Sudden cardiac death (SCD) refers to death after an unexpected sudden cardiac arrest in a patient with or without known structural heart disease. The incidence of SCD in the United States ranges from 300,000 to 460,000 events per year, depending on the criteria for SCD used for surveillance. SCD results from a complex interaction between preexisting cardiac substrates, either structural or genetic, with superimposed physiological or environmental triggers. The most common underlying causal disorders for SCD in adults aged ≥35 are coronary heart disease (65%–80%) and dilated cardiomyopathy (10%–15%). Various types of cardiomyopathy (eg, hypertrophic cardiomyopathy, arrhythmogenic right ventricular cardiomyopathy, infiltrative, inflammatory, and valvular diseases), genetically determined rhythm disorders (eg, long-QT syndrome, Brugada syndrome, and catecholaminergic polymorphic ventricular tachycardia), or developmental disorders (anomalous origins of coronary arteries) account for most of the remaining SCDs.

**Abstract:** Ventricular arrhythmia is the leading cause of sudden cardiac death (SCD). Deranged cardiac metabolism and abnormal redox state during cardiac diseases foment arrhythmogenic substrates through direct or indirect modulation of cardiac ion channel/transporter function. This review presents current evidence on the mechanisms linking metabolic derangement and excessive oxidative stress to ion channel/transporter dysfunction that predisposes to ventricular arrhythmias and SCD. Because conventional antiarrhythmic agents aiming at ion channels have proven challenging to use, targeting arrhythmogenic metabolic changes and redox imbalance may provide novel therapeutics to treat or prevent life-threatening arrhythmias and SCD. (Circ Res. 2015;116:1937-1955. DOI: 10.1161/CIRCRESAHA.116.304691.)

**Key Words:** cardiac arrhythmias ■ death, sudden, cardiac ■ metabolism ■ oxidative stress

**Mechanisms of Sudden Cardiac Death**

**Oxidants and Metabolism**

Kai-Chien Yang, John W. Kyle, Jonathan C. Makielski, Samuel C. Dudley Jr

Original received October 19, 2014; revision received February 8, 2015; accepted February 9, 2015. In April 2015, the average time from submission to first decision for all original research papers submitted to Circulation Research was 13.84 days.

From the Department of Pharmacology (K.-C.Y.) and Division of Cardiology, Department of Internal Medicine (K.-C.Y.), National Taiwan University Hospital, Taipei, Taiwan; Division of Cardiovascular Medicine, Department of Medicine, University of Wisconsin, Madison (J.W.K., J.C.M.); and Lifespan Cardiovascular Institute, the Providence VA Medical Center, and Brown University, RI (S.C.D.).

Correspondence to Samuel C. Dudley, MD, PhD, The Warren Alpert Medical School of Brown University, 593 Eddy St, APC730, Providence, RI 02903. E-mail samuel_dudley@brown.edu; or Jonathan C. Makielski, MD, Department of Medicine, University of Wisconsin, 600 Highland Ave, MC CSC-3248, Madison, WI 53792. E-mail jcm@medicine.wisc.edu

© 2015 American Heart Association, Inc.

Circulation Research is available at http://circres.ahajournals.org

DOI: 10.1161/CIRCRESAHA.116.304691
Overview of Cardiac Ionic Channels and Membrane Excitability

Normal functioning of the mammalian heart depends on proper electric activity involving the initiation of the electric impulse from pacemaker cells, the propagation of the electric activity through specialized conduction system and myocardium, and the generation of action potentials in individual myocytes. A normal cardiac cycle begins with the action potential originating from the cells in sinoatrial node, conducts through the atria, atrioventricular node, His bundle, Purkinje fibers, and then spreads throughout the entire ventricular myocardium. The proper propagation of cardiac electric impulse depends on low resistance pathways between cells via gap junctions, which are formed by docking of 2 connexin hemichannels on appositional sarcolemmal membranes.

Cardiac action potentials are generated through the coordinated activity of various types of ion channels and transporters (Figure 1). Inward current conducting through the voltage-gated Na⁺ channel rapidly depolarizes the cell (phase 0), which is followed by early repolarization (phase 1) attributed to the transient outward K⁺ current (Iₒ). Depolarizing L-type Ca²⁺ currents (I_CaL) and multiple repolarizing delayed rectifier K⁺ currents (I_K) form the plateau phase (phase 2), which is followed by rapid repolarization (phase 3) with inactivated I_CaL and increasing I_K. The repolarization continues with the contribution of inwardly rectifying K⁺ current, I_K1, until return to the resting membrane potential (phase 4). The Ca²⁺ influx during the subsequent depolarizations earlier (early afterdepolarization [EAD]) or later (delayed afterdepolarization [DAD]) in the repolarization phase.

All 3 mechanisms can result from abnormal functioning of myocardial ion channels and transporters, leading to disordered initiation or propagation of cardiac action potentials. Accumulating evidence suggests that altered cardiac ion channel/transporter function is closely linked to abnormal myocardial metabolic activity and imbalanced redox states in a wide range of cardiac pathology. This review presents the current evidence on the acute effects of abnormal myocardial metabolism and increased oxidative stress on myocardial ion channel/transporters that predispose to ventricular arrhythmias and SCD. It is important, however, to recognize that abnormal metabolism and oxidative stress in noncardiac myocytes tissues can also contribute to the development of ventricular arrhythmias. For example, altered metabolism and redox state have been implicated in vascular tissue leading to atherosclerosis, which underlies the main substrate for SCD and coronary heart disease. Another example is that abnormal metabolism and increased oxidative stress affect autonomic nervous system, thereby contributing to ventricular arrhythmias and SCD. Within the ventricle, chronic effects of such entities as diabetes mellitus and ischemia from atherosclerosis work through mechanisms involving metabolic and oxidative abnormalities to create the substrate of structural heart diseases that leads to SCD. Both the role of aberrant metabolism and oxidants in the chronic creation of such substrate will not be further considered.

### Membrane Excitability

<table>
<thead>
<tr>
<th>Nonstandard Abbreviations and Acronyms</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>ΔΨ_m</td>
<td>mitochondrial membrane potential</td>
</tr>
<tr>
<td>AMPK</td>
<td>AMP-activated protein kinase</td>
</tr>
<tr>
<td>APD</td>
<td>action potential duration</td>
</tr>
<tr>
<td>CaMKII</td>
<td>calcium/calmodulin-dependent protein kinase II</td>
</tr>
<tr>
<td>Cav1</td>
<td>caveolin-1</td>
</tr>
<tr>
<td>DAD</td>
<td>delayed afterdepolarization</td>
</tr>
<tr>
<td>EAD</td>
<td>early afterdepolarization</td>
</tr>
<tr>
<td>ETC</td>
<td>electron-transport chain</td>
</tr>
<tr>
<td>FADH₂</td>
<td>flavin adenine dinucleotide</td>
</tr>
<tr>
<td>I_CaL</td>
<td>L-type Ca²⁺ currents</td>
</tr>
<tr>
<td>I_K</td>
<td>delayed rectifier K⁺ currents</td>
</tr>
<tr>
<td>IMAC</td>
<td>inner membrane anion channel</td>
</tr>
<tr>
<td>INa</td>
<td>Na⁺-current</td>
</tr>
<tr>
<td>IPC</td>
<td>ischemic preconditioning</td>
</tr>
<tr>
<td>I_to</td>
<td>transient outward K⁺ current</td>
</tr>
<tr>
<td>Kv</td>
<td>voltage-gated K⁺ channels</td>
</tr>
<tr>
<td>NADH</td>
<td>nicotinamide adenine dinucleotide</td>
</tr>
<tr>
<td>NADPH</td>
<td>nicotinamide adenine dinucleotide phosphate</td>
</tr>
<tr>
<td>NCX</td>
<td>Na⁺/Ca²⁺ exchanger</td>
</tr>
<tr>
<td>NHE</td>
<td>Na⁺/H⁺ exchanger</td>
</tr>
<tr>
<td>NO⁺</td>
<td>nitric oxide</td>
</tr>
<tr>
<td>NOS</td>
<td>nitric oxide synthase</td>
</tr>
<tr>
<td>O²⁻</td>
<td>superoxide</td>
</tr>
<tr>
<td>PK</td>
<td>protein kinase</td>
</tr>
<tr>
<td>PTP</td>
<td>mitochondrial permeability transition pore</td>
</tr>
<tr>
<td>RAS</td>
<td>renin-angiotensin system</td>
</tr>
<tr>
<td>ROS</td>
<td>reactive oxygen species</td>
</tr>
<tr>
<td>RyR2</td>
<td>ryanodine receptor 2</td>
</tr>
<tr>
<td>sarcoK_ATP</td>
<td>sarcolemmal K⁺ ATPase</td>
</tr>
<tr>
<td>SCD</td>
<td>sudden cardiac death</td>
</tr>
<tr>
<td>SERCA</td>
<td>sarco/endoplasmic reticulum Ca²⁺-ATPase</td>
</tr>
<tr>
<td>SR</td>
<td>sarcoplasmic reticulum</td>
</tr>
</tbody>
</table>

and causes of SCD have been extensively reviewed previously and in the articles on epidemiology of sudden cardiac death in this compendium series.

The physiological mechanisms reported to cause SCD vary with the patient population and the criteria used to define SCD. In general, ventricular tachyarrhythmias, including ventricular fibrillation and pulseless ventricular tachycardia, are the most common electrophysiological mechanisms leading to SCD. Other physiological events that can result in SCD include pulseless electric activity, bradyarrhythmias, and asystole. This review will focus only on SCDs attributed to ventricular tachyarrhythmias.

Ventricular tachyarrhythmias, like all cardiac arrhythmias, result from 1 of 3 primary mechanisms: reentry, abnormal automaticity, or triggered activity. Reentry is the most common mechanism for ventricular tachyarrhythmias, involving the presence of an anatomic or functional disturbance of cardiac electric impulse propagation and of heterogeneous conduction. Abnormal automaticity results from the accelerated generation of an action potential by a region of ventricular cells. Triggered activity occurs when an action potential elicits
Action potential (AP) waveform and major ion channels/transporters in human ventricular cardiac myocytes. The AP waveform begins with the activation of Na⁺ channels ($I_{Na}$), leading to the rapid depolarization (phase 0), followed by inward repolarizing transient outward K⁺ currents ($I_{to}$) in phase 1, forming the notch of the AP waveform. The balance between the inward L-type Ca²⁺ current ($I_{CaL}$) and outward delayed rectifier K⁺ currents ($I_{Kdr}$) contributes to the plateau (phase 2) of the AP waveform. $I_{Na}$ continues to repolarize the membrane potential (phase 3) and the inwardly rectifying K⁺ currents ($I_{K1}$) contributes to the later part of phase 3 repolarization and the maintenance of resting membrane potential (phase 4). During the AP, Ca²⁺ influx via $I_{CaL}$ triggers the release of Ca²⁺ ions from the sarcoplasmic reticulum (SR) via ryanodine receptor 2 (RyR2), coined Ca²⁺-induced Ca²⁺-release. During the diastolic phase, SR/endoplasmic reticulum Ca²⁺-ATPase (SERCA) retrieves cytosolic Ca²⁺ to SR, and Na⁺/Ca²⁺ exchanger (NCX) extrudes Ca²⁺ from the cell, bringing cytosolic [Ca²⁺] back to baseline. Sarcolemmal K⁺ channels (sarcKATP), gated by intracellular ATP/ADP levels and acidosis, does not contribute to AP waveform under normal circumstances, but play an important role in regulating cellular metabolism and electrophysiological responses to metabolic and oxidative stresses. Na⁺/K⁺ ATPase consumes ATP to pump Na⁺ outside of the cell in exchange for K⁺, which is critical for the maintenance of Na⁺ and K⁺ gradients across the plasma membrane. Cx43 indicates connexin 43.

Cardiac Oxidants and Oxidative Stress

In biological systems, partially reduced forms of oxygen (O₂⁻) lead to the formation of oxidants or reactive oxygen species (ROS), which oxidize other molecules when these molecules accept electrons. The main ROS in cardiac myocytes exist in radical forms, such as superoxide (O₂⁻•) and hydroxyl radicals, or in nonradical forms, such as hydrogen peroxide (H₂O₂), hypochlorite, nitric oxide (NO), and peroxynitrite. It is important to note that although grouped under the acronym of ROS, different reactive species containing molecular oxygen vary greatly in diffusion coefficient, reactivity, and oxidation potential.
Under physiological conditions, trace amounts of signaling ROS, which are tightly regulated by the balance of ROS production and ROS scavengers, form a network that coordinates metabolism with gene transcription and enzymatic activity. Low levels of ROS produced during short periods of ischemia, for example, trigger signaling pathways that confer cardiac protection, a phenomenon coined ischemic preconditioning (IPC), which is mediated, at least in part, through mitochondrial KATP channels (see below). In addition, low levels of nicotinamide adenine dinucleotide phosphate (NADPH) oxidases, xanthine oxidase, nitric oxide synthase (NOS), and mitochondria. Figure 3 illustrates the major ROS synthesis pathways in cardiac myocytes. NADPH oxidases are membrane-bound enzymes that use NADH or NADPH as the electron donor to generate $O_2^-$ from $O_2$. Xanthine oxidase catalyzes the oxidation of hypoxanthine/xanthine and produces $O_2^-$, NOS, on the contrary, produces a single nitrogen radical (NO•) through metabolizing l-arginine to l-citrulline. NOS becomes uncoupled under conditions such as depletion of l-arginine, oxidation of the NOS cofactor tetrahydrobiopterin, or increased S-glutathionylation of NOS. Under these conditions, NOS produces $O_2^-$ instead of NO•. NO• itself is a weak thiol oxidant, but it regulates various physiological functions by activating cGMP/protein kinase G (PKG) pathways or by covalent modification of protein cysteine residues via S-nitrosylation or S-glutathiolation (GSS-). NO• also reacts with $O_2^-$ and forms a more potent oxidative molecule peroxynitrite. Oxidative molecules derived from NO•, such as NO•, $N_2O_3$, and peroxynitrite, are termed reactive nitrogen species. As the metabolic hub in cardiac myocytes, mitochondria consume most of the $O_2$ for energy production, thereby serving as the major source of ROS in the heart.

In the mitochondria, ROS are generated as an inevitable byproduct of OXPHOS, resulting from the premature leak of electrons from the ETC to molecular oxygen. It has been estimated that 0.1% to 1% of the electrons passing through the ETC prematurely leak to $O_2$ at complexes I, II, or III.
Mitochondria represent the predominant source of ROS in cardiac myocytes, producing superoxide ($O_2^•−$) by the premature leak of electrons from the respiratory chain at complexes I, II, or III. Mitochondrial manganese superoxide dismutase can dismutate $O_2^•−$ to hydrogen peroxide ($H_2O_2$), which can be broken down by transition metals such as Fe$^{2+}$ and converted to hydroxyl radicals (‘$OH$). Superoxide can also be generated by the reduction of $O_2$ by free electrons released from (1) NAD(P)H oxidases, which oxidize nicotinamide adenine dinucleotide (NADH) or nicotinamide adenine dinucleotide phosphate (NADPH) to form NAD+ or NADP+, (2) xanthine oxidase, which catalyzes the conversion of hypoxanthine to xanthine or xanthine to uric acid, (3) endothelial nitric oxide synthase (eNOS), which switches from a coupled state (producing NO$^•−$ to an uncoupled oxide (producing $O_2^−$) under conditions such as the reduction in $\Delta$-arginine, the deficiency of cofactor tetrahydrobiopterin (BH$_4$), or increased glutathionylation of eNOS. Superoxide can also interact with preformed NO$^•−$ and generate peroxynitrite ONOO$^−$.

Figure 3. Major reactive oxygen species (ROS) generation pathways in cardiac myocytes. Mitochondria represent the predominant source of ROS in cardiac myocytes, producing superoxide ($O_2^•−$) by the premature leak of electrons from the respiratory chain at complexes I, II, or III. Mitochondrial manganese superoxide dismutase can dismutate $O_2^•−$ to hydrogen peroxide ($H_2O_2$), which can be broken down by transition metals such as Fe$^{2+}$ and converted to hydroxyl radicals (‘$OH$). Superoxide can also be generated by the reduction of $O_2$ by free electrons released from (1) NAD(P)H oxidases, which oxidize nicotinamide adenine dinucleotide (NADH) or nicotinamide adenine dinucleotide phosphate (NADPH) to form NAD+ or NADP+, (2) xanthine oxidase, which catalyzes the conversion of hypoxanthine to xanthine or xanthine to uric acid, (3) endothelial nitric oxide synthase (eNOS), which switches from a coupled state (producing NO$^•−$) to an uncoupled oxide (producing $O_2^−$) under conditions such as the reduction in $\Delta$-arginine, the deficiency of cofactor tetrahydrobiopterin (BH$_4$), or increased glutathionylation of eNOS. Superoxide can also interact with preformed NO$^•−$ and generate peroxynitrite ONOO$^−$.

The mitochondrial ETC efficiency is impaired under pathological conditions, such as myocardial ischemia and heart failure, and diabetes mellitus resulting in increased electron leak and ROS production. Accumulating ROS levels trigger the opening of mitochondrial channels, including mitochondrial permeability transition pore (PTP) and inner membrane anion channel (IMAC) (Figure 2), leading to depolarization of the $\DeltaW_m$ and further acceleration of ROS production, a phenomenon termed mitochondrial ROS-induced ROS release. The PTP is a nonselective channel residing on the inner mitochondrial membrane, and it opens with elevated matrix [Ca$^{2+}$], increased ROS and phosphate levels, and by reduced
adenine nucleotides (ADP or ATP). Brief and reversible PTP opening is considered physiological and serves as a release mechanism to prevent overt mitochondrial cation (especially Ca\(^{2+}\)) and ROS accumulation. During myocardial ischemia, myocytes experience Ca\(^{2+}\) overload, high phosphate and low ATP concentrations. These conditions prime PTP for opening when reperfusion triggers ROS generation and mitochondrial Ca\(^{2+}\) accumulation, leading to prolonged PTP opening and burst ROS release from mitochondria. Pharmacological inhibition of PTP (with cyclosporine A or sanglifehrin A) provides protection against ischemia-reperfusion injury in various experimental models. Recent evidence suggests that dimers of mitochondrial ATP-synthase form the pore-forming component of PTP, and the c subunit of the ATP-synthase complex may be required for PTP-dependent mitochondrial fragmentation and cell death. IMAC, on the contrary, is responsible for anionic currents across the mitochondrial membrane, allowing the passage of O\(_2^•−\) generated by ETC from the matrix to the cytoplasmic side of the inner membrane, thereby serving as a mitochondrial O\(_2^•−\) efflux and ΔΨ\(_{\text{m}}\) depolarization mechanism. During ischemia and reperfusion, mitochondrial O\(_2^•−\) production is increased, which leads to increased permeability of IMAC, resulting in a burst in ROS release and ΔΨ\(_{\text{m}}\) depolarization. The resulting reversible ΔΨ\(_{\text{m}}\) collapse and membrane potential oscillation foster lethal ventricular arrhythmias, which can be abrogated by IMAC inhibitors.

During mitochondrial ROS-induced ROS release, depolarized mitochondrial ΔΨ\(_{\text{m}}\) depletes NADPH (the level of which is coupled to [NADH] by nicotinamide nucleotide transhydrogenase in mitochondria), thereby leading to rapid dissipation of ROS-eliminating capacity, allowing acceleration of ROS emission from mitochondria.

Elevated mitochondrial ROS also impairs ATP production through various mechanisms. Excessive ROS cause oxidative damage to components of the ETC, such as complex I, II, IV, and ATP synthase, impairing ATP generation and further aggravate electron leakage. In addition, O\(_2^•−\) activates mitochondrial uncoupling proteins, resulting in increased electron leak and uncoupled O\(_2\) consumption from ATP synthesis. Taken together, excessive cardiac ROS leads to abnormal electric function directly by ROS-mediated signaling and oxidative damage, as well as indirectly by decreasing ATP generation that is required for normal ion channel/transporters functioning.

Impaired cardiac metabolism and increased myocardial oxidative stress, depending on the cause and clinical course, can be divided into 2 categories: acute and chronic. Acute cardiac oxidative and metabolic derangements typically occur with myocardial ischemia/reperfusion that results from coronary artery occlusion. Chronic oxidative and metabolic abnormalities, in contrast, occur with conditions such as cardiac hypertrophy, heart failure, and diabetes mellitus. The impact of chronic oxidative and metabolic stresses on cardiac electrophysiology often involves changes in the transcript and protein expression of cardiac ion channels/transporters, which is commonly defined as electric remodeling. Electric remodeling during these chronic cardiac conditions have been extensively reviewed elsewhere.
dynamics under physiological and pathological conditions. Recently, Williams et al provide quantitative data, suggesting that mitochondria do not function as a significant buffer of cytosolic Ca\(^{2+}\) under physiological conditions, with prolonged elevation of cytosolic [Ca\(^{2+}\)]; however, mitochondrial Ca\(^{2+}\) uptake can increase \(\leq1000\)-fold and affect cellular Ca\(^{2+}\) dynamics significantly. Nevertheless, it has been shown that mitochondrial dysfunction results in cardiac myocytes Ca\(^{2+}\) transient alternans by impairing the mitochondrial capacity of Ca\(^{2+}\) handling, thereby predisposing to cardiac arrhythmias.

In addition, pharmacological inhibition of mitochondrial uptake through mitochondrial Ca\(^{2+}\) uniporner with Ru360 has been shown to reduce the incidence of ventricular arrhythmias induced by ischemia-reperfusion in the rodent heart. As the metabolic center in cardiac myocytes, therefore, mitochondria play an important role in transducing changes in metabolic states to the dynamic regulation of sarcolemmal Ca\(^{2+}\), membrane excitability, and electric functioning.

**Table 1. Effects of Metabolic Changes on Cardiac Ion Channel/Transporter Function and Arrhythmogenicity**

<table>
<thead>
<tr>
<th>Channel/Transporter Effects</th>
<th>Metabolic Changes</th>
<th>Effects on Electric/Ionic Homeostasis</th>
<th>Proarrhythmic Mechanism</th>
</tr>
</thead>
<tbody>
<tr>
<td>Na(^{+}/K(^{-}) ATPase ↓</td>
<td>Ischemia/hypoxia</td>
<td>Na(^{+}) overload</td>
<td>Ca(^{2+}) overload and DAD</td>
</tr>
<tr>
<td>Cx hemichannel ↑</td>
<td>Ischemia</td>
<td>Na(^{+}) overload</td>
<td>Ca(^{2+}) overload and DAD</td>
</tr>
<tr>
<td>Peak (I_{\text{Na}}) ↓</td>
<td>Ischemia/heart failure</td>
<td>↓ Na(^{+}) influx</td>
<td>Slow conduction</td>
</tr>
<tr>
<td>Late (I_{\text{Na}}) ↑</td>
<td>Ischemia/hypoxia acidosis, ↑ LPC AMPK mutation</td>
<td>↑ Na(^{+}) influx, prolonged APD</td>
<td>EAD</td>
</tr>
<tr>
<td>Kv ↓</td>
<td>Diabetes mellitus, heart failure</td>
<td>↓ K(^{+}) influx, prolonged APD</td>
<td>EAD</td>
</tr>
<tr>
<td>Kv↑</td>
<td>Insulin treatment in diabetic heart, PI3K activation, exercise training</td>
<td>↑ K(^{+}) channel expression</td>
<td>Protective</td>
</tr>
<tr>
<td>(I_{\text{ATP}}) ↑</td>
<td>Ischemia</td>
<td>↑ K(^{+}) influx, shortened APD</td>
<td>Current sink, slow conduction</td>
</tr>
<tr>
<td>RyR2</td>
<td>↓ during ischemia, ↑ on reperfusion</td>
<td>SR Ca(^{2+}) load ↑, spontaneous Ca(^{2+}) waves</td>
<td>Ca(^{2+}) transient/action potential alternans</td>
</tr>
</tbody>
</table>

APD, action potential duration; Cx, connexin; DAD, delayed afterdepolarization; EAD, early afterdepolarization; \(I_{\text{ATP}}\), ATP-sensitive K\(^{+}\) current; Kv, voltage-gated K\(^{+}\) current; Late \(I_{\text{Na}}\), late Na\(^{+}\) current; LPC, lysophosphatidylcholine; Peak \(I_{\text{Na}}\), peak Na\(^{+}\) current; RyR2, ryanodine receptor 2; and SR, sarcoplasmic reticulum.

**Metabolic Regulation of Na\(^{+}\) Homeostasis and Na\(^{+}\) Channel Function**

Intracellular [Na\(^{+}\)] plays a critical role in regulating the energetics, electric functioning, and contractility of cardiac myocytes, which can be attributed to the direct and indirect effects of cytosolic [Na\(^{+}\)] on Ca\(^{2+}\) homeostasis, mitochondrial function, and cellular signaling. Multiple ion channels/transporters function to maintain the [Na\(^{+}\)] homeostasis in cardiac myocytes: Na\(^{+}/K\(^{-}\) ATPase, for example, consumes ATP to pump Na\(^{+}\) outside of the cell in exchange of K\(^{+}\), representing the main mechanism of Na\(^{+}\) efflux, whereas Na\(^{+}\) channels, NCX, and the Na\(^{+}/H\(^{+}\) exchanger (NHE) mediate Na\(^{+}\) influx in cardiac myocytes.

Cardiac voltage-gated Na\(^{+}\) channels form by the assembly of a pore-forming α subunit and auxiliary β subunits that modulate channel activities. Nav1.5 (SCN5A) is the predominant Nav α subunit expressed in the mammalian myocardium. Currents conducting through Na\(^{+}\) channels generate the rapid upstroke (phase 0) of the action potential. In addition, Na\(^{+}\) channels, together with cardiac gap junctions, govern the electric conduction velocity in the myocardium. There are 2 effects of Na\(^{+}\) channel/transporter dysfunction on arrhythmogenesis: (1) through affecting Na\(^{+}/Ca\(^{2+}\) homeostasis, thereby promoting delayed afterdepolarizations and creating reentry substrates and (2) through electrophysiological effects of the channel function. For example, increased late \(I_{\text{Na}}\) predisposes to action potential duration (APD) prolongation and EADS, both of which are arrhythmogenic and predispose to SCD (Table 1; Figure 4).

During the metabolic stress that ensues with myocardial ischemia/reperfusion, reduced Na\(^{+}/K\(^{-}\) ATPase (because of reduced ATP and elevated ADP/phosphate levels), increased late \(I_{\text{Na}}\) (see below for detailed mechanisms), and enhanced NHE (owing to increased cytosolic acidosis) activity result in increased cytosolic [Na\(^{+}\)], thereby leading to Ca\(^{2+}\) overload and increased propensity for ventricular arrhythmias (see above). It has been shown that pharmacological inhibition or knockout of NHE renders cardiac function resistant to ischemia-reperfusion injury.

In addition to altered Na\(^{+}/K\(^{-}\) ATPase and NHE function, it has been demonstrated that ischemia/metabolic inhibition can trigger Na\(^{+}\) influx through connexin hemichannels. Under normal conditions, connexin hemichannels located on adjacent intercellular sarcolemmal membranes combine to form gap junctions at the intercalated discs between cardiac myocytes. During ischemia/metabolic inhibition, connexin hemichannels localize to the nonjunctional membrane, turning into nonselective cation channels that are permeable to Na\(^{+}\), K\(^{+}\), and Ca\(^{2+}\). With the high conductance of these connexin hemichannels, it is suggested that the opening of even a small number of connexin 43 hemichannels can lead to doubling of Na\(^{+}\) influx in ventricular cardiac myocytes, contributing to increased Na\(^{+}\) load and increased propensity for arrhythmias (Table 1; Figure 4).

Elevated cytosolic [Na\(^{+}\)] also impairs cardiac metabolism reciprocally through affecting mitochondrial function. Mitochondrial [Na\(^{+}\)] is regulated by NHE-mediated Na\(^{+}\) uptake and mitochondrial Na\(^{+}/Ca\(^{2+}\) exchanger-mediated Na\(^{+}\) efflux. Under physiological conditions, energized mitochondria extrude H\(^{+}\), and the resulting pH gradient drives the Na\(^{+}\) gradient between mitochondrial matrix (lower [Na\(^{+}\)]) and cytosol (higher [Na\(^{+}\)]). Intracellular Na\(^{+}\) levels increase significantly in pathological conditions, such as myocardial
ischemia. The rise in cytosolic [Na+] during ischemia widens the Na+ gradient across mitochondria, leading to stronger driving force for Ca2+ extrusion from mitochondria via mitochondrial Na+–Ca2+ exchanger, thereby leading to reduced mitochondrial [Ca2+] and altered mitochondrial energetics, including decreased activities of Ca2+-dependent dehydrogenases in the tricarboxylic acid cycle, the net oxidation of the matrix NADH/NADPH pool, and hence the deficiency of NADPH-dependent ROS scavengers, thereby increasing mitochondrial ROS emission. Increased ROS levels further aggravate cytosolic Na+ accumulation by increasing late Ito, through direct Na+ channel modification or by activating signaling molecules such as PKC or CaMKII (see below).

Therefore, there may be a feed-forward mechanism between increased cytosolic [Na+] , impaired mitochondrial metabolism, increased ROS emission, and ROS-induced elevation of cytosolic [Na+] .

In addition to cytosolic [Na+] , cardiac metabolism also influences electric functioning through modulating sarcolemmal Na+ channel activity. During myocardial ischemia, various metabolic derangements result in altered Na+ channel properties and function. The ischemic metabolite lysophosphatidylcholine, for example, has been shown to decrease peak Na+ current (peak Ito) and to slow its inactivation. In addition, lysophosphatidylcholine also causes a marked increase in late Ito, thereby increasing myocardial Na+ loading. Myocardial acidosis during ischemia is also known to affect the inactivation states of Na+ channel, leading to increased late Ito (Table 1; Figure 4). Cardiac Na+ channels are reported to be modulated by AMP-activated PK (AMPK). AMPK is a serine/threonine kinase that is activated by increased cytosolic AMP/ATP ratio to increase ATP production and decrease ATP consumption, thereby functioning as a master metabolic regulator in cardiac myocytes. Mutations in PRKAG2 gene, which encodes the γ 2 subunit of AMPK, have been reported to be associated with electric instability and lethal ventricular arrhythmias. For example, PRKAG2 T172D, a constitutively active AMPK mutant, has been shown to slow Na+ channels inactivation, leading to increased late Ito, prolonged APD and arrhythmogenic EADs.

**Metabolic Regulation of K+ Channels and Cardiac Repolarization**

Various types of voltage-gated K+ (Kv) channels and nonvoltage-gated inwardly rectifying channels contribute to myocardial action potential repolarization. Under pathological conditions, such as myocardial ischemia, diabetes mellitus, and heart failure, repolarizing Kv currents in ventricular cardiac myocytes are reduced, leading to delayed repolarization and prolonged APD, which predispose to the development of ventricular arrhythmias and SCD (Figure 4).

Among several types of inwardly rectifying K+ channels functioning in mammalian heart, I K1 contributes to the terminal phase of repolarization and the maintenance of resting membrane potentials in ventricular myocytes, whereas currents conducted via sarcolemmal KATP (sarcKATP) channels, gated by intracellular ATP/ADP levels and acidosis, function as immediate sensors of cellular metabolism and play an essential role in electrophysiological responses to metabolic stresses such as myocardial ischemia.

Abnormal cardiac repolarization, including QT-interval prolongation and T-wave abnormalities, are often observed in patients with diabetes mellitus, one of the leading chronic metabolic disorders associated with ventricular arrhythmias and SCD. Studies on cellular mechanisms of diabetes mellitus–induced repolarization abnormalities have consistently revealed downregulation of cardiac Kv currents in diabetic heart. Multiple mechanisms have been proposed to account for reduced Kv currents during diabetes mellitus. First, post-translational inhibition of Kv channel function occurs in diabetes mellitus. Plasma levels of free fatty acid metabolites, such as palmitoylcarnitine and palmitoyl-CoA, are increased with diabetes mellitus. They have been shown to inhibit cardiac Kv currents directly. Acute treatment with insulin has been demonstrated to reverse diabetes mellitus–induced Kv current reduction, suggesting the primary role of insulin-dependent signaling on Kv channel function. It has been suggested that improved glucose utilization by insulin may be responsible for restored Kv current expression because compounds known to improve glucose utilization (such as dicholoroacetate or l-carnitine) have been shown to normalize cardiac Kv current density with diabetes mellitus.

Increased ROS levels also contribute to reduced cardiac Kv currents during diabetes mellitus, and direct incubation with antioxidant glutathione has been shown to restore Kv current expression in myocytes isolated from diabetic hearts. Second, transcriptional regulation of Kv channel expression may reduce Kv currents. Peroxisome proliferator-activated receptor α , a critical regulator of glucose/fatty acid metabolism, is upregulated in diabetic heart and has been suggested to transcriptionally repress cardiac Kv channel expression.

Cardiac-specific overexpression of peroxisome proliferator-activated receptor α downregulates the mRNA and protein expression levels of Kv4.2 and KChIP2, the channel subunits encoding Ito whereas peroxisome proliferator-activated receptor α knockout upregulates Ito. Interestingly, it has been recently demonstrated that the activation of phosphoinositide 3-kinase α , a key component of insulin receptor signaling pathway that can be triggered by insulin treatment or exercise training, upregulates mRNA levels of various Kv channels through an Akt-independent mechanism, which may provide the mechanistic explanation for insulin-mediated Kv current modulation, as well as the metabolic impact of exercise training on cardiac repolarization. It is important to note that the changes in Kv current observed with diabetes mellitus or exercise training reflect the chronic effects of metabolic stress on Kv channel remodeling, rather than acute channel effects.

The sarcKATP channels are present at high density in the sarcolemmal membrane. SarcKATP channels are inhibited by ATP and activated by ADP, Mg2+, or low pH, conditions that are associated with ischemia, insufficient fuel supply, and increased metabolic stress. On increased metabolic stress with ATP depletion and acidosis (eg, during myocardial ischemia), sarcKATP channels are activated, allowing an inwardly rectifying repolarizing K+ current. SarcKATP channels open...
within seconds in response to acute ischemia, which is likely attributed to the rapid drop in pH (also within seconds of ischemia) in the ischemic tissue. ATP levels, by contrast, deplete at a slower rate and remain in the millimolar range (levels that are sufficient to keep sarcK ATP channels closed) until 10 to 15 minutes after the onset of ischemia. Taken together, these data suggest that abnormal cardiac metabolism can lead to ventricular arrhythmias and SCD through non–ion channel mechanisms.

The cell surface expression density of sarcK ATP channels is high. It has been estimated that the opening of merely 1% of the sarcK ATP channels is sufficient to significantly shorten myocardial APD. The opening of sarcK ATP channels shortens APD and decreases inward Ca2+ currents, thereby reducing Ca2+-mediated cardiac energy consumption and preventing Ca2+ overload-induced cell death. With adequate numbers of sarcK ATP channels in the open state, however, cardiac myocytes become hyperpolarized and rendered unexcitable. This creates a current sink that slows or blocks electric propagation in the myocardium, predisposing to the development of conduction block that potentiates re-entrant type ventricular arrhythmias. Indeed, pharmacological inhibition of sarcK ATP channels has been shown to reduce the incidence of ventricular arrhythmias in animal models and in humans. Reduced mitochondrial ATP production with impaired respiratory chain function leads to the activation of sarcK ATP channels. During ROS-induced ROS release, for example, dissipation of ΔΨm results in cessation of mitochondrial ATP production, leading to sarcK ATP activation, altered myocyte APDs and even ventricular arrhythmias.

Interestingly, it has been shown that ATP generated from glycolysis plays a bigger role in modulating sarcK ATP activity than ATP generated from OXPHOS, which is in accordance with recent findings that the key glycolytic enzymes, including glyceraldehyde 3-phosphate dehydrogenase (GAPDH) and pyruvate kinase, serve as components of the sarcK ATP channel macrocomplex and regulate its function. SarcK ATP channels have also been shown to exhibit reduced sensitivity to ATP-mediated inhibition in diabetic heart, resulting in higher sarcK ATP conductance and greater APD shortening in response to hypoxia.

Non–Ion Channel Mechanisms Linking Metabolism to Ventricular Arrhythmias and SCD

As mentioned above, AMPK functions as a key metabolic sensor and regulator in cardiac myocytes. It has been reported that specific mutations in PRKAG2, the γ2 regulatory subunit of AMPK, lead to a glycogen storage cardiomyopathy characterized by ventricular pre-excitation and cardiac hypertrophy. The structural abnormalities associated with PRKAG2 mutations, including the glycogen accumulation and ventricular hypertrophy, result in abnormal conduction that predisposes to ventricular arrhythmias.

In addition to AMPK, patients with inherited fatty acid oxidation disorders exhibit high incidence of ventricular arrhythmias, conduction defects and SCD, which can be attributed to the accumulation of arrhythmogenic intermediary metabolites of fatty acids. Fatty acid metabolites, such as lysophosphatidylcholine, accumulate during myocardial ischemia, contributing to the formation of arrhythmogenic substrates. Consistent with these findings, a transgenic mouse model of cardiac-specific overexpression of fatty acid transport protein 1 (α-MHC FATP1) has been shown to exhibit increased myocardial uptake and accumulation of free fatty acids, leading to arrhythmogenic electric changes including QT prolongation. Taken together, these data suggest that abnormal cardiac metabolism can lead to ventricular arrhythmias and SCD through non–ion channel mechanisms.

Mechanisms Linking Increased Cardiac Oxidative Stress to Ventricular Arrhythmias and SCD

Myocardial Oxidative Stress and Cardiac Na+ Channels

Abnormal cardiac Na+ channel function is often observed in cardiac diseases, such as myocardial ischemia and heart failure, that are associated with increased cardiac oxidative stress. Na+ channel abnormalities can be categorized as loss- or gain-of function. Loss-of-function Na+ channel abnormality leads to reduced peak INa, resulting in conduction block that potentiates re-entrant type ventricular arrhythmias. Gain-of-function Na+ channel abnormality, in contrast, results in the increase in late INa, which can lead to prolongation of APDs, EADs, reverse mode NCX transport, and subsequent Ca2+ overload in sarcromere, all of which predispose to ventricular arrhythmias.

ROS have been demonstrated to affect Na+ channel function through multiple mechanisms. Transcriptionally, ROS have been shown to reduce Nav1.5 channel expression by reducing mRNA expression. At the protein level, ROS have been shown to impair Nav1.5 channel inactivation through direct oxidation at the methionine residues of Nav1.5 channel protein. Elevated mitochondrial ROS are known to reduce peak INa by modifying Nav1.5 conductance through post-translational mechanisms without affecting cell surface expression of Nav1.5. The slowly inactivating component of Na+ current (late INa), on the contrary, is known to be augmented by ROS in cardiac myocytes. For example, increased H2O2 has been shown to increase the open probabilities of Na+ channel, leading to increased late INa and arrhythmogenic changes including prolongation of APDs, EADs, and cytosolic Ca2+ overload (Figure 5).

In addition to direct ROS-dependent effects on Na+ channels, ROS also modulate Na+ channel activities indirectly by altering membrane lipid environment or by activating signaling molecules, such as PKC or CaMKII. Elevated mitochondrial ROS activate PKC, for example, which may reduce peak INa by affecting Nav1.5 phosphorylation. ROS-induced CaMKIIb activation, on the contrary, has been shown to mediate H2O2-induced increases in late INa.

Increased myocardial oxidative stress is often associated with impaired cardiac metabolism. The cardiac NADH level, for example, is elevated during cardiac ischemia and cardiomyopathy, which leads to increased mitochondrial ROS production and reduced peak INa. Using a mouse model of non–ischemic cardiomyopathy, we have demonstrated recently that the cytosolic NADH and mitochondrial ROS levels are increased with cardiomyopathy, resulting in a cardiac peak INa reduction without...
altering membrane Na⁺ channel protein expression levels.⁵⁷ The reduced peak I_{Na} can be restored by mitochondrial antioxidants NAD⁺ or Mito-TEMPO [(2,2,6,6-Tetramethylpiperidin-1-yl) oxyl] treatment.⁵⁷ Consistent with these findings, NAD⁺ treatment has been shown to improve myocardial conduction abnormality that is associated with reduced peak I_{Na} in human failing heart.⁵⁷

The link between mitochondrial ROS and I_{Na} regulation is also observed in the hereditary arrhythmia syndromes. One of the Brugada syndrome–causing mutations, A280V mutation in glycerol-3-phosphate dehydrogenase 1–like protein, has been demonstrated to reduce peak I_{Na} by increasing cytosolic NADH levels and mitochondrial ROS.⁵³,¹⁷⁹ Peak I_{Na} reduction by A280V glycerol-3-phosphate dehydrogenase 1–like is abrogated by the treatment of NAD⁺ or Mito-TEMPO.¹²³ Taken together, these data suggest the important role of mitochondrial ROS in transducing the altered cardiac metabolic states (NADH/NAD⁺ levels) to the regulation of cardiac Na⁺ channels and membrane excitability.

During acute ischemia, myocardial NO⁻ production is enhanced owing to increased NOS (especially eNOS) activity and conversion of tissue nitrate to NO⁻.¹⁸¹ Increased NO⁻ production and elevated NOS activities play a critical role in the cardioprotective effects of IPC (see Myocardial ROS and Potassium Channels Section of this article), which has been extensively reviewed previously.¹⁸²,¹⁸³ Elevated NO⁻ has been shown to increase late I_{Na} which can be abolished by reducing agents glutathione and DTT,¹⁸⁴,¹⁸⁵ suggesting that NO⁻ modulates Na⁺ channel activities by oxidizing Na⁺ channel or its regulatory proteins. Interestingly, others report that NO⁻ decreases peak I_{Na} through reducing the open probability and the surface expression of functional Na⁺ channels, which are mediated indirectly through a PKG/PKA-dependent pathway.¹⁸⁶ Peroxynitrite, formed by NO⁻ and O₂⁻ interaction, has also been shown to augment late I_{Na}⁵⁷

### Cardiac Redox State and Ca²⁺ Homeostasis

As mentioned earlier in this review, intracellular [Ca²⁺] is tightly regulated by various Ca²⁺-handling proteins, including voltage-gated Ca²⁺ channels, SERCA, RyR2, NCX, and signaling molecules such as CaMKII, PKA, and PKC. Many of these Ca²⁺-handling and regulatory proteins harbor methionines or thiol groups that are susceptible to the direct modification by ROS or reducing agents, thereby sensitive to the altered redox states during cardiac diseases.

In general, elevated ROS levels in cardiac myocytes are known to result in a net increase in intracellular [Ca²⁺]⁵⁸. The direct application of H₂O₂ or increased mitochondrial ROS has been shown to increase I_{CaL} and its sensitivity to isoproterenol stimulation in ventricular myocytes.¹⁸⁰ In addition, oxidized low-density lipoprotein has been demonstrated to stimulate I_{CaL} through lysophosphatidylcholine-induced mitochondrial ROS.¹⁸⁰ Nevertheless, there are conflicting reports showing reduced cardiac I_{CaL} in the presence of elevated oxidative stress.¹⁹¹,¹⁹² The discrepant findings in the effects of ROS on cardiac I_{CaL} may be explained by the differences in the experimental design, animal species, and type of ROS involved.

The results on the impact of increased oxidative stress on other Ca²⁺-handling proteins, in contrast, are more consistent. Excessive ROS increases the open probability of RyR2, enhancing the release of Ca²⁺ from SR.¹⁹³ Calcium sparks from RyR2 are increased in response to photoactivated or chemical-induced mitochondrial ROS.¹⁹⁴ In contrast to RyR2, SERCA activity is reduced on increased cardiac ROS,¹⁹⁵–¹⁹⁷ which can be, at least in part, attributed to reduced ATP supply for SERCA pump owing to mitochondrial dysfunction.³⁸ Cardiac NCX, which normally extrudes Ca²⁺ from cytosol in exchange for Na⁺, has been shown to be activated in the reverse mode by increased oxidative stress, thereby increasing cytosolic [Ca²⁺] (Table 2; Figure 5).¹⁹⁸,¹⁹⁹

CaMKII is activated by ROS, and it is known to play a critical role in regulating calcium-handling proteins in response to increased cardiac oxidative stress.³⁹ For example, activated CaMKII augments I_{CaL} by phosphorylating Cav1.2 channel subunit and increasing its open probability.²⁰¹ Phosphorylation of RyR2 by CaMKII increases SR Ca²⁺ leak, promoting cytosolic Ca²⁺ overload and delayed afterdepolarizations.²⁰² Taken together, the net impact of increased oxidative stress
on Ca²⁺-handling proteins leads to cytosolic Ca²⁺ overload and depletion of SR Ca²⁺ store, resulting in detrimental changes, such as arrhythmogenic delayed afterdepolarizations and contractile dysfunction.

As discussed earlier in this review, mitochondria also play an important role in the regulation of Ca²⁺ in cardiac myocytes. Increased oxidative stress is known to modulate mitochondrial [Ca²⁺] by altering mitochondrial ion channel activities, contributing to the perturbation of cytosolic Ca²⁺ homeostasis. For example, increased ROS during myocardial ischemia depolarize ∆Ψm,202 forcing mitochondrial NCX into reverse mode and drives Ca²⁺ from cytosol into mitochondria.117,203 Physiologically, mitochondrial Ca²⁺ is required for normal ETC function and serves as a positive effector of OXPHOS. Ca²⁺ overload, however, leads to increased mitochondrial ROS production through mechanisms including increased electron leakage,204,205 enhanced NO⁺ production, which is known to inhibit complex IV and augment ROS production from complex III,206 and reduced cytochrome c-mediated respiration.207,208 In addition, pathological mitochondrial Ca²⁺ overload has been shown to increase NO⁺ and ROS production in cardiac myocytes.209 These data suggest a positive feedback loop between ROS-induced Ca²⁺-overload and Ca²⁺-induced ROS production. Under pathological conditions, such as myocardial ischemia, cellular and mitochondrial [Ca²⁺] increase, leading to increased ROS production. ROS overproduction induced by elevated mitochondrial [Ca²⁺] results in further increase in mitochondrial Ca²⁺ and ROS levels. The Ca²⁺ load and ROS produced from this positive feedback loop can overwhelm the cellular capacity of ROS scavenging and Ca²⁺ clearance, resulting in cellular damage and electric instability that predispose to ventricular arrhythmias and SCD.120

Increased oxidative stress is known to inhibit repolarizing Kv currents including Ito and several delayed rectifier K⁺ currents (Ikr, Ik, and Ic) in mammalian ventricular myocytes.210,211 These effects are reversible by raising cellular antioxidant glutathione levels.140,219 ROS have been shown to regulate Kv current expression by reducing the transcript/protein expression of the Kv channel subunits218 and by modulating the phosphorylation of these channel subunits through PKs such as PKA, PKC, or protein tyrosine phosphatases.220–222 Cardiac Kv channels are also regulated by NO⁺. It has been reported that NO⁺ inhibits human cardiac Kv4.3 channel, thereby reducing the transient outward K⁺ current Ito.221 NO⁺ also blocks Kv1.5 channel through S-nitrosylation and the activation of cGMP/PKG pathway.223 Myocardial ROS also regulates myocyte membrane excitability through sarKATP channels. During myocardial ischemia, fuel substrate deprivation and increased oxidative stress depolarize the mitochondrial network, leading to fluctuated on Ca²⁺-handling proteins.

Table 2. Effects of Increased Oxidative Stress on Cardiac Ion Channel/Transporter Function and Arrhythmogenicity

<table>
<thead>
<tr>
<th>Channel/Transporter</th>
<th>Effects of ROS</th>
<th>Effects on Electric/Ionic Homeostasis</th>
<th>Proarrhythmic Mechanism</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peak INa</td>
<td>↓</td>
<td>↓ Na⁺ influx</td>
<td>Slow conduction</td>
</tr>
<tr>
<td>Late INa</td>
<td>↑</td>
<td>↑ Na⁺ influx, APD prolonged</td>
<td>EAD</td>
</tr>
<tr>
<td>Ito</td>
<td>↓</td>
<td>↓ K⁺ influx, APD prolonged</td>
<td>EAD</td>
</tr>
<tr>
<td>Ic</td>
<td>↓</td>
<td>↓ K⁺ influx, APD prolonged</td>
<td>EAD</td>
</tr>
<tr>
<td>Ic</td>
<td>↓</td>
<td>↑ K⁺ influx, APD prolonged</td>
<td>EAD</td>
</tr>
<tr>
<td>Ic</td>
<td>↑</td>
<td>↑ K⁺ influx, APD shortened</td>
<td>Current sink, slow conduction</td>
</tr>
<tr>
<td>NCX</td>
<td>Reverse mode</td>
<td>Ca²⁺ overload</td>
<td>DAD</td>
</tr>
<tr>
<td>Cx43</td>
<td>↓</td>
<td>Impaired gap junction function</td>
<td>Slow conduction</td>
</tr>
<tr>
<td>SERCA</td>
<td>↑</td>
<td>Ca²⁺ overload</td>
<td>DAD</td>
</tr>
<tr>
<td>RyR2</td>
<td>↑</td>
<td>Ca²⁺ overload</td>
<td>DAD</td>
</tr>
<tr>
<td>Mito-NCX</td>
<td>Reverse mode</td>
<td>Ca²⁺ overload</td>
<td>DAD</td>
</tr>
<tr>
<td>Mito-KATP</td>
<td>↑</td>
<td>↑ Mitochondrial K⁺ influx</td>
<td>Protective, ischemic-preconditioning</td>
</tr>
</tbody>
</table>

APD indicates action potential duration; Cx43, connexin 43; DAD, delayed afterdepolarization; EAD, early afterdepolarization; Ito, delayed rectifier K⁺ current; Ikr, inwardly rectifying K⁺ current; Ic, ATP-sensitive K⁺ current; Ic, transient outward K⁺ current; Late INa, late Na⁺ current; mito-KATP, mitochondrial ATP-sensitive K⁺ current; mito-NCX, mitochondrial Na⁺/Ca²⁺ exchanger; NCX, Na⁺/Ca²⁺ exchanger; Peak INa, peak Na⁺ current; RyR2, ryanodine receptor 2; and SERCA, sarco/endoplasmic reticulum Ca²⁺-ATPase.

Myocardial ROS also favors ROS production (by impairing mitochondrial energetics and oxidizing NAD(P)H).119,120 NO⁺ and peroxynitrite also play important roles in modulating cardiac Ca²⁺ channel and Ca²⁺-handling proteins. Increased NO⁺ inhibits L-type Ca²⁺ channel activity through increased S-nitrosylation211,212 and cGMP-PKG-dependent phosphorylation13 of Ca²⁺ channel subunits, which may contribute to the cardioprotective effects of IPC by limiting Ca²⁺ influx during ischemia.212 The effects of NO⁺ on RyR2 is concentration dependent. Physiological NO⁺ levels (∼1 μmol/L) do not affect RyR2 activity, whereas supraphysiological levels (≥100 μmol/L) inhibit RyR2 activity.214 Peroxynitrite, on the contrary, activates RyR2 by oxidizing its cysteine residues.214 SERCA2a, by contrast, is inhibited by peroxynitrite-mediated tyrosine nitration. PKA pretreatment can prevent the inhibitory effects of peroxynitrite on SERCA2a by inducing dissociation of phospholamban from SERCA2a.215

Myocardial ROS and Potassium Channels

Increased oxidative stress is known to inhibit repolarizing Kv currents including Ito and several delayed rectifier K⁺ currents (Ikr, Ik, and Ic) in mammalian ventricular myocytes.210,211 These effects are reversible by raising cellular antioxidant glutathione levels.140,219 ROS have been shown to regulate Kv current expression by reducing the transcript/protein expression of the Kv channel subunits218 and by modulating the phosphorylation of these channel subunits through PKs such as PKA, PKC, or protein tyrosine phosphatases.220–222 Cardiac Kv channels are also regulated by NO⁺. It has been reported that NO⁺ inhibits human cardiac Kv4.3 channel, thereby reducing the transient outward K⁺ current Ito.221 NO⁺ also blocks Kv1.5 channel through S-nitrosylation and the activation of cGMP/PKG pathway.223
\( \Delta V_{m} \) and ATP production levels, which result in oscillation of sarc\( \mathrm{Ca}^{2+} \) currents and ADP. The \( \Delta V_{m} \) depolarization of the mitochondrial network is potentiated by widespread ROS production induced by focal increases in mitochondrial ROS (ROS-induced ROS release). IMAC plays a critical role in \( \Delta V_{m} \) depolarization on increased ROS; pharmacological inhibition of IMAC has been shown to prevent \( \Delta V_{m} \) depolarization and sarc\( \mathrm{Ca}^{2+} \) currents and ADP, as well as preventing ventricular arrhythmias in mammalian hearts during myocardial ischemia.

Another group of K\textsubscript{ATP} channels, residing on the mitochondrial inner membrane (mito-K\textsubscript{ATP} channels), contribute importantly to the protective effects of IPC, an endogenous cellular protective mechanism involving brief periods of ischemia that confers protection against infarction produced by a subsequent prolonged ischemia. Under physiological conditions, mito-K\textsubscript{ATP} channel opening enhances mitochondrial ROS production, triggering downstream signaling pathways involved in gene transcription and cell growth. During IPC, the activation of mito-K\textsubscript{ATP} channels allows mitochondrial K\textsuperscript{+} influx, resulting in partially depolarized \( \Delta V_{m} \), which leads to a compensatory increase in proton pumping and cellular respiration to maintain \( \Delta \Psi \). IPC,238 and several mito-K\textsubscript{ATP} channel openers are shown to potentiate mito-K\textsubscript{ATP} opening blunts mitochondrial \( \mathrm{Ca}^{2+} \) accumulation during ischemia and reduces PTP opening on reperfusion, both of which contribute to the protection against cell death by IPC. The opening of mito-K\textsubscript{ATP} channel also triggers the production of protective ROS, which activates a PKC-dependent pathway that confers cardiac protection by reducing deleterious postischemic ROS production. Interestingly, NO\textsuperscript{−}NO\textsuperscript{•} is an important free radical, has been shown to potentiate mito-K\textsubscript{ATP} opening and triggers both early and delayed IPC. In addition, mito-K\textsubscript{ATP} opening during ischemia provides additional K\textsuperscript{+} influx to maintain mitochondrial volume, which is critical to maintain the normal function of mitochondria and ETC. Pharmacological inhibition of mito-K\textsubscript{ATP} channels abrogates the antiarrhythmic effects of IPC, and several mito-K\textsubscript{ATP} channel openers are shown to provide protective effects against ischemia-induced cardiac arrhythmias.

**Mitochondrial ROS and Cardiac Gap Junction Regulation**

Cardiac gap junctions are formed by the assembly of a pair of juxtaposed intercellular hemichannels with each hemichannel consisting of 6 connexin (connexin) proteins. Gap junctions mediate the intercellular communication of small metabolites and ions and play an essential role in cardiac electric conduction. Among the 3 principal connexin isoforms (connexin 40, connexin 43, and connexin 45) expressed in the heart, connexin 43 is the predominant isoform expressed in ventricular myocytes. Ventricular connexin 43 expression has been shown to be downregulated with myocardial ischemia, and heart failure, which can lead to slowed conduction, increased electric heterogeneity and abnormal anisotropic properties of the ventricles. These changes can facilitate the initiation and maintenance of ventricular arrhythmias and SCDs.

Cardiac renin–angiotensin system (RAS) activity is increased on acute ischemia, contributing to the acute and chronic myocardial remodeling in response to ischemia. RAS activation is known to increase myocardial oxidative stress and downregulate ventricular gap junction protein connexin 43. Transgenic mouse models with enhanced cardiac RAS activity have high incidence of conduction abnormality, ventricular arrhythmias and sudden death owing to reduced cardiac connexin 43 and abnormal gap junction function. Using a transgenic mouse model of cardiac-restricted angiotensin-converting enzyme overexpression (ACE8/8), we have demonstrated that increased cardiac RAS activity leads to increased activity of cSrc, a redox-sensitive tyrosine kinase, by increasing cSrc phosphorylation at Tyr\textsuperscript{416}, in the ventricular myocardium. The activation of cSrc leads to connexin 43 downregulation, impaired gap junction function, slowed cardiac conduction, and increased incidence of ventricular arrhythmias and SCD. The downregulation of connexin 43 and increased risk for arrhythmias in angiotensin-converting enzyme 8/8 mice are alleviated by pharmacological inhibition of RAS and cSrc. It has been shown that increased myocardial p-cSrc leads to connexin 43 reduction through the competition between p-cSrc and connexin 43 for the binding with zonula occludens-1, a scaffolding protein at the intercalated disk, resulting in connexin 43 destabilization and degradation. Elevated p-cSrc levels also impair gap junction function by phosphorylating connexin 43 at tyrosine residues. Using the same ACE 8/8 mouse model, we have also shown that cardiac mitochondrial ROS were markedly increased with enhanced RAS activity. Treatment with mitochondria-targeted antioxidant Mito-TEMPO, but not with other types of antioxidants, decreases cSrc phosphorylation, preserves connexin 43 expression, improves gap junction function, and abolishes ventricular arrhythmias and SCD in ACE 8/8 mice. Although the clinical evidence suggests that exogenous antioxidants do not prevent SCD, these data support a role for ROS, and suppression of endogenous, mitochondria-specific ROS may be an effective therapeutic approach. Mechanistically, we have recently demonstrated that enhanced RAS signaling increases S-nitrosylation of cardiac caveolin-1 (Cav1), an intrinsic inhibitor of cSrc, resulting in Cav1-cSrc dissociation and subsequent cSrc activation. Cav1 S-nitrosylation upon enhanced RAS signaling is mediated by increased Cav1-eNOS binding that is dependent on elevated mitochondrial ROS. Consistent with these findings, Cav1 knockout mice exhibit increased cSrc activation, reduced connexin 43 expression, myocardial conduction defect, and increased inducibility of ventricular arrhythmias. Taken together, these data suggest the critical roles of mitochondrial oxidative stress and Cav1 in AngII–induced gap junction remodeling and arrhythmia. As mitochondrial ROS are increased in myocardial ischemia and heart failure, both of which are associated with RAS activation, reduced ventricular connexin 43 and increased risk for ventricular arrhythmias and SCD, it would be of great interest to test whether the treatment with mitochondria-targeted antioxidant...
Additional studies are required to address these possibilities. Changes and simply occur on an arrhythmogenic background. Metabolic/oxidative changes or whether acute changes are unrelated to the chronic metabolic/oxidative derangement. It is difficult to distinguish whether acute rapidly reversed by targeting the underlying metabolic/oxidative stress in these chronic conditions, which can be suggest the contribution of the acute, modifiable metabolic/oxidative changes related to these chronic conditions involve electric remodeling15,21 and the creation of arrhythmogenic substrates such as fibrosis,266,267 which may not be seen with acute metabolic and oxidative stress. Nevertheless, the effects of acute and chronic metabolic/oxidative stress on cardiac electric function may not be mutually exclusive. The acute application of mitochondria-targeted antioxidant, for example, can restore the reduced peak INa observed with chronic heart failure.57 In addition, reduced Kv currents in diabetic heart can be reversed by acute insulin treatment.140 These observations suggest the contribution of the acute, modifiable metabolic/oxidative stress in these chronic conditions, which can be rapidly reversed by targeting the underlying metabolic/oxidative derangement. It is difficult to distinguish whether acute metabolic/oxidative changes exacerbate and amplify chronic changes or whether acute changes are unrelated to the chronic changes and simply occur on an arrhythmogenic background. Additional studies are required to address these possibilities.

Conclusions
In summary, metabolic derangement and increased oxidative stress are prevalent in arrhythmogenic cardiac conditions, particularly during myocardial ischemia. Impaired cardiac metabolism and increased ROS production can lead to malfunction of various cellular mechanisms that are required to maintain normal electric functioning and intracellular ionic homeostasis in cardiac myocytes. The impact of altered cardiac metabolism and increased oxidative stress on cardiac arrhythmogenicity is summarized in Table 1, Figure 4 and Table 2, Figure 5, respectively. Because the conventional antiarrhythmic drugs targeting ion channels are often proarrhythmic, understanding the mechanisms linking abnormal metabolism and oxidative stress to cardiac arrhythmias may help to develop novel therapeutics to reduce the risk of life-threatening arrhythmias and SCD in patients with cardiac diseases. These observations suggest that therapeutics tailored to ameliorate metabolic derangement and oxidative stress may prove a more efficacious alternative to traditional ion channel blocking drugs to address arrhythmia in associated with cardiac diseases, and a list of potential novel therapeutics are summarized in Table 3.

Sources of Funding
This work was funded by National Institutes of Health grants RO1 HL104025 (Dr Dudley), HL106592 (Dr Dudley), a Veterans Affairs MERIT grants BX000859 (Dr Dudley), RO1 HL 122109 (J.C. Makielski), RO1 HL 57414 (J.C. Makielski), RO1 HL 071092 (J.C. Makielski), a Taiwan Ministry of Science Technology Grant 103-2320-B-002-068-MY2 (K.-C. Yang) and a Taiwan National Health Research Institute Career Development grant NHRI-EX104-104185C (K.-C. Yang).

Disclosures
Dr Dudley is an inventor of 13/551 790 A Method for Ameliorating or Preventing Arrhythmic Risk Associated with Cardiomyopathy by Improving Conduction Velocity, 13/507 319 A Method for Modulating or Controlling Connexin43 (Cx43) Level of a Cell and Reducing Arrhythmic Risk, PCT/US2008/011919 Modulation of Sodium Current by Nicotinamide Adenine Dinucleotide, US 12/929 786 Modulating Mitochondrial Reactive Oxygen Species to Increase Cardiac Sodium Channel Current and Mitigate Sudden Death, and 13/551 790 Method for Ameliorating or Preventing Arrhythmic Risk Associated with Cardiomyopathy.


Elevated cytosolic Na + decreases mitochondrial Ca 2+ uptake during excitation-contraction coupling and impairs energetic adaptation of cardiac myocytes. Circ Res. 2006;99:172–182. doi: 10.1161/01.RES.0000232546.92777.05.

10.1161/01.RES.0000232546.92777.05


Yang et al. Oxidants, Metabolism, and Sudden Cardiac Death


Mechanisms of Sudden Cardiac Death: Oxidants and Metabolism
Kai-Chien Yang, John W. Kyle, Jonathan C. Makielski and Samuel C. Dudley, Jr

doi: 10.1161/CIRCRESAHA.116.304691

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circres.ahajournals.org/content/116/12/1937

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Circulation Research can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Circulation Research is online at:
http://circres.ahajournals.org//subscriptions/