Abstract: Studies in humans and in mice have highlighted the importance of short telomeres and impaired mitochondrial function in driving age-related functional decline in the heart. Although telomere and mitochondrial dysfunction have been viewed mainly in isolation, recent studies in telomerase-deficient mice have provided evidence for an intimate link between these two processes. Telomere dysfunction induces a profound p53-dependent repression of the master regulators of mitochondrial biogenesis and function, peroxisome proliferator-activated receptor gamma coactivator (PGC)-1α and PGC-1β in the heart, which leads to bioenergetic compromise due to impaired oxidative phosphorylation and ATP generation. This telomere-p53-PGC mitochondrial/metabolic axis integrates many factors linked to heart aging including increased DNA damage, p53 activation, mitochondrial, and metabolic dysfunction and provides a molecular basis of how dysfunctional telomeres can compromise cardiomyocytes and stem cell compartments in the heart to precipitate cardiac aging. (Circ Res. 2012;110:1226-1237.)

Key Words: heart aging ■ telomere ■ mitochondria ■ metabolism ■ PGC-1α ■ myocardial regeneration ■ telomerase ■ transcription factors ■ transcriptional coactivator

Societies around the world are experiencing a dramatic increase in the elderly population (people 65 years and older). It is predicted that by the year 2030, 1 billion individuals (1 in 8 individuals) will be 65 years or older worldwide. In the United States, the percentage of the elderly population is expected to increase from 12.9% to 19.7% (40.2 to 88.5 million people total) from 2010 to 2050. Within the elderly, the oldest old (85 years and older) is the fastest growing section and is projected to grow from 1.8% to 4.2% (5.7 to 19.0 million) in the same time period.1 This rise of an

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aging population is accompanied by a sharp increase in the prevalence of age-related chronic diseases ranging from cardiovascular disease (CVD) and cancer to metabolic syndrome and Alzheimer’s disease.

CVD includes coronary artery disease (CAD), hypertension, and chronic heart failure and is a major cause of chronic disability and the leading cause of death in the United States. The prevalence of CVD for men and women combined jumps from 12% in individuals younger than 40% to 38% in the 40 to 60 group to a staggering 83% in those older than 80 years of age. CAD is the most common CVD and manifests as angina pectoris and myocardial infarction, both of which are significantly more common in the elderly. Eighty-two percent of patients who die of CAD are 65 years or older. Similarly, aging is associated with a dramatic increase in the prevalence of hypertension and chronic heart failure. Although substantial progress in the prevention and treatment of CVD has led to a significant decline in death rates over the last 2 decades, morbidity and mortality rates associated with CVD continue to be high and remain a tremendous burden for the national health care system. In 2010 alone, CVD-related costs were estimated to be $444 billion.

A long-term solution to this social and health care crisis will require a better understanding of how aging per se drives cardiovascular decline. This would facilitate the development of effective preventive and therapeutic strategies. Telomeres, repetitive TTAGGG sequences at the ends of chromosomes, have been strongly implicated in aging. Cumulative studies in humans with telomere maintenance disorders and telomerase knock-out mice have demonstrated that short telomeres precipitate functional decline in different tissues, including the cardiovascular system. Mechanistically, telomere dysfunction–driven tissue compromise is thought to be secondary to the activation of DNA damage signaling pathways that converge on p53, a central executor of the DNA damage response pathway. p53 activation induces senescent and apoptosis pathways, particularly in stem cell and progenitor compartments of highly regenerative organs. The elimination of stem and progenitor cells is thought to be the driving force in the development of tissue defects. Telomeres are also linked to aging and disease development in more quiescent tissues such as the heart, and recent studies have uncovered a different mechanism that could explain how telomere dysfunction affects not only proliferative but also quiescent tissues that rely less on stem cell–driven regeneration. These studies have demonstrated that telomere dysfunction–activated p53 directly leads to mitochondrial and metabolic compromise through the repression of the master regulators of mitochondrial biogenesis and function, peroxisome proliferator-activated receptor gamma coactivator (PGC)-1α and PGC-1β. PGC repression is associated with a profound compromise in mitochondrial biogenesis and function and subsequent decline in ATP generation, indicating that a fundamental problem of energy maintenance drives the aging process.

Independent studies in the aging heart have noted a significant decline of mitochondrial function, including compromised oxidative phosphorylation, ATP generation capacity, and β-oxidation. This has lead to the hypothesis that human heart failure is a consequence of mitochondrial dysfunction and energy starvation. In the present report, we review the role of these two drivers of cardiac aging—the mitochondrial/energy maintenance and the telomere hypotheses—in CVD, with the main focus on heart disease, and discuss how telomere dysfunction converges on mitochondria and energy maintenance pathways to drive cardiac aging and failure.

**Mitochondrial Alterations in the Aging Heart**

The heart undergoes a number of functional and structural changes during aging, even when no contributing cardiac risk factors such as hypertension or diabetes exist (Table). These changes include a 35% loss of cardiomyocytes and a 20% decrease of sinoatrial nodal pacemaker cells due to apoptosis, necrosis, and potentially autophagy. The aging heart also contains more senescent cardiomyocytes as defined by the expression of senescence markers p53, p21, and p16 and the presence of shorter telomeres. This continued loss of functional resident cardiac cells is paralleled by a decline in regenerative activity from 1% per year at age 20 to 0.4% at the age of 75. Age-related changes often include left ventricular hypertrophy and diastolic dysfunction, which lead to increased end-diastolic pressure. The cumulative effect of all these changes together with the coexistence of other diseases predisposes older individuals to develop typical aged-associated heart disease: diastolic heart failure (defined as presence of heart failure symptoms in patients who have preserved left ventricular function but impaired diastolic function), atrial fibrillation, and cardiac ischemia.

The heart contains a very high density of mitochondria due to its extraordinary demand for ATP, which is mainly provided by oxidative phosphorylation (OXPHOS). Other biochemical processes involved in energy production such as fatty acid oxidation (FAO), Krebs cycle, and tricarboxylic acid cycle also take place in mitochondria. Except for 13 subunits of the oxidative phosphorylation chain that are encoded by the mitochondrial genome, all mitochondrial bioenergetic processes take place in mitochondrial compartments.
proteins are encoded within the nucleus. Mitochondrial biogenesis is regulated on the transcriptional level by a set of nuclear encoded coactivators and transcription factors. The first coactivator demonstrated to regulate mitochondrial biogenesis and function was identified as a coactivator of nuclear receptor peroxisome proliferator-activated receptor (PPAR) γ in brown adipose tissue and named PGC-1α.16 Subsequently, 2 more related coactivators were identified: PGC-1β and PGC-related coactivator (PRC).17,18 Of these 3 PGC mem-

bers, PGC-1α and PGC-1β have been well studied and demonstrated to coordinate mitochondrial biogenesis and function through association with different transcription factors, including nuclear respiratory factors 1 and 2 (NRF-1 and NRF-2), estrogen-related receptor α (ERRα), and PPAR, to drive the expression of mitochondrial genes important for mitochondrial energy transduction, ATP generation, and mitochondrial biogenesis.19

Age-related mitochondrial changes include increased reactive oxygen species (ROS), impaired OXPHOS, reduced ATP generation, impaired FAO, and increased mtDNA mutations.20–23 The pathophysiological relevance of alterations in each of these processes for cardiac disease has been documented in patients with genetic disorders.24 Patients with loss-of-function mutations in genes essential for mtDNA integrity, OXPHOS, or β-oxidation develop significant cardiac compromise and cardiomyopathy.24–28 Studies in genetically modified mice have supported these clinical observations. Mice carrying mutations in genes resulting in compromised fatty acid transport and oxidation, ATP generation, protection from ROS, and mtDNA proofreading capacity all develop cardiac dysfunction. Although the study of single gene mutations has highlighted the importance of specific processes for the maintenance of cardiac function, it has been noted that there is often compromise of multiple mitochondrial processes in aged heart tissues.27,28 According to the mitochondrial variant of the free radical theory of aging, this is due to increasing levels of mitochondrial ROS, which leads to a progressive forward-feedback spiral of increasing damage to mtDNA and other mitochondrial components. This in turn induces more ROS through damage to the electron transport chain, furthering ROS production and more mtDNA damage and ultimately leading to bioenergetic failure.27,28 Support for an important role of ROS in driving cardiac aging comes from findings in mice overexpressing catalase (an antioxidant enzyme) selectively in heart mitochondria.29 Mitochondrial catalase overexpression reduces ROS-mediated damage to proteins and mtDNA, prevents functional decline in the heart and increases median lifespan.29 Interestingly, catalase overexpression also improves the cardiomyopathy observed in mice expressing a proofreading-impaired variant of mitochondrial polymerase γ, which is involved in mtDNA replication and repair.30 The expression of this variant is associated with a substantial increase in mtDNA point mutations and deletions and an accelerated aging phenotype including cardiomyopathy (so-called mutator mouse).31,32 The beneficial effect of catalase overexpression on cardiac function in the mutator mice is deleterious.33 Increased mtDNA damage observed in mutator mice are beyond the levels seen during natural aging in human and rodents, the development of cardiomyopathy in these mice highlights the importance of mtDNA integrity for normal cardiac function.33 Increased mtDNA
mutations and deletions have been documented in aged heart tissues of humans and rodents, and this might be facilitated by the lack of mtDNA histones and the limited mtDNA repair capacity. Mechanistically, the increase in mtDNA damage is thought to compromise the electron transport chain through dysregulated transcription of mtDNA encoded respiratory chain components. This view is supported by the findings in the mutator mouse, which have depressed complex I and IV activities, reduced ATP generation, and subsequently develop dilated cardiomyopathy.32 The decline in electron transport chain activity has been also noted in naturally aged hearts in humans and rodents but might affect only a subset of mitochondria.27,34 The decline in electron chain activity is often associated with impaired ATP generation.8,34 Other age-dependent changes in mitochondrial energy pathways have been noted and are thought to contribute to ATP depletion phenotype, including decreased FAO rates and an increase in glycolysis in animal models of pressure overload-induced cardiac hypertrophy and heart failure.8

In humans, there is growing evidence that cardiac hypertrophy and heart failure have also altered energy pathways, although the changes are not uniform. For example, whereas hypertension-induced cardiac hypertrophy and heart failure is characterized by impaired FAO and a shift to glucose utilization resembling the fetal situation with reliance of glycolysis for ATP generation,35 the heart of diabetic patients utilizes primarily fatty acids.36–38 Furthermore, the degree of cardiac dysfunction appears to be an important determinant for fuel preference based on studies demonstrating that FAO is unchanged or increased in early heart failure but significantly decreased in later stages. Similarly, glucose utilization is increased early in heart failure but drops significantly in advanced heart failure.8 The progression to heart failure in humans is associated with a decline of ATP content and reduction in the ATP transfer capacity.8 PCr/ATP ratios are reduced with heart failure severity and are good predictors of cardiovascular mortality.39

The Role of PGCs in Mitochondrial and Heart Decline

Multiple signaling pathways converge to compromise mitochondrial function during aging in the heart (Figure 1). Among these, PGC family members have gained particular interest because of their capacity to drive virtually all aspects of mitochondrial biogenesis in the heart, including mitochondrial number, mitochondrial respiration, expression of OXPHOS and FAO genes, and ROS levels.19,40 Gain-of-function and loss-of-function studies highlight the critical role of PGC family members in the heart. One study showed that transgenic PGC-1α overexpression in mice leads to significant increase in mitochondrial number in the heart, leading to lethal cardiomyopathy.40 Mice lacking either PGC-1α or PGC-1β have no overt cardiac phenotype.41–43 However, more detailed analysis has uncovered a compromise in bioenergetics (reduced FAO, ATP synthesis, and myocardial efficiency) and defects in cardiac function (isolated hearts showed decreased cardiac work and output) at baseline.41,43 Under conditions of increased hemodynamic stress and increased bioenergetic demands, PGC-1α− and PGC-1β−deficient mice develop overt cardiac failure.42,44 Decreased PGC-1α expression has been also linked to the development of heart failure in other mouse models, including mice overexpressing cyclin-dependent kinase 9 (Cdk9).45 Cardiac failure is even more pronounced in mice lacking both coactivators, indicating a functional overlap of these coacti-
The relevance of reduced telomeres and activation of p53 play a role. This issue will be discussed next.

**Telomere Structure**

Telomeres consist of several kilobases of tandem-repeat double-stranded TTAGGG sequences that are followed by a single-stranded, few hundred nucleotide-long 3′-overhang that loops back into telomeres to form a so-called t-loop. The essential function of the t-loop is to prevent telomere ends from being recognized as break points by the DNA damage repair machinery. Telomeres associate with a number of proteins, 6 of which are highly telomere specific (TRF1, TRF2, POT1, TIN2, TPP1, and Rap1) and form the shelterin complex. The shelterin complex is essential for the formation of the t-loop, maintenance of telomere length, and the suppression of the DNA damage response (DDR; Figure 2). Other non–telomeric-specific proteins contribute to telomere maintenance and include DNA repair (XPF/ERCC1, Rad51D, Ku70/80, Apollo, Mre11), DNA replication (RecQ helicase and ORC), and DNA damage signaling proteins (Mre11 and 9–1–1 complex). Although it was widely believed that the only role of telomeres is to provide structural stability to chromosomal ends, recent studies have demonstrated that mammalian telomeres are transcribed into telomeric repeat-containing RNA (TERRA). Akin to previous work in flies, trypanosomes, and birds, TERRA transcription in mammalian cells starts at subtelomeric regions and moves outward to generate TERRA of various lengths which localize at telomeres.

**Figure 2. Telomere structure and telomerase.** Telomeres (shown in green by fluorescence in situ hybridization) are repetitive TTAGGG sequences at the ends of chromosomes. The 3′ end contains a G-rich, single-stranded overhang that loops and invades the double-stranded telomeric region to form a so-called t-loop (not depicted). Telomeres exist in association with many different proteins that contribute to the overall stability. Six telomere-binding proteins (POT1, TRF1, TRF2, TIN2, TPP1, and Rap1) are telomere-specific and form the shelterin complex. Three, POT1, TRF1, and TRF2 directly bind to telomeres, whereas TIN2, TPP1, and Rap13 connect them (A). Telomeres are maintained by the enzyme telomerase, which consists of a RNA subunit (AAUCCC) and the reverse transcriptase, TERT, which are illustrated. The catalytic component TERT uses the RNA component to synthesize new telomeres (B). Progressive telomere shortening leads to telomere shortening and activation of the DNA damage machinery (S3BP1, Mre11, and (phosphorylated) p-H2AX, p-ATM, and others) and subsequent activation of p53 and induction of apoptosis, growth arrest, and senescence (C). Telomere length is measured traditionally by Terminal Restriction Fragment (TRF) length analysis by Southern blot. TRF assesses the average telomere length within a cell population with up to 1-kb resolution. Telomere length can be alternatively measured by fluorescence in situ hybridization (FISH) with greater sensitivity (resolution, 0.3 kb) on metaphase or interphase chromosomes using directly fluorescently labeled (CCCTAA)3 peptide nucleic acid (PNA) probes. This method can be combined with immunohistochemical staining to detect TIFs. A variant of FISH has been adopted for flow (Flow-FISH). Newer methods use PCR methods with greater sensitivity (Single Telomere Length Analysis or STELA, Q-PCR, and monochrome multiplex Q-PCR).

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regulated in a species-, tissue- and cell type–specific manner. There is a growing appreciation that telomere length is not fixed and can be modulated by exercise or psychological factors.60–64 Telomeres are maintained by the enzyme telomerase. Telomerase is composed of a catalytic active component TERT and the telomerase RNA component (TERC), which serves as a template during telomere biosynthesis (Figure 2). In the absence of telomerase, cells are not capable of solving the end-replication problem and experience telomere shortening with each successive round of DNA replication.65,66 In humans, with the exception of progenitor and stem cell compartments and some other notable exceptions such as activated lymphocytes, most tissues have no or very little telomerase activity primarily due to transcriptional repression of TERT. However, even in compartments with documented telomerase activity, such as CD34 positive hematopoietic stem cells, telomere length is not maintained as a function of age in humans, pointing to other levels of regulation in the maintenance of telomeres.4

Telomeres and Telomerase in Aging
The importance of adequate telomerase activity and maintenance of telomere length for both replicative potential in cultured cells and organismal aging was initially inferred from a series of seminal studies using cultured human fibroblasts. These in vitro experiments demonstrated that continuous passage of fibroblasts resulted in telomere shortening and eventual cellular senescence after a finite number of cell divisions, a barrier known as the Hayflick limit.67 The critical role of adequate telomere length in this senescence process was demonstrated by the capacity of enforced TERT expression to stabilize telomere length and enable unlimited replicative potential.68 These paradigmatic studies strongly implicated telomeres in processes of cellular senescence or “aging,” which were further substantiated in humans and in telomerase deficient mice.

Patients with autosomal dominant dyskeratosis congenita carry mutations in TERC or TERT and have shortened telomeres, signs of accelerated aging, and a reduced lifespan.69 TERC or TERT mutations have also been linked to more organ-restricted disease manifestations such as idiopathic pulmonary fibrosis (a fatal lung disease with excessive fibrosis) and bone marrow failure syndromes.70 Interestingly, telomerase mutations, although not widely recognized to confer predisposition to develop cardiac disease, have been recently linked to the development of cardiac disease such as cardiac fibrosis and cardiomyopathy in few reports (see below). The factors that determine disease localization and extent of clinical manifestations in patients with telomerase mutations remain unknown, although the degree of telomere dysfunction plays an important role as severity and onset of disease manifestation correlate with telomere length.71

Similarly, Werner syndrome patients with mutations in the WRN gene, which encodes a RecQ helicase involved in DNA repair, DNA recombination, and telomere maintenance, manifest clinically with multiple premature aging pathologies after puberty, including senile cataracts, osteoporosis, hair graying, myocardial infarction, and cancer.71 A potential role of telomeres in human disease pathogenesis was inferred by the capacity of enforced TERT expression to rescue the limited replicative potential of Werner syndrome fibroblasts with shortened telomeres.72–74 Taken together, studies of inherited human degenerative conditions support the view that telomere dysfunction is a rate-limiting pathogenic element underlying degenerative phenotypes and shortened lifespan.

The understanding of the in vivo relevance and specific roles of telomeres in aging and degenerative diseases was initially provided by the generation of mice null for either the Tert or Terc component of telomerase.4,75 Deletion of either component was found to be fully compatible with normal embryonic and postnatal development, indicating that telomerase per se is dispensable for life.75 However, the limited impact of telomerase deficiency was found to be related to the long starting telomeres in inbred laboratory mouse strains. Generation of mice with increasingly shorter telomeres through continuous interbreeding (G1×G1×G2, G2×G2×G3, etc) revealed that onset of telomere dysfunction dramatically diminished organismal fitness, promoted age-associated diseases, and reduced lifespan.76 On the tissue level, “late-generation” Terc−/− mice with short dysfunctional telomeres show widespread tissue atrophy with impaired organ function.76 Whereas proliferative tissues such as the hematopoietic system (anemia and lymphopenia), intestine (atrophy), skin (alopecia, hair graying, decreased wound healing), reproductive system (infertility), and skeletal system (kyphosis and osteoporosis) are prominently affected, there is also significant tissue pathology in more quiescent tissues such as the heart (cardiomyopathy, see also below) as well as generalized metabolic aberrations including insulin resistance. The severity of these conditions is dependent on the degree of telomere dysfunction, as measured by the number of chromosomal fusions, anaphase bridges, and number of DNA damage foci at telomeres (telomere-induced foci [TIFs]).77 Studies in the mouse have further established that telomere dysfunction in a few chromosomes is sufficient to drive degenerative changes as observed in “classic” late-generation TERC−/− mice.78 The relevance of limiting telomeres to the aging process is further substantiated through intercrossing telomerase deficient mice with mice with classic human degenerative-disease mutations such as WRN or ATM.79,80 Strikingly, these mutant mice fail to recapitulate the classic human phenotypes unless telomeres are reduced to a more human-like length, thus supporting the view that telomere dysfunction plays a critical role in the pathogenesis of these conditions.

The Relevance of Telomeres in CVD
Studies in Tert and Terc knock-out mice have advanced our understanding of the relevance of critical short telomeres for cardiovascular aging and CVD. Late-generation Terc and Tert knock-out mice develop hallmark features of cardiomyopathy with severe systolic and diastolic dysfunction. This cardiomyopathy is linked to increased p53 levels and cardiomyocyte apoptosis.81 Cardiomyocyte specific Tert overexpression maintains telomere length in the adult mouse heart, inhibits apoptosis and extends the capacity of cardiomyocytes to proliferate from 8 to 12 weeks.82 Intriguingly, continued
expression of Tert induces cardiac hypertrophy without fibrosis or impaired myocardial function, indicating a role for telomerase beyond modulation of proliferation and apoptosis in the heart. 

A causal role for telomere dysfunction in the pathogenesis of human CVD is supported by 3 lines of evidence. First, case reports indicate that CVD might be more common in patients with telomerase germline mutations than previously appreciated. Two members of a family with autosomal dominant mutations in TERT developed cardiac disease at an early age. One member developed dilated cardiomyopathy, whereas a second one had severe cardiac fibrosis. In a different study, myocardial infarction at a young age was documented in a patient carrying a mutation in telomerase. Accordingly, other telomere maintenance disorders such as Werner syndrome are associated with premature CVD including atherosclerosis, aortic stenosis, valvular disease, mitral regurgitation, and calcification. Although a mouse model of Werner syndrome develops cardiac fibrosis, only 1 instance of florid cardiac fibrosis in a 17-year-old with Werner syndrome has been reported. Although these reports are intriguing, systematic studies in asymptomatic patients are needed to more precisely quantify the prevalence and extent of cardiac compromise in patients with telomere maintenance disorders. Second, human cardiomyocytes and cardiac stem cells display an age-dependent decline in telomere length and express senescence markers such as p53, p21, and p16. These changes correlate with a decreased proliferative and regenerative capacity of aged cardiomyocytes and cardiac stem cells. Notably, failing human hearts with decreased expression of telomere repeat binding-factor 2 (TRF2) contain 25% shorter telomeres than age-matched healthy control subjects. Consistent with this documented telomere attrition, there is robust activation of DNA damage kinase, checkpoint kinase 2 (Chk2) and increased cardiomyocyte apoptosis. Third, the relevance of critical short telomeres for human heart aging and disease is inferred from a large number of epidemiological studies demonstrating associations between shortened telomeres, aging, and CVD. In these studies, leukocyte telomere length correlates inversely with multiple cardiovascular risk factors, including cigarette smoking, obesity, diabetes, systemic inflammation, and physical inactivity. In case-control studies, decreased leukocyte telomere length is associated with CAD and premