Mutations in STAP1 Are Associated With Autosomal Dominant Hypercholesterolemia

Sigrid W. Fouchier, Geesje M. Dallinga-Thie, Joost C.M. Meijers, Noam Zelcer, John J.P. Kastelein, Joep C. Defesche, G. Kees Hovingh

Rationale: Autosomal-dominant hypercholesterolemia (ADH) is characterized by elevated low-density lipoprotein cholesterol levels and increased risk for coronary vascular disease. ADH is caused by mutations in the low-density lipoprotein receptor, apolipoprotein B, or proprotein convertase subtilisin/kexin 9. A number of patients, however, suffer from familial hypercholesterolemia 4 (FH4), defined as ADH in absence of mutations in these genes and thereafter use the abbreviation FH4.

Objective: To identify a fourth locus associated with ADH.

Methods and Results: Parametric linkage analysis combined with exome sequencing in a FH4 family resulted in the identification of the variant p.Glu97Asp in signal transducing adaptor family member 1 (STAP1), encoding signal transducing adaptor family member 1. Sanger sequencing of STAP1 in 400 additional unrelated FH4 probands identified a second p.Glu97Asp carrier and 3 additional missense variants, p.Leu69Ser, p.Ile71Thr, and p.Asp207Asn. STAP1 carriers (n=40) showed significantly higher plasma total cholesterol and low-density lipoprotein cholesterol levels compared with nonaffected relatives (n=91).

Conclusions: We mapped a novel ADH locus at 4p13 and identified 4 variants in STAP1 that associate with ADH. (Circ Res. 2014;115:552-555.)

Key Words: hypercholesterolemia, autosomal dominant ■ physical chromosome mapping ■ STAP1 gene, human

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steps (Online Table I) of the exome sequence data led to the identification of a variant in \textit{STAP1}, p.Glu97Asp (c.291G>C). This variant, which was verified by Sanger sequencing, affected a highly conserved amino acid and was predicted to be pathogenic by SIFT, Polyphen-2, and MutationTaster. The variant was absent in 400 controls of matched ancestry, the Genome of the Netherlands (http://www.nlgenome.nl/), the 1000G population, and the National Heart, Lung, and Blood Institute-Exome Sequencing Project data set (http://evs.gs.washington.edu/EVS/), suggesting that it is not a polymorphism. Testing of remaining family members by Sanger sequencing revealed 11 affected carriers (total cholesterol or LDL-C levels >95th percentile), 4 possibly affected carriers (between 75th and 95th percentile) and 1 unaffected carrier (<75th percentile), reflecting a disease penetrance of 0.94 (Figure).

We subsequently sequenced all coding regions of \textit{STAP1} in 400 unrelated FH4 probands (Online Table II) and identified a second p.Glu97Asp carrier and 3 additional missense variants, p.Leu69Ser (c.206T>C), p.Ile71Thr (c.212T>C), and p.Asp207Asn (c.619G>A). All variants were located in exon 3, except for p.Asp207Asn located in exon 6, and all variants altered highly conserved amino acid residues. SIFT, Polyphen-2, and MutationTaster predicted the p.Leu69Ser variant to be pathogenic, whereas p.Ile71Thr and p.Asp207Asn were predicted to possibly affect protein function. None of these variants were detected in 400 controls, the 1000G population, or in the Genome of the Netherlands. In the National Heart, Lung, and Blood Institute-Exome Sequencing Project data set, p.Ile71Thr was found once and p.Asp207Asn was found 3×. However, variants at low frequency associated with an hypercholesterolemic phenotype can be expected, considering the substantial number of patients at increased cardiovascular risk included in National Heart, Lung, and Blood Institute-Exome Sequencing Project.

The frequency of rare \textit{STAP1} variants predicted to be pathogenic or possibly pathogenic in our FH4 cohort was 1.3% (5 of 400 individuals). This was 3× and 6× higher than the rate observed in National Heart, Lung, and Blood Institute-Exome Sequencing Project (0.4%, 17 of 4300 European American individuals).

Figure. Family FH4-0102. Half-blackened symbols indicate affected members (>95th percentile), quarter-blackened symbols indicate possibly affected members (between 75th and 95th percentile), and white symbols indicate unaffected members (<75th percentile). A dot indicates carriers of the p.Glu97Asp variant. The triangle represents the proband. Age (in years) at lipid measurement, total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol (HDL-C), triglycerides (TG) in mmol/L, and history and age onset of coronary vascular disease are given. Percentiles matched for age and sex, pTC, pLDL, pHDL, and pTG are given between brackets, a percentile indicated by 99 resembles a >95th percentile. Individuals with mapping data are marked with +. For individuals with exome data symbols are marked with brackets. AMI indicates acute myocardial infarction; CABG, coronary artery bypass graft; CVD, coronary vascular disease; and UPN, unique personal number.
individuals; \( P = 0.035 \) and the Genome of the Netherlands (0.2%, 1 of 499 individuals; \( P = 0.067 \), respectively (Online Table III).

The STAP1 variants found in the 4 additional probands were further evaluated in the relatives of the replicate families. In total, 45 carriers of the rare STAP1 variants and 91 noncarrier relatives were identified. Individuals carrying a rare STAP1 variant had significantly higher total cholesterol and LDL-C levels compared with relatives without the STAP1 variants (Table). High-density lipoprotein cholesterol levels were not significantly different, whereas triglycerides levels were slightly increased. Similar results were observed when the carriers of the discovery family were excluded (Online Table IV).

We additionally compared the ADH phenotype of the STAP1 carriers with the ADH phenotype observed in a large cohort of LDLR and APOB mutation carriers. Total cholesterol and LDL-C levels were significantly lower in STAP1 carriers when compared with patients molecularly diagnosed with familial hypercholesterolemia (FH, OMIM 606945), caused by mutations in LDLR. However, cholesterol levels in STAP1 carriers were comparable with those observed in patients molecularly diagnosed with familial defective apolipoproteinemia B (OMIM 107730), caused by mutations in APOB.

In studies looking at ADH, individuals are considered affected if their plasma cholesterol levels are >95th percentile adjusted for age and sex, and in ADH-affected families a disease penetrance of 0.9 is usually assumed. Nevertheless, the ADH phenotype of patients affected by well-established pathogenic mutations in the LDLR and APOB varies enormously from severe to moderate, as recently demonstrated.7,8 In the 5 STAP1 families identified here, such variation was also observed, most evident in family FH4-0150 (Online Figure IIA), which displayed only a moderate penetrance of 0.76. The variants p.Ile71Thr in family FH4-0247 (Online Figure IIB) and p.Glu97Asp in family FH4-317 (Online Figure IIC) displayed complete penetrance. Further expansion of family FH4-0356 was unfortunately not possible (Online Figure IID). Overall, the disease penetrance of the missense STAP1 variants in all 5 families is 0.94±0.10 with a phenocopy rate of 0.19±0.11.

### Discussion

We report here for the first time association of rare STAP1 variants with increased total cholesterol and LDL-C levels. The lipid phenotype of STAP1 carriers is not as severe as generally observed in FH patients, but resembles the minor lipid and coronary vascular disease phenotype observed in familial defective apolipoproteinemia B patients.4 It is of note that in genome-wide association studies, single nucleotide polymorphism in the STAP1 locus have not been shown to be associated with cholesterol levels. However, single nucleotide polymorphisms in the STAP1 locus have been associated with the risk for Parkinson disease.10 Although the exact biological explanation for this association remains elusive, increased cholesterol levels might play a role because high total serum cholesterol levels have been associated with a slow clinical progression of Parkinson disease.11

The function of STAP1, also known as BRDG1 (BCR downstream signaling protein 1) or stem cell adhesion protein 1, is largely unknown. STAP1 contains a Pleckstrin homology domain, a Src homology 2 domain, and several tyrosine phosphorylation sites.12 It has been suggested that the Pleckstrin homology domain of STAP1 functions as a phosphoinositide-binding domain and facilitates the association of STAP1 with membranes.12 Intriguingly, the p.Leu69Ser, p.Ile71Thr, and p.Glu97Asp alterations all occur within the Pleckstrin homology domain of STAP1 and may thus affect its interaction with membranes or membrane proteins (Online Figure III). The STAP1 p.Asp207Asn variant is located in the Src homology 2 domain, a domain implicated in propagation of signal transduction emanating from upstream receptor tyrosine kinase pathways.13 Indeed, STAP1 has been suggested to act downstream of the receptor tyrosine kinases c-kit and c-fms.12

