Hypertension affects one-third of Western populations and increases in frequency with age, such that 70% of adults develop this disease by age 70. Hypertension is also a major risk factor for stroke, myocardial infarction, renal failure, and heart failure, and therefore is an enormous healthcare burden. Despite its prevalence, the pathogenesis of most cases of hypertension remains unclear.

Abstract: For >50 years, it has been recognized that immunity contributes to hypertension. Recent data have defined an important role of T cells and various T cell–derived cytokines in several models of experimental hypertension. These studies have shown that stimuli like angiotensin II, deoxycorticosterone acetate-salt, and excessive catecholamines lead to formation of effector like T cells that infiltrate the kidney and perivascular regions of both large arteries and arterioles. There is also accumulation of monocyte/macrophages in these regions. Cytokines released from these cells, including interleukin-17, interferon-γ, tumor necrosis factorα, and interleukin-6 promote both renal and vascular dysfunction and damage, leading to enhanced sodium retention and increased systemic vascular resistance. The renal effects of these cytokines remain to be fully defined, but include enhanced formation of angiotensinogen, increased sodium reabsorption, and increased renal fibrosis. Recent experiments have defined a link between oxidative stress and immune activation in hypertension. These have shown that hypertension is associated with formation of reactive oxygen species in dendritic cells that lead to formation of gamma ketoaldehydes, or isoketals. These rapidly adduct to protein lysines and are presented by dendritic cells as neoantigens that activate T cells and promote hypertension. Thus, cells of both the innate and adaptive immune system contribute to end-organ damage and dysfunction in hypertension. Therapeutic interventions to reduce activation of these cells may prove beneficial in reducing end-organ damage and preventing consequences of hypertension, including myocardial infarction, heart failure, renal failure, and stroke. (Circ Res. 2015;116:1022-1033. DOI: 10.1161/CIRCRESAHA.116.303697.)

Key Words: angiotensin II ■ antigen presenting cell ■ cytokines ■ effector T cell ■ nitric oxide synthase ■ sodium
Inflammation and Hypertension

Nonstandard Abbreviations and Acronyms

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Definition</th>
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<tr>
<td>CCL</td>
<td>ligands for chemokines with two adjacent cysteines</td>
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<td>CD</td>
<td>cluster of differentiation</td>
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<td>DC</td>
<td>dendritic cells</td>
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<td>DOCA</td>
<td>deoxycorticosterone acetate</td>
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<td>eNOS</td>
<td>endothelial nitric oxide synthase</td>
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<td>IFN</td>
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<td>MHC</td>
<td>major histocompatibility complex</td>
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<td>RAG</td>
<td>recombinase activating gene</td>
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<td>TH</td>
<td>T helper</td>
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<td>TNF</td>
<td>tumor necrosis factor</td>
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Adult hypertension, or essential hypertension, remains poorly defined. Perturbations of the kidneys, vasculature, and central nervous system have all been implicated in hypertension. In the past several years, it has become increasingly evident that hypertension is an inflammatory process that involves the transmigration and accumulation of both innate and adaptive immune cells into the interstitium of affected tissues where they release cytokines and promote oxidative stress. In this review, we will discuss how these cells contribute to dysfunction of the kidney and vasculature, promoting blood pressure elevation and end-organ damage.

Historical Perspectives

The concept that immune cells contribute to hypertension is not new. Almost one-half century ago, Grollman and White showed that immunosuppression lowers blood pressure in rats with partial renal infarction and found that these animals showed that immunosuppression lowers blood pressure in rats.1 Almost one-half century ago, Grollman and White showed that immunosuppression lowers blood pressure in rats. They noted “The cellular reaction was predominantly composed of mononuclear cells derived from the blood. The majority … looked like lymphocytes, and the rest like typical monocytes.” He went on to describe the time course and location of the cellular infiltration. “The reaction began as a sticking phenomenon corresponding to the damaged endothelium followed by a penetration of mononuclear cells into the arteriolar walls … A marked periarteriolar cellular infiltration like that seen in cases of chronic hypertensive vascular disease in different experimental animals was produced…” In a subsequent article published in 1972,2 Dr Olsen showed that vascular inflammation occurs in humans with a variety of causes of hypertension. Again, he noted “The cellular infiltration was composed of mononuclear cells exclusively which adhered to the surface of the endothelium of the vessels or had penetrated into the tunica media or the adventitia.” Indeed, subsequent studies as described below have identified the adventitia and perivascular adipose tissue of both large and small vessels as sites of immune cell accumulation in hypertension.

After the early observations by Grollman, White, and Olsen, several studies appeared supporting the role of immune cells in hypertension. These described perturbations of antibodies in the spontaneously hypertensive rat,3−7 and reduced hypertensive responses in athymic nude mice. Bendich et al found that treatment with antithymocyte serum lowers blood pressure in the spontaneously hypertensive rat,8 and the immunosuppressant cyclophosphamide was also found to have antihypertensive effects.9 Subsequent experiments by Finn Olsen showed that transfer of splenocytes from rats with deoxycorticosterone acetate (DOCA)-salt hypertension raises blood pressure in recipient rats.10

Thus, by the 1980s, a substantial body of data suggested that immune cells participate in hypertension, although the mechanisms were poorly understood. Unfortunately, this field seemed to stagnate for nearly 2 decades after these initial observations. This may partly have been as a result of a lack of understanding of the immune system and a paucity of tools available to further study this topic. Fortunately, the field of immunology has dramatically expanded in recent years. Our immunologist colleagues have defined subsets of innate and adaptive immune cells and gained insights into mechanisms by which the innate and adaptive immune systems interact. Myriad cytokines and chemokines have been identified that orchestrate inflammatory reactions, as have the signaling pathways that guide their synthesis. There has been an explosion of therapeutic interventions for immune/autoimmune diseases that can be used to probe events in hypertension, not only in animals but also in humans. Numerous mouse models have been produced that have provided enormous insight into the role of immune cells, cytokines, and cell trafficking in hypertension. These models have facilitated our ability to study hypertension in mice and rats.

In this review, we will discuss the more recent observations that support a role of the immune system in hypertension. We will consider the contribution of immunity and inflammation in blood pressure elevation and its parallel roles in producing end-organ damage. We will also consider mechanisms by which hypertensive stimuli activate the immune system and finally discuss therapeutic options that might prove useful in the treatment of hypertension.

Brief Primer of the Immune System

The immune system has 2 major components, the innate and the adaptive systems, which closely interact. Innate immunity represents the first line of defense against invading organisms and comprises epithelial cells that provide a barrier to invasion, phagocytes that engulf and destroy foreign organisms, and the complement system that helps to kill and clear pathogens. Innate immune cells contain pattern recognition receptors and Toll-like receptors that recognize pathogen-associated molecular motifs, such as lipopolysaccharide, lipoteichoic acid, flagellin, double stranded RNA, and unmethylated cytosine and guanine triphosphate deoxynucleotides with phosphodiester linkage. On binding, these signal a series of cellular events, including cytokine and chemokine production, expression of the inducible nitric oxide (NO) synthase, and production of reactive oxygen species (ROS). Importantly, the effects of
Toll-like receptor ligation can be influenced by the autonomic nervous system. As an example, Tracey and colleagues have described an inflammatory reflex, in which locally released cytokines and prostaglandins activate vagal afferent nerves that transmit information to the brainstem, the hypothalamus, and higher centers. Reflex signals from the brain stem alter behavior, reduce heart rate variability, increase vagal efferent activity, and increase sympathetic outflow. Vagal and sympathetic nerves promote acetylcholine release from a unique population of T cells, which, in turn acts on nicotinic receptors on nearby macrophages to dampen cytokine production and reduce the inflammatory response. A recent study from Harwani et al has shown that although Toll-like receptor priming normally reduces production of interleukin (IL)-6 and IL-1β in response to nicotinic stimulation, these responses are paradoxically increased in young spontaneously hypertensive rat, even before the onset of hypertension.

Monocytes and macrophages are components of the innate immune system that are particularly relevant to cardiovascular diseases. Uptake of oxidized low-density lipoprotein by macrophages leads to foam cell formation and represents an early step in atherogenesis. Macrophages are also a source of ROS that alter vascular reactivity, promote inflammation, and together with matrix metalloproteinases, contribute to vascular remodeling. These cells are also potent sources of cytokines, including IL-6 and tumor necrosis factor (TNF)α, which are discussed in depth later in this review. Historically, macrophages were thought to derive from circulating monocytes; however, it is now clear that some tissues contain resident macrophages that derive from a unique lineage. Moreover, recent evidence indicates that monocytes can traffic in and out of tissues without becoming macrophages.

In contrast to the innate immune response, the adaptive immune response is designed to respond specifically to foreign antigens. In the case of T cell activation, antigen-presenting cells (APCs), including dendritic cells (DCs), macrophages, and B cells, process foreign proteins to small peptides that are presented in major histocompatibility complexes (MHC). There is division of duty, so that MHC-I activates cluster of differentiation (CD)8+ T cells, whereas MHC-II present to CD4+ T cells. The peptide/MHC complex is recognized by a unique T cell receptor (TCR), representing signal 1. In addition, other receptor/ligand interactions occur together with the TCR/MHC interaction for the immunologic synapse. One such signal is the costimulatory interaction between CD28 and the B7 ligands, referred to as Signal 2. In the absence of this interaction, full T cell activation does not occur. T cells possess numerous accessory receptors that modify response to Signals 1 and 2, proliferation, and cytokine production (Signal 3).

Figure 1. Classical pathway for T cell activation. Antigen presenting cells, including dendritic cells, B cells, macrophages, and others process foreign antigens to peptides and present these in major histocompatibility complexes (MHC). MHC-I present to CD8+ T cells, whereas MHC-II present to CD4+ T cells. The peptide/MHC complex is recognized by a unique T cell receptor (TCR), representing signal 1. In addition, other receptor/ligand interactions occur together with the TCR/MHC interaction for the immunologic synapse. One such signal is the costimulatory interaction between CD28 and the B7 ligands, referred to as Signal 2. In the absence of this interaction, full T cell activation does not occur. T cells possess numerous accessory receptors that modify response to Signals 1 and 2, proliferation, and cytokine production (Signal 3).

In treatment of autoimmune diseases and transplant rejection. We have used one such agent, abatacept, to show that T cell costimulation is essential for hypertension and that treatment of mice with this agent after onset of either angiotensin II– or DOCA-salt–mediated hypertension lowered blood pressure. We also found that B7-deficient mice are resistant to blood pressure elevation in response to angiotensin II. Of note, hypertension was associated with an increase in CD86 in spleen and lymph node DCs. This study defined an important role of costimulation and, more importantly, DCs in hypertension.

T cells develop in the thymus from bone marrow–derived precursors and migrate to secondary lymphoid organs, such as the spleen and lymph nodes, where they reside in a quiescent (naïve) state until activated by an antigen-presenting cell. TCR ligation and costimulation lead to a cascade of signaling events that cause T cell oligoclonal expansion, cytokine production, and an alteration in surface proteins that allow egress of cells from secondary lymphoid organs to peripheral tissues. These effector T cells are guided to sites of infection or tissue damage by chemokines and cytokines released from the affected tissues. Once present in the peripheral tissues, effector T cells produce mediators that further orchestrate an inflammatory response. Importantly, this response is limited in duration, and resolution signals, such as cytotoxic lymphocyte antigen 4, programmed cell death protein 1, and others lead to apoptosis of most effector T cells. A few of these cells remain as memory cells, which can either return to the secondary lymphoid organs as central memory cells or remain in the periphery as effector memory cells. These memory cells can rapidly respond to a second challenge when the original antigen is re-encountered.

Four distinct effector phenotypes of CD4+ cells exist, 3 of which seem to have evolved to respond to specific pathogen challenges. T helper (Th1) cells, which produce interferon (IFN)-γ, are classically associated with intracellular bacteria...
and viruses. TH1 cells respond to helminthes and other allergens. The newest T helper subclass, T\textsubscript{H}\textsubscript{17} cells, protect against extracellular bacteria and fungi. A fourth T helper subset, termed T regulatory cells (Tregs), are immunosuppressive. The cytokine milieu and the nature of antigen presentation that guide naive T cells to these various subsets have been reviewed extensively elsewhere\textsuperscript{16,17} and will be mentioned in the context of hypertension and cardiovascular disease in this review. Of note, CD8\textsuperscript{+} T cells, although generally considered to function by releasing cytotoxic molecules like perforin and granzyme B, can also produce cytokines and can be classified in a manner similar to CD4\textsuperscript{+} T cells (ie, T\textsubscript{c}1, T\textsubscript{c}2, T\textsubscript{c}17, and cytokotoxic T reg cells). These aspects of T cell function have been reviewed in depth.\textsuperscript{18}

A concept relevant to cardiovascular disease and hypertension is immune senescence. After involution of the thymus in early adulthood, naive T cells decline and memory cells, particularly effector memory CD8\textsuperscript{+} T cells, expand.\textsuperscript{19} This is in part driven by recurrent and persistent viral infections.\textsuperscript{20} After repeated divisions, these cells assume a senescent phenotype characterized by shortened telomeress, loss of the costimulatory factors CD27 and CD28, and an increase in the surface marker CD57. Because of the absence of costimulatory receptors, senescent T cells are incapable of classical activation by TCR engagement. Nevertheless, these cells exhibit a state of persistent proinflammatory activation, producing IFN-\gamma, IL-6, and TNF\textalpha. Senescent CD8\textsuperscript{+} T cells also produce large amounts of cytotoxic granzyme. Senescent T cells have been recovered from atherosclerotic plaques of humans with unstable angina,\textsuperscript{21} and rheumatoid arthritis has been associated with premature T cell aging and accumulation of senescent T cells in the synovium.\textsuperscript{22} Recently, Youn et al found that relatively young hypertensive humans have increased circulating CD8\textsuperscript{+} T cells that are deficient in CD28 and produce excessive IFN-\gamma, perforin, and granzyme.\textsuperscript{23} The contribution of these cells in human hypertension has yet to be defined, but their profile of cytokine production indicates that they might play a critical role.

**Role of the Immune System in Blood Pressure Elevation**

As mentioned earlier, the immunology field has generated several genetically altered animal models that have proven extremely useful for studies of hypertension. The recombinase activating genes (RAG) 1 and 2 are responsible for recombining the genetic sequences that encode immunoglobulins and the T cell receptor. Mice lacking either of these fail to develop B and T lymphocytes. Several years ago, our group found that RAG-1\textsuperscript{-/-} mice develop blunted hypertension in response to either angiotensin II or DOCA-salt challenge. In these studies, we found that hypertension was associated with accumulation of T cells with an effector phenotype in the perivascular adipose tissue and adventitia. Adoptive transfer of T (but not B) cells into RAG-1\textsuperscript{-/-} mice restored hypertension and its attendant end-organ dysfunction.\textsuperscript{24} Subsequently, we found that stress-induced hypertension is also reduced in RAG-1\textsuperscript{-/-} mice and restored by adoptive transfer of T cells. Others have confirmed these findings. Crowley and coworkers showed that mice with severe combined immune deficiency are protected against hypertension.\textsuperscript{25} In parallel with our findings in mice, Mattson et al deleted the RAG-1 gene in Dahl salt–sensitive rats using zinc finger DNAse technology and showed that this not only blunted hypertension but also reduced renal damage caused by salt feeding.\textsuperscript{26} In more recent studies, this group has deleted CD247, the CD3\epsilon chain, selectively removing T cells in Dahl salt–sensitive rats. This led to a similar phenotype to that observed with RAG-1 deletion, blunting the blood pressure elevation and renal damage caused by salt feeding.

