) aorta of control (Ctrl) or anti-CD8α treated Ldlr−/− mice fed a HFD for 6 weeks (n=6-8/group). *p<0.05.
Figure S X: Myeloid progenitors in chow fed mice bone marrow.
A) Frequencies of MP among total cells and B) absolute numbers of MP in the femur; C) frequencies of CMP among MP and D) absolute cell numbers; E) frequencies of GMP among MP and F) absolute cell numbers in bone marrow of control (Ctrl) or anti-CD8α-treated Ldlr<sup>−/−</sup> mice fed a normal chow for 6 weeks (n=6/group). *p<0.05.
Figure S XI: Myeloid progenitors in spleens of chow fed mice.
A) Absolute numbers of MP; B) frequencies of CMP among MP and C) absolute cell numbers; D) frequencies of GMP among MP and E) absolute cell numbers in spleens of control (Ctrl) or anti-CD8α-treated Ldlr<sup>−/−</sup> mice fed a HFD for 6 weeks (n=6/group).
Figure S XII: Monocyte levels in bone marrow and spleens of chow fed mice.

a) Percent of CD115^+Ly6C\textsuperscript{hi} monocytes in bone marrow cells and B) splenic counts of CD115^+Ly6C\textsuperscript{hi} monocytes in control (Ctrl) or anti-CD8\textalpha treated Ldlr^-/- mice fed a normal chow for 6 weeks (n=6/group). **p<0.01.
**Figure S XIII: CD8⁺ T cell depletion and plaque formation in anti-CD8β treated mice.**

CD8⁺ T cells counts in the A) blood and B) spleen; C) Lesion coverage in the aortic arch, D) lesion size in the aortic root and E) macrophage content of aortic root plaques in mice treated with Ctrl or anti-CD8β antibodies and fed a HFD for 6 weeks. n=7-11/group. *p<0.05, ***p<0.001.
Figure S XIV: Monocytes and bone marrow myeloid progenitors in anti-CD8β treated mice.

Blood counts of A) Ly6C\textsuperscript{hi} and B) Ly6C\textsuperscript{lo} monocytes and of C) neutrophils; bone marrow counts of D) Ly6C\textsuperscript{hi} monocytes, E) common myeloid progenitors (CMP) and F) granulocyte-monocyte progenitors; and G) splenic counts of Ly6C\textsuperscript{hi} monocytes in mice treated with Ctrl or anti-CD8β antibodies and fed HFD for 6 weeks. n=8-9/group.
Figure S XV: CD8+ T cell depletion reduces levels of GM-CSF and IL-6 producing cells in the bone marrow. Representative FACS plots of A) GM-CSF and D) IL-6 expression in total bone marrow cells and quantification of B) GM-CSF+ cells, E) IL-6+ cells, C) CD45-GM-CSF+ and F) CD45-IL-6+ cells in the bone marrow of Ldlr−/− mice fed a HFD for 6 weeks and treated with control (Ctrl) or anti-CD8β antibodies. n=10-11/group. *p<0.05, **p<0.01.
Figure S XVI: CD8⁺ T cells are the main source of IFNγ in the bone marrow.
Representative gating of A) IFNγ-producing cells among total CD45⁺ leukocytes and B) CD8⁺ vs. CD8⁻ cells; C) distribution of CD8⁺ vs. CD8⁻ cells among IFNγ-producing cells in the bone marrow of Control (Ctrl) antibody treated mice after 6 weeks of HFD feeding. D) Percentages of IFNγ-producing cells among total CD45⁺ cells in the bone marrow of Ctrl vs. or anti-CD8α treated Ldlr⁻/⁻ mice after 6 weeks of HFD feeding (n=7-8/group). **p<0.01, ***p<0.001.
Figure S XVII: IFNγ neutralization in hypercholesterolemic mice
Analysis of A) MP frequencies in percent of total bone marrow cells and B) CMP and C) GMP frequencies among MP in bone marrow, absolute counts of D) Ly6Chi and e) Ly6Clo monocytes in blood and F) total monocytes in percent of total bone marrow cells in control and anti-IFNγ treated Ldlr−/− mice after 3 weeks of HFD feeding (n=5-12/group); *p<0.05, **p<0.01.