In line with STAP1 functioning downstream of c-kit to control systemic cholesterol levels, it is interesting to point out that W/Wv mice, which harbor a loss-of-function mutation in c-kit, show increased plasma cholesterol levels.14,15 Similarly, gain-of-function mutations in c-kit cause leukemia,16 and leukemic patients often present with hypercholesterolemia.17–21 Intriguingly, therapeutic administration of receptor tyrosine kinase inhibitors to these patients is associated with increases in plasma cholesterol.22,23 This raises the possibility that STAP1 and potentially other downstream genes in this signaling pathway have an effect on circulating levels of cholesterol.

Our study provides strong genetic evidence for involvement of STAP1 in controlling cholesterol homeostasis, yet the molecular mechanism behind this is presently unknown. Expression of STAP1 in the liver, the main organ maintaining whole-body cholesterol homeostasis, is low (not shown). Studies aimed at identifying the extrahepatic mechanism underlying the potential

### Table. Clinical Characteristics of Carriers and Noncarriers

<table>
<thead>
<tr>
<th></th>
<th>STAP1 Carriers, mean [SD]</th>
<th>STAP1 Noncarriers, mean [SD]</th>
<th>PValue</th>
<th>PValue n</th>
<th>LDLR Carriers, mean [SD]</th>
<th>STAP1 vs LDLR, PValue</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex (men), %</td>
<td>91</td>
<td>43</td>
<td>40</td>
<td>50</td>
<td>2471</td>
<td>49</td>
</tr>
<tr>
<td>TC, mmol/L</td>
<td>80</td>
<td>5.76 [1.25]</td>
<td>40</td>
<td>6.63 [1.66]</td>
<td>0.001</td>
<td>2471</td>
</tr>
<tr>
<td>LDL, mmol/L</td>
<td>89</td>
<td>3.83 [1.10]</td>
<td>35</td>
<td>4.27 [1.24]</td>
<td>0.005</td>
<td>1963</td>
</tr>
<tr>
<td>HDL, mmol/L</td>
<td>78</td>
<td>1.37 [0.37]</td>
<td>36</td>
<td>1.33 [0.28]</td>
<td>0.949</td>
<td>1970</td>
</tr>
<tr>
<td>TG, mmol/L</td>
<td>78</td>
<td>1.41 [0.79]</td>
<td>37</td>
<td>1.85 [1.32]</td>
<td>0.052</td>
<td>1969</td>
</tr>
</tbody>
</table>

*All family members, including branches in which the STAP1 variants were not present.

APOB indicates apolipoprotein B; HDL, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol; LDLR, low-density lipoprotein receptor; STAP1, signal transducing adaptor family member 1; TC, total cholesterol; and TG, triglycerides.
role of STAP1 in cholesterol metabolism are required and are expected to yield new insight into the pathogenesis of ADH.

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Disclosures

None.

References


Novelty and Significance

What Is Known?

• Autosomal-dominant hypercholesterolemia (ADH) leads to an increase in total and low-density lipoprotein cholesterol and accelerated atherosclerotic disease.

• Mutations in the genes encoding low-density lipoprotein receptor, apolipoprotein B, or proprotein convertase subtilisin/kevin 9 are established causes of ADH.

• A substantial proportion of clinically diagnosed hypercholesterolemic patients do not carry mutations in the 3 established causal genes for ADH.

• The identification of novel ADH genes is essential to identifying additional targets to lower low-density lipoprotein cholesterol to reduce coronary vascular disease risk.

What New Information Does This Article Contribute?

• We identified signal transducing adaptor family member 1 (STAP1) as a novel gene associated with ADH.

• This finding reveals a previously unrecognized signaling pathway that contributes to the pathogenesis of ADH.

• Identification of STAP1 as an ADH gene might expand current genetic strategies for the molecular diagnosis of ADH patients and lead to new therapeutic targets for ADH.

The prevalence of ADH is ≈1:200 individuals in most Western countries. The diagnosis is usually made on the basis of clinical symptoms. However, because these symptoms develop late in life, establishing the diagnosis in younger patients is often difficult. Genetic analysis, that is, the demonstration of a causative mutation, provides unequivocal diagnosis. A proportion of the patients with a clinical diagnosis of ADH does not carry mutations in the established ADH genes but might carry mutations in genes that have not been associated previously with the hypercholesterolemic phenotype. Here, we identify STAP1 as a novel ADH gene, a finding of prognostic value that could also yield new insight into the pathogenesis of ADH.
Mutations in \textit{STAP1} Are Associated With Autosomal Dominant Hypercholesterolemia

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SUPPLEMENTAL MATERIAL

DETAILED METHODS

Family recruitment

The probands were clinically diagnosed with familial hypercholesterolemia using an uniform protocol and internationally accepted criteria\(^1,2\) and were screened for the presence of mutations in \(LDLR\), \(APOB\) (amino acids 2955 to 3092, 3303 to 3492, 3441 to 3615 and 4330 to 4440), and \(PCSK9\) as described before.\(^3,4\)

Relatives were included into the study after written informed consent. The study protocol was approved by AMC's institutional review board (MEC 07/331).

After an overnight fast, blood was sampled and plasma levels of total cholesterol, HDL-cholesterol and triglycerides were measured by commercially available kits (Diasys, Holzheim, Germany and Randox, Northern Ireland, United Kingdom). LDL-c levels were calculated by the Friedewald formula only when triglycerides were below 4.5 mmol/L.\(^5\)

An affected ADH phenotype was defined by levels of TC or LDL-c above the 95\(^{th}\) percentile for age and gender.\(^6\) A possible ADH phenotype was defined by levels of TC or LDL-c between the 75\(^{th}\) and 95\(^{th}\) percentile for age and gender.\(^7,8\) When untreated values were not available, estimated baseline LDL-c values were back calculated based on the potency of cholesterol-lowering therapy.\(^9\)

Linkage analysis

Genomic DNA was extracted from 10 ml whole blood on an AutopureLS apparatus according to a protocol provided by the manufacturer (Gentra Systems, Minneapolis, MI, USA).

Fifteen individuals from Family FH4-0102 (III:2, III:3, III:4, III:5, III:7, III:8, III:9, III:11, III:12, III:13, IV:1, IV:3, IV:4, IV:10, IV:11) were genotyped using the HumanCytoSNP-12 SNP array (Illumina). To verify the relationship between individuals, the data were subjected to the PedCheck program.\(^10\) Multi-point linkage analysis was performed using SNPs with a spacing of 0.1 cM, computed in overlapping sets of 175 and 200 markers. Multi-point linkage analysis was first performed based on a model defined by a disease allele frequency of 0.001 and an autosomal dominant mode of inheritance with the assumption of 90% penetrance and 10% phenocopy rate using Genehunter\(^11\) and Allegro\(^12\). The Allegro haplotypes were further studied in detail using HaploPainter\(^13\) to identify phenocopies, non-penetrants and shared haplotypes.

The high penetrance of affected individuals in the 3\(^{rd}\) generation of family FH4-0102 was unexpected and discordant with the expected Mendelian inheritance pattern. This suggested that two different ADH traits might be present in this family. Therefore, the three phenocopies identified in this generation were excluded and parametric LOD scores were re-calculated based on a model defined by a disease allele frequency of 0.001 and an autosomal dominant mode of inheritance with the assumption of 100% penetrance and 0% phenocopy using Genehunter.\(^11\)

Exome sequencing

Whole exome sequencing was performed at the Broad Institute (Boston, USA). Briefly, genomic DNA underwent library construction and in-solution hybrid selection as previously described\(^14\) using an Agilent (Agilent Technologies, Santa Clara, CA, USA) whole exome kit targeting 33Mb of genomic sequence. The target includes 188,260 exons from 18,560 genes. The resulting exonic DNA was sequenced using 75-base pair paired-end reads on an Illumina GA-II sequencer.\(^14\) Raw sequence reads were aligned to the human genome reference (HG19) using the Burroughs-Wheeler Alignment tool\(^15\) in paired-end mode. The aligned reads underwent base quality recalibration using the Genome Analysis Toolkit (GATK).\(^16,17\) Duplicated reads from sequencing the same DNA molecule and sequencing reads not corresponding to the exome target were discarded.

Supplementary note
Single nucleotide substitutions and short insertion and deletion events were identified and genotyped using the GATK UnifiedGenotyper tool in multisample mode. The quality scores of the resulting variants were recalibrated using the GATK Variant Score Recalibration.

Cohorts
By the participation of 64 Lipid Clinics, evenly distributed across The Netherlands, a representative group of over ~17,000 clinically diagnosed ADH patients has been collected. All participants gave written informed consent. From this cohort 400 unrelated cases, without any mutation in LDLR, APOB (amino acids 2955 to 3092, 3303 to 3492, 3441 to 3615 and 4330 to 4440) and PCSK9 and with baseline TC and LDL-c levels above the 95th percentile for age and gender, were selected for further study.

The control cohort of ancestry-matched individuals and the cohorts of APOB and LDLR mutation carriers were recruited via the national genetic cascade screening program for FH. The control cohort consisted of 400 unrelated and untreated individuals, with baseline LDL-c levels below the 20th percentile for age and gender and tested negative for the familial mutation. The cohorts of APOB and LDLR mutation carriers consisted of related and unrelated individuals tested positive for the familial mutation. Probands were excluded. When baseline values of the APOB and LDLR carriers were not available, estimated baseline LDL values were back calculated based on the potency of cholesterol-lowering therapy. All subjects gave written informed consent for genetic screening. Lipid profiles were routinely measured with the LDX analyzer.

The National Heart, Lung, and Blood Institute (NHLBI) Exome Sequencing Project (ESP) and the Genome of the Netherlands (GoNL) datasets were used to evaluate the prevalence of individuals carrying rare STAP1 variants.

Genetic analysis of STAP1
The nine exons, including the intronic boundaries, of STAP1 were analyzed by Sanger sequencing. For mutation nomenclature, numbering was based on the cDNA with nucleotide c.1 being A of the ATG initiation codon p.1. The STAP1 reference sequence NM_000384.2 was used.

A variant predicted to be tolerated by SIFT, benign by Polyphen-2 and a polymorphism by MutationTaster was considered to be non-pathogenic. A variant predicted to be deleterious by SIFT, Polyphen-2 and MutationTaster was considered to be pathogenic. All other variants were classified as possible pathogenic.

Phenocopy rate was determined by the count of non-carriers with an affected status divided by the count of all non-carriers in each family. Penetrance rate was determined by the count of carriers with an affected status divided by the count of all carriers in each family.

Statistical analysis
SPSS 20 software (SPSS, Inc, Chicago, Illinois, USA) was used for data analysis. Differences in cholesterol levels corrected for age, gender, and relatedness were determined using a general linear mixed model analysis. Triglyceride levels values were log-transformed before analysis. Probands were excluded from the analysis. A p-value <0.05 was considered statistically significant.

Differences in the frequency of rare STAP1 variants between the Dutch FH4 cohort, GoNL and EVS-EA were determined using an one-tailed Fisher’s exact test (GraphPad Software, La Jolla, California, USA).

References


### Online Table I. Variant-filtering pipeline for family FH4-0102

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<tr>
<th>criteria</th>
<th># exome variants</th>
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<td>Variants fitting a dominant model located in the linked intervals on</td>
<td>462</td>
</tr>
<tr>
<td>chromosome 4, 11 and 13</td>
<td></td>
</tr>
<tr>
<td>Variants with frequency &lt;0.5% (EVS and GoNL)</td>
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<tr>
<td>Variants not found in control exomes (GQ&gt;90)</td>
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<tr>
<td>Not located in a gene with nonsense mutations with frequency &gt;0.5% (stop-</td>
<td>5</td>
</tr>
<tr>
<td>gained, frame-shift, and splice-3/5 in EVS and GoNL)</td>
<td></td>
</tr>
<tr>
<td>Not involving 1) non-coding variants predicted not to affect splicing</td>
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</tr>
<tr>
<td>and low phastcons (&lt;0.5), 2) synonymous coding variants predicted not to</td>
<td></td>
</tr>
<tr>
<td>affect splicing or 3) coding variants predicted to be non-pathogenic by</td>
<td></td>
</tr>
<tr>
<td>SIFT, polyphen-2 and MutationTaster</td>
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### Online Table II. Genetic variation in STAP1 in 400 FH4 patients

<table>
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<tr>
<th>exon</th>
<th>rs number</th>
<th>c.Nomen</th>
<th>p.Nomen</th>
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<th>AB</th>
<th>BB</th>
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<td>1</td>
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<td>c.12G&gt;A</td>
<td>p.= (p.Lys4Lys)</td>
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<td>1</td>
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<td>c.192+10T&gt;C</td>
<td>p.?</td>
<td>9</td>
<td>71</td>
<td>297</td>
<td>0.882</td>
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<tr>
<td>2</td>
<td>rs2306810</td>
<td>c.193-12T&gt;C</td>
<td>p.?</td>
<td>195</td>
<td>140</td>
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<tr>
<td>3</td>
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<td>p.Ile71Thr</td>
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<td>1</td>
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<tr>
<td>3</td>
<td>rs141647940</td>
<td>c.291G&gt;C</td>
<td>p.Glu97Asp</td>
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<td>4</td>
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<td>p.?</td>
<td>223</td>
<td>153</td>
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<td>c.354A&gt;G</td>
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<td>1</td>
<td>0</td>
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<td>c.730-41G&gt;C</td>
<td>p.?</td>
<td>92</td>
<td>177</td>
<td>103</td>
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<td>8</td>
<td>rs192899556</td>
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<td>p.?</td>
<td>371</td>
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</tr>
<tr>
<td>9</td>
<td>c.827-53C&gt;T</td>
<td>p.?</td>
<td></td>
<td>392</td>
<td>1</td>
<td>0</td>
<td>0.001</td>
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### Online Table III. Genetic variation in the coding region of STAP1 identified in GoNL and EVS-EA

<table>
<thead>
<tr>
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<tbody>
<tr>
<td>4:68424532</td>
<td>c.5T&gt;A</td>
<td>p.Met2Lys</td>
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<td>1</td>
<td>D</td>
<td>D</td>
<td>D</td>
<td>pathogenic</td>
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<tr>
<td>4:68424562</td>
<td>c.35G&gt;A</td>
<td>p.Arg12His</td>
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<td>D</td>
<td>D</td>
<td>D</td>
<td>pathogenic</td>
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<td>4:68424573</td>
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<td>p.Gln16*</td>
<td>nonsense</td>
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<td>N/A</td>
<td>N/A</td>
<td>N/A</td>
<td>pathogenic</td>
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<tr>
<td>4:68424619</td>
<td>c.92T&gt;A</td>
<td>p.Phe31Tyr</td>
<td>missense</td>
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<td>T</td>
<td>B</td>
<td>D</td>
<td>possible pathogenic</td>
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<td>p.Leu69Ser</td>
<td>missense</td>
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<td>T</td>
<td>D</td>
<td>D</td>
<td>pathogenic</td>
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<td>p.Ile71Thr</td>
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<td>p.Thr78Ile</td>
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<td>D</td>
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<td>p.Pro176Ser</td>
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<td>3</td>
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<td>pathogenic</td>
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<td>4:68449354</td>
<td>c.596dup</td>
<td>p.Asn199Lysfs*8</td>
<td>frame-shift</td>
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<td>N/A</td>
<td>N/A</td>
<td>pathogenic</td>
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<td>4:68449380</td>
<td>c.619G&gt;A</td>
<td>p.Aspr207Asn</td>
<td>missense</td>
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<td>3</td>
<td>T</td>
<td>B</td>
<td>possible pathogenic</td>
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<td>p.Arg209Gly</td>
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<td>T</td>
<td>B</td>
<td>P</td>
<td>non-pathogenic</td>
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<td>4:68456651</td>
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<td>p.Thr237Ala</td>
<td>missense</td>
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<td>D</td>
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<td>pathogenic</td>
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<tr>
<td>4:68459021</td>
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<td>missense</td>
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<td>D</td>
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<td>possible pathogenic</td>
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<td>4:68472019</td>
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<td>p.Glu278Lys</td>
<td>missense</td>
<td>1</td>
<td>T</td>
<td>D</td>
<td>P</td>
<td>possible pathogenic</td>
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</table>

S:SIFT; P-2:Polyphen-2; MT:MutationTaster

### Online Table IV. Clinical characteristics of non-carriers versus carriers of the replicate families.

<table>
<thead>
<tr>
<th></th>
<th>N STAP1 non-carriers* means [SD]</th>
<th>N STAP1 carriers means [SD]</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>sex (% male)</td>
<td>91</td>
<td>43</td>
<td>25</td>
</tr>
<tr>
<td>Age (years)</td>
<td>91</td>
<td>48 [16]</td>
<td>25</td>
</tr>
<tr>
<td>TC (mmol/L)</td>
<td>80</td>
<td>5.76 [1.25]</td>
<td>25</td>
</tr>
<tr>
<td>LDL (mmol/L)</td>
<td>89</td>
<td>3.83 [1.10]</td>
<td>23</td>
</tr>
<tr>
<td>HDL (mmol/L)</td>
<td>78</td>
<td>1.37 [0.37]</td>
<td>23</td>
</tr>
<tr>
<td>TG (mmol/L)</td>
<td>78</td>
<td>1.41 [0.79]</td>
<td>23</td>
</tr>
</tbody>
</table>

TC: total cholesterol; LDL: LDL-cholesterol; HDL: HDL-cholesterol; TG: triglycerides. *All family members, including branches in which the STAP1 variants were not present.
Online Figure I. Parametric LOD scores obtained in family FH4-0102

(A) Multi-point linkage analysis computed in sets of 200 markers with 0.1000 cM spacing, indicating suggestive linkage (LOD 3.0) on chromosome 4p15.1-q13.3, 11p13-q14.1 and 13q14.13-q32.1. (B) Linkage peaks computed in sets of 175 markers with 0.1000 cM spacing corresponded to a 38 Mb region at chromosome 4 (chr. 4: 32,194,465-70,584,037), a 47 Mb region at chromosome 11 (chr. 11: 35,802,039-82,312,013), and a 49 Mb region at chromosome 13 (chr 13: 46,960,154-96,027,407).
Online Figure II. STAP1 families

(A) STAP1 families

(B) STAP1 families

Supplementary note
Online Figure II. *STAP1* families. Half-blackened symbols indicate affected members, quarter-blackened symbols indicate possibly affected members, and unblackened symbols indicate unaffected members. Age (in years) at lipid measurement, total cholesterol (TC), LDL cholesterol (LDL-c), HDL cholesterol (HDL-c), triglycerides (TG) in mmol/L, history and age onset of coronary artery disease and therapeutic cholesterol-lowering intervention are given. Percentiles matched for age and gender, pTC, pLDL, pHDL, pTG and pLDL corrected for cholesterol-lowering therapy when applicable are given in brackets, a percentile indicated by 99 resembles a percentile above 95th. (a) pedigree FH4-0150 with p.Leu69Ser carriers represented by the dot, (b) pedigree FH4-0247 with p.Ile71Thr carriers represented by the dot, (c) pedigree FH4-0317 with p.Glu97Asp carriers represented by the dot, (d) pedigree FH4-0356 with p.Asp202Asn carriers represented by the dot. The triangles represent the probands. Only branches in which the *STAP1* variant was present are depicted here.


Supplementary note
Online Figure III. Gene structure of *STAP1*.

PH: Pleckstrin homology domain, SH2: Src homology 2 domain. The positions of tyrosine residues are indicated by arrowheads.