In keeping with these experimental findings, several studies have shown that the T cell suppressing agent mycophenolate mofetil reduces blood pressure and renal inflammation in experimental models of hypertension.\textsuperscript{27-29} This agent has also reduced blood pressure in a small number of hypertensive patients with rheumatoid arthritis or psoriasis.\textsuperscript{30}

There is also evidence that innate immune cells have an important role in hypertension. De Ciuceis et al studied mice lacking the macrophage colony stimulating factor, also referred to as osteoporosis spontaneous mutation factor, which exhibit severe osteoporosis and growth retardation and are deficient in monocytes and macrophages.\textsuperscript{31} These animals exhibited minimal elevation of blood pressure in response to chronic angiotensin II infusion and had preserved endothelial–dependent vasodilatation of the resistance mesenteric vessels. Although angiotensin II stimulated an increase in NADPH oxidase activity in the vessels of normal mice, this response was markedly blunted in the osteoporosis spontaneous mutation mice. Subsequent studies further demonstrated that osteoporosis spontaneous mutation mice mice are also resistant to DOCA-salt hypertension.\textsuperscript{32} Wenzel et al used lysozyme M–targeting of the diphtheria toxin receptor to delete monocytes from mice.\textsuperscript{33} The authors showed that angiotensin II–induced hypertension increased aortocyte/macrophages and markers of aortic inflammation, characterized by increased mRNA for the vascular cell adhesion molecule-1, cyclooxygenase 2, and the inducible nitric oxide synthase. Hypertension was also associated with increase in vascular superoxide production and endothelial dysfunction. Diphtheria toxin treatment prevented virtually all of these vascular perturbations and the hypertension caused by angiotensin II infusion. Hypertension could be restored by adoptively transferring monocytes but not granulocytes to these mice. As discussed later, we have identified a critical role of monocyte-derived DCs in hypertension, and it is conceivable that loss of these cells in the osteoporosis spontaneous mutation mice mice and the elimination of monocytes in the study by Wenzel et al reduced the contribution of these important cells.

**Cytokines Contributing to Hypertension**

At first glance, it is difficult to understand how the immune system could contribute to hypertension. Studies from our group and others have shown that hypertension is associated with accumulation T cells and monocyte/macrophages in vessels and the kidney. In keeping with Olsen’s observations 40 years ago, the vascular accumulation is predominantly in the adventitia and the perivascular fat. In the kidney, T cells increase in both the medulla and renal cortex. We have found
that these cells have markers suggestive of effector memory cells. Data from our group and others indicate that these cells produce potent cytokines that affect vascular and renal function. In the past several years, important roles of ≥5 cytokines have been identified in hypertension, which will be discussed below.

**Interleukin 17**

A particularly important cytokine is IL-17, produced by a unique set of CD4+ T cells, referred to as Th17 cells. In addition to CD4+ T cells, γ/δ T cells, the Th17 subset of CD8+ T cells, some B cells and natural killer T cells, and miscellaneous other cells produce IL-17. There are 6 isoforms of IL-17, classified as A–F, which share varying degrees of sequence homology. IL-17A and IL-17F are the most closely related isoforms (sharing 50% sequence homology) and are generally produced by the same cell types. These isoforms are encoded on the same chromosome and can bind as homo- or heterodimers to the IL17 receptor complex composed of IL17RA and IL17RC. These receptor chains are thought to heterodimerize on ligand binding and signal through the adaptor molecule Act1 and tumor necrosis factor–associated factor-6, leading to nuclear factor kappa B, mitogen-activated protein kinase and CCAAT-enhancer–binding protein-δ activation.

Importantly, IL-17A seems to act synergistically with other cytokines, in particular TNFα, to stimulate these signals. As an example, Sharma et al have shown that IL-17 and TNFα synergistically enhance lung inflammation and chemokine C-X-C motif-1, CCL-2, and CCL-5 production after ischemia/reperfusion in an NADPH oxidase–dependent fashion. IL-17A has been implicated in a variety of autoimmune diseases, including psoriasis, experimental autoimmune encephalomyelitis, asthma, Crohn’s disease, and rheumatoid arthritis. The anti–IL-17A antibodies, Ixekizumab and Secukinumab, as well as the anti-IL17RA receptor antibody, Brodalumab, have proven effective in phase II/III human trials for the treatment of psoriasis and are being studied in other inflammatory diseases.

In recent years, several reports indicate that IL-17A contributes to hypertension. We found that angiotensin II infusion increases IL-17A production from mouse T cells and that plasma levels of IL-17A are increased in humans with hypertension. Importantly, mice lacking IL-17A develop blunted hypertension and do not develop endothelial dysfunction in response to angiotensin II infusion. In keeping with this, the increase in vascular superoxide production generally observed in hypertension does not occur in mice lacking IL-17A. A particularly striking finding was that the vascular infiltration of total leukocytes and T cells was markedly reduced in IL-17A−/− mice. In keeping with this finding, we found that when coadministered with TNFα to human vascular smooth muscle cells in culture, IL-17A increased expression of a variety of cytokines and chemokines, including CCL8, macrophage colony–stimulating factor-3, chemokine C-X-C motif-2, and CCL7. Thus, it seems that IL-17A might coordinate an inflammatory response, leading to accumulation of multiple cell subtypes in hypertension.

Subsequent studies have confirmed a role of IL-17A in hypertension. Nguyen et al made the fascinating discovery that IL-17A can induce phosphorylation of threonine 495 on the endothelial nitric oxide synthase (eNOS) in a Rho kinase–dependent manner. Phosphorylation at this site prevents calmodulin binding and leads to conformational changes of eNOS, reducing production of NO. These investigators also showed that infusion of IL-17A in mice caused a modest elevation of blood pressure in the absence of other hypertensive stimuli.

Recently, Amador et al found that rats with DOCA-salt hypertension have a striking increase in circulating Th17 cells and reduced Tregs and that treatment with spironolactone reverses this pattern. The investigators also found a marked increase in the mRNA for IL-17A in the heart and kidney, and these values were normalized by spironolactone. Interestingly, triple therapy with hydralazine, reserpine, and hydrochlorothiazide, which normalized blood pressure, did not reduce these elevated IL-17 mRNA levels, suggesting that the increase in this cytokine was not purely a consequence of blood pressure elevation. The investigators also found that treating rats with an antibody against IL-17A reduced blood pressure and levels of collagen I in the heart and kidneys.

In keeping with a role of IL-17A in collagen deposition, we have recently discovered that this cytokine plays a critical role in aortic stiffening. A normal compliant aorta expands during systole and collapses during diastole, respectively, storing and then ejecting a portion of the cardiac stroke volume during these 2 phases of the cardiac cycle. Aortic stiffening occurs in a variety of disease conditions, including aging and hypertension, leading to loss of this Windkessel or capacitance function. This results in rapid transmittance of blood volume to the periphery, an increase in systolic blood pressure, and a decrease in diastolic blood pressure. Recently, we found that both angiotensin II and DOCA-salt–induced hypertension lead to a striking deposition of collagen in the adventitia of mice and a marked loss of aortic compliance. Interestingly, this aortic stiffening was found to be caused by the presence of T cells because it did not occur in RAG-1−/− mice and was restored by adoptive transfer of T cells to these animals. Moreover, our studies implicated IL-17A as a causative cytokine in aortic stiffening, as collagen deposition and aortic stiffening did not occur in IL-17A−/− mice. Subsequent studies of cultured aortic fibroblasts showed that IL-17A induced mRNA for collagens I, III, and V in a p38 mitogen–activated protein kinase–dependent fashion.

Of interest, transforming growth factor-β alone promotes formation of Tregs and there is substantial plasticity of these cells, such that exposure to IL-6 in conjunction with transforming growth factor-β can convert Tregs to Th17 cells. Preeclampsia has been associated with an imbalance of Tregs and Th17 cells. Toldi et al found a modest increase in circulating T cells containing IL-17A in patients with preeclampsia. In keeping with this, Cornelius et al found a striking increase in circulating Th17 cells in a placental ischemia model of preeclampsia and showed that IL-17 inhibition using a soluble form of the IL-17 receptor C ameliorated many aspects of preeclampsia in this model, including the blood
pressure elevation. An important cause of preeclampsia is the production of agonistic antibodies against the angiotensin type I receptor, and these were markedly reduced by this novel treatment strategy.

There is one study that has shown a worsening of renal damage in mice lacking either IL-17 or IL-23. These authors, however, subjected the experimental animals to uninephrectomy, DOCA-pellet implantation, salt-feeding, and an extraordinarily high dose of angiotensin II infusion. Other investigators, including our group, have found that either lower doses of angiotensin II or the DOCA-salt challenge alone is sufficient to cause hypertension, and their combined use of high dose angiotensin II with DOCA-salt hypertension likely induced severe vascular and renal injury. Given the multiple other papers showing a role of IL-17 in promoting hypertension, it is likely that the paradoxical finding of this report was as a result of the unusual model used by these authors.

**Interferon Gamma**

The type II interferon, IFN-γ, is the signature cytokine of Tc1 cells, but is also produced in high amounts by CD8 T cells (Tc1 cells) and natural killer T cells. We have consistently found an increase in IFN-γ-forming CD4+ and CD8+ T cells in hypertensive mice; however, it seems that this cytokine might have mixed effects on the response to various hypertensive stimuli. Ishimitsu et al found that subcutaneous injections of IFN-γ attenuated hypertension, proteinuria, and glomerular injury in Dahl salt–sensitive rats, although having no effect in spontaneously hypertensive rats. This study and others where the cytokine was administered may not reflect the role of endogenously produced IFN-γ in hypertension. More recently, Garcia et al demonstrated a blunted hypertensive response to chronic aldosterone infusion (combined with uninephrectomy and salt-feeding) in IFN-γ−/− mice compared with wild-type animals. However, IFN-γ−/− mice exhibited exaggerated left ventricular (LV) hypertrophy, reduced LV cavity size, and worse diastolic dysfunction.

We have recently shown that mice deficient in the lymphocyte adapter protein, LNK, exhibit severe hypertension and renal/vascular dysfunction. Interestingly, these mice have elevated numbers of CD4+ and particularly CD8+ T cells that produce IFN-γ. In addition, we showed that IFN-γ deficiency results in blunted hypertension in response to angiotensin II infusion.

Marko et al recently examined a role of IFN-γ in hypertension by studying mice lacking IFN-γ receptor 1. Although these animals did not exhibit an alteration in the hypertensive response to high dose angiotensin II infusion, they developed less renal fibrosis and maintained their glomerular filtration rate. These animals also demonstrated reduced cardiac fibronectin and collagen and fewer inducible arrhythmias than observed in wild-type mice subjected to angiotensin II infusion.

An important mechanism by which IFN-γ might promote hypertension relates to its capacity to induce angiotensinogen expression in both hepatocytes and renal proximal tubular cells. Although angiotensinogen is not considered rate limiting for the systemic production of angiotensin II, its role in the tubular production of angiotensin II seems more critical. Navar and colleagues have shown that proximal tubular cells produce angiotensinogen, which is subsequently converted to angiotensin I and angiotensin II within the tubule or within epithelial cells. This locally formed angiotensin II promotes sodium and volume reabsorption in both the proximal and distal nephron. In the proximal tubule, this is mediated by actions on apical sodium hydrogen exchanger 3, basolateral Na/HCO3 cotransport, and the Na-K ATPase. In the distal nephron, renin produced by collecting duct cells act on proximally derived angiotensinogen to form additional intratubular angiotensin II, which has multiple actions, including activation of the epithelial sodium channel and the sodium chloride cotransporter. Of particular interest, recently Satou et al have shown that prolonged exposure of renal proximal tubule cells to IFN-γ increases angiotensinogen protein production >2-fold in a Janus kinase-2/signal transducer and activator of transcription-3–dependent fashion. Thus, it is conceivable that infiltrating T cells that release IFN-γ might modulate the local production of angiotensinogen, enhance sodium reabsorption, and worsen hypertension in a feed-forward fashion. In keeping with this, Bravo et al demonstrated that mycophenolate mofetil reduced the presence of angiotensin II–producing cells of the kidney, some of which were proximal tubular epithelial cells.

Interestingly, Kamat et al recently showed that mice deficient in IL-17A or IFN-γ exhibit alterations in proximal or distal tubule sodium transporters, such as sodium hydrogen exchanger 3 and the sodium chloride cotransporter, leading to enhanced pressure natriuresis and decreased distal sodium reabsorption, respectively. Thus, in addition to affecting the local renin–angiotensin system, proinflammatory cytokines may have direct effects on the expression of renal sodium transporters and thus on salt and water balance.

**Tumor Necrosis Factor α**

TNFα is produced by a variety of cells, including T cells, macrophages, endothelial cells, fibroblasts, and neuronal cells. It acts on 2 receptors TNFRI and TNFR2, which are ubiquitously expressed and form homotrimers on TNFα binding. These receptors in turn activate multiple signals, including death and survival pathways, NADPH oxidase activation, c-Jun N-terminal kinases, and nuclear factor kappa B. Nuclear factor kappa B and NAPDH oxidase activation contribute to several of the cardiovascular and renal effects of TNFα, including chemokine and adhesion molecule expression, vascular remodeling, and sodium retention by the kidney. TNFα has a multitude of untoward effects on endothelial NO production that could contribute to hypertension. Superoxide production by the NADPH oxidase rapidly reacts with NO, forming the strong oxidant peroxynitrite. TNFα also inhibits the eNOS promoter and causes destabilization of the eNOS mRNA, ultimately reducing eNOS protein levels and the ability of the endothelium to produce NO. Recent studies have shown that microRNA 155 contributes to destabilizing the eNOS mRNA and reducing endothelium-dependent vasodilatation in response to TNFα. Thus, like IL-17A, TNFα impairs the ability of the endothelium to produce NO, promoting vasoconstriction.

In addition to its vascular effects, TNFα has renal effects that could affect blood pressure. Ramseyer and Garvin have
recently reviewed this topic. As in the case of the endothelium, TNFα decreases eNOS expression in medullary thick ascending limb cells within 24 hours of exposure in a Rho-kinase–dependent manner. NO inhibits sodium reabsorption at several sites along the renal tubule, including the mTAL and collecting duct, and its loss would therefore lead to sodium retention. Loss of NO in the vasa vasorum could also promote sodium reabsorption in the renal medulla by perturbing tubulovascular crosstalk. Prolonged and sustained exposure of renal parenchymal cells to TNFα could promote renal injury and thus shift the pressure natriuresis curve to favor blood pressure elevation.

In keeping with a role of TNFα in hypertension, we found that angiotensin II infusion stimulates T cells to produce TNFα and that the TNFα antagonist etanercept blunts the blood pressure elevation and vascular superoxide production caused by angiotensin II. TNFα blockade has also proven effective in either reducing blood pressure or protecting against renal injury in various models of hypertension, including preeclampsia, a model of lupus erythematosus, angiotensin II infusion in rats, transgenic rats, and fructose feeding. Despite the fact that TNFα antagonists have been used extensively to treat humans with a variety of autoimmune diseases, there is no evidence that they have blood pressure lowering effects. There are conflicting reports that these agents affect vascular stiffness in humans.

Sriramula et al demonstrated a virtual absence of blood pressure elevation and a reduction of left ventricular hypertrophy in response to chronic angiotensin II infusion in TNFα−/− mice. An interesting finding in this study was that the water drinking behavior induced by angiotensin II was markedly reduced in the TNFα-deficient mice, indicating a central role of this cytokine. In keeping with this, activation of nuclear factor kappa B in the hypothalamus occurred in wild-type, but not TNFα−/− mice. These findings are in keeping with a role of hypothalamic inflammation in hypertension.

TNFα inhibitors are widely used in the treatment of humans with autoimmune diseases. To date, there are no reports that these agents lower blood pressure in humans; however, mixed effects on arterial stiffness have been described.

**Interleukin 6**

IL-6 is a small, 21 kDa glycoprotein produced by numerous cells, including DCs, macrophages, monocytes, subsets of TH1 T cells, and vascular cells. IL-6 binds to its receptor, IL-6R, which dimerizes with the transmembrane protein glycosylated protein 130, which is ubiquitously expressed and has a large intracellular signaling domain that activates the Janus kinase tyrosine kinases and the downstream signal transducer and activator of transcription-3. Cells not expressing IL-6R can be transactivated by IL-6 binding to soluble IL-6R, which in turn binds to glycosylated protein 130. Via these signaling pathways and others, IL-6 induces multiple effects on target tissues, stimulating bone resorption, neutrophil chemotaxis, and polarization of helper T cells. Because it stimulates hepatocyte production of C-reactive protein, elevations of this acute phase reactant often reflect the actions of IL-6. As mentioned earlier, IL-6 is a major signal to promote polarization of CD4+ T cells to produce IL-17. Experimental studies have implicated IL-6 in diverse inflammatory conditions, including malignancies, autoimmune diseases, atherosclerosis, and hypertension. Circulating levels of IL-6 are increased in humans with polymyalgia rheumatica and giant cell arteritis and are suppressed by corticosteroid therapy. The humanized anti-IL-6R antibody Tocilizumab has proven effective in treatment of rheumatoid and juvenile arthritis and is being evaluated for treatment of several other inflammatory diseases, including Crohn’s disease, systemic sclerosis, and ankylosing spondylitis.

There is substantial evidence that IL-6 contributes to hypertension. Levels of IL-6 correlate with blood pressure in hypertensive subjects and are reduced by treatment with angiotensin II-receptor blockade. In keeping with this, angiotensin II infusion in humans increases IL-6 levels, and this is blocked by treatment with spironolactone, implicating activation of the mineralocorticoid receptor. Lee et al found that IL-6−/− mice develop blunted hypertension and albuminuria in response to high salt and angiotensin II infusion compared with wild-type mice. In cultured cortical collecting duct cells, IL-6 increases the protein levels and activity of the epithelial sodium channel and therefore has the potential to enhance sodium and volume resorption in vivo. This direct effect of IL-6, together with its ability to skew T cells from a regulatory phenotype toIL-17-producing cells, is likely important in hypertension.

**Interleukin 10/T Regulatory Cells**

Interleukin 10 is an anti-inflammatory cytokine that was originally discovered based on its ability to inhibit IL-2 and IFN-γ production. A variety of cells, almost all lymphocytes, monocytes, macrophages, DCs, and endothelial cells, can produce IL-10. IL-10 inhibits cytokine production by T cells and monocyte/macrophages and reduces DC maturation, diminishing antigen presentation, MHC, and the costimulatory B7 molecule expression. In T cells, ligation of the TCR stimulates IL-10 production in a feedback loop fashion.

There is evidence that IL-10 has protective functions in hypertension. The -627 polymorphism of the IL-10 promoter is associated with a reduced incidence of essential hypertension in Russian Tatars. Didion et al have defined a critical role of IL-10 in modulating endothelial function in hypertension. These investigators found that direct application of angiotensin II doubled superoxide production in carotid arteries of IL-10−/− mice, but not in WT mice. These investigators also found that direct application of angiotensin II markedly impaired vasodilatation caused by acetylcholine in IL-10−/− mice, although having no effect in vessels of WT mice. This defect in vascular function was corrected by treatment with a cell permeable form of superoxide dismutase, linking increased oxidative stress to vascular dysfunction in IL-10-deficient vessels. The hypertensive response to 10 days of angiotensin II infusion was similar between WT and IL-10−/− mice. Of interest, the aortic levels of IL-6 mRNA were markedly increased, and TNFα mRNA slightly increased by angiotensin II in IL-10−/− mice compared with values observed in WT aortas.

There is also evidence that IL-10 ameliorates hypertension associated with pregnancy. Placentas from women with preeclampsia exhibit reduced staining for IL-10 compared with women with normal gestation, and serum levels of IL-10
are reduced in preeclampsia. Tinsley et al demonstrated that daily IP injections of recombinant IL-10 normalized blood pressure and endothelial function in pregnant rats with DOCA-salt hypertension. This model was associated with proteinuria and increased circulating levels of endothelin-1 and IFN-γ, all of which were reduced by IL-10. In addition, placental levels of IFN-γ and PECAM-1 were elevated by DOCA-salt hypertension and corrected by IL-10 treatment. Likewise, Lai et al showed that exposure of wild-type pregnant mice to hypoxia led to hypertension, proteinuria, reduced fetal weight, and renal injury and that these parameters were more severe in IL-10−/− mice. Treatment with recombinant IL-10 corrected hypertension, proteinuria, and fetal weight in the IL-10-deficient animals. Of note, increased circulating levels of the soluble vascular endothelial cell growth factor-l receptor soluble fms–like tyrosine kinase have been implicated in the pathogenesis of preeclampsia. The investigators found that hypoxia also increased soluble fms–like tyrosine kinase in this model and that IL-10 treatment normalized these levels.

Although numerous cells can produce IL-10, it is a key product of Tregs. Adoptive transfer of Tregs has been shown to lower angiotensin II– and aldosterone–induced hypertension, cardiac fibrosis, coronary inflammation, and electric remodeling. These effects are likely mediated at least in part by release of IL-10 from Tregs. As Kassan et al showed, IL-10−/− mice exhibit enhanced hypertension, endothelial dysfunction, and increased NADPH oxidase activity in response to angiotensin II, and these effects were ameliorated by adoptive transfer of Tregs from WT but not IL-10−/− mice. In keeping with this concept, Viel et al studied Dahl salt–sensitive rats that had undergone transfer of chromosome 2 from normotensive Brown Norway rats. These animals exhibited enhanced T regulatory cell markers, increased IL-10 levels, and reduced parameters of vascular inflammation. Taken together, these studies support an anti-inflammatory and antihypertensive role of T regulatory cells in response to multiple stimuli.

**Novel Mechanism of T Cell Activation in Hypertension: Role of Oxidative Stress**

Despite the aforementioned studies, it has remained unclear as to how and why T cells are activated in hypertension. Recently, we have established a new mechanism of T cell activation related to oxidative stress. More than 15 years ago, we discovered that angiotensin II and DOCA-salt hypertension lead to an increase in vascular superoxide production because of activation of the NADPH oxidase. Studies from others have shown that the NADPH oxidase plays a role in hypertension at many levels. In the subfornical organ of the brain, ROS produced by this enzyme complex stimulate neuronal firing, and in the kidney, this enzyme stimulates sodium reabsorption. We now have discovered that the NADPH oxidase is activated in DCs in hypertension, and this leads to formation of γ-ketoaldehydes, also known as isoketals. These are products of fatty acid oxidation, which rapidly react with protein lysines. Recent evidence indicate that these oxidatively modified proteins are immunogenic, and we find that isoketal protein adducts abundantly accumulate in DCs of mice made hypertensive by either angiotensin II or DOCA-salt hypertension. This is particularly evident in CD11c+CD11b+ DCs, which are stimulated to express high levels of CD80 and CD86 when they form isoketal adducts. DCs affected in this manner produce large amounts of IL-6, IL-23, and IL-1β, known to drive T cell polarization. Indeed, we find that these DCs drive T cell proliferation and production of IL-17A, TNFα and IFN-γ, which as noted above contribute to hypertension. Adoptive transfer of these DCs to recipient mice primes severe hypertension in response to low dose angiotensin II. Moreover, induction of oxidative stress in DCs by addition of tert-buty1 hydroperoxide promotes formation of isoketal-added proteins and hypertension when these tert-buty1 hydroperoxide–treated DCs are transferred to recipient mice.

In these studies, we also examined the efficacy of 2-hydroxybenzylamine, a compound known to scavenge isoketals before they adduct proteins. We found 2-hydroxybenzylamine virtually eliminated isoketal-added proteins in DCs, prevented DC expression of CD80 and CD86, and production of polarizing cytokines. Importantly, 2-hydroxybenzylamine prevented the capacity of DCs to drive T cell proliferation and the ability of DCs to convey hypertension to recipient mice when administered to the donor mice. We studied several related compounds, some of which were known to scavenge isoketal and others modified to be inactive and found that the active compounds had antihypertensive properties.

These recent studies from our group define a new mechanism of hypertension and T cell activation as depicted in Figure 2. Given that oxidative stress contributes to a variety of diseases, including atherosclerosis, diabetes mellitus, and obesity, it is possible that isoketal adducts promote immune mechanisms in these conditions as well and that drugs like 2-hydroxybenzylamine might be beneficial in these conditions.
Concluding Remarks

Classic teachings from Guyton and coworkers indicate sustained hypertension requires an alteration in renal function followed by an increase in systemic vascular resistance. Dr Guyton described several physiological and pathophysiological phenomena that alter the renal pressure natriuresis curve and proposed that this renal event is followed by systemic autoregulation, which causes an increase in systemic vascular stiffness and increased recruitment of immune cells, propagating the inflammatory response. These effects result in vascular dysfunction. In the renal medulla and cortex, activated T cells produce cytokines, such as IL-6 and interferon (IFN)γ that stimulate production of angiotensinogen. Angiotensinogen is converted to angiotensin I (Ang I) by intrarenal renin and subsequently to angiotensin II (Ang II) by intrarenal angiotensin-converting enzyme. Angiotensin II upregulates and stimulates transport channels in the proximal and distal convoluted tubules, including the sodium hydrogen exchanger 3 (NHE3) and sodium chloride cotransporter (NCC). In conjunction with salt and water retention, T cell activation causes an increase in renal ROS production, and renal injury and fibrosis, all of which lead to renal dysfunction. The culmination of vascular and renal dysfunction caused by T cell–derived cytokines exacerbates hypertension (Illustration credit: Ben Smith).

We devote substantial effort to defining blood pressure goals in the treatment of hypertension, but the major reason to treat this disease is to prevent end-organ damage. If hypertension did not cause strokes, myocardial infarctions, heart failure, renal failure, and dementia, it is unlikely that physicians would even measure blood pressure. The data discussed in this review supports the notion that an important component of the end-organ damage associated with hypertension is mediated by inflammation. Thus, although blood pressure lowering is important, an underlying goal should clearly be prevention of the local inflammation that accompanies this disease. We currently have no drugs to specifically accomplish this, except by indirect means. Moreover, our ability to detect local inflammation is limited to assays of circulating markers. Substantial additional research is needed to understand this field in detail and to develop diagnostic and therapeutic tools for management of this widespread health problem.

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References


68. Lee DL, Sturgis LC, Labazi H, Osborne JB Jr, Fleming C, Pollock JS, Manhiani M, Imig JD, Brands MW. Angiotensin II hypertension is
Inflammation, Immunity, and Hypertensive End-Organ Damage
William G. McMaster, Annet Kirabo, Meena S. Madhur and David G. Harrison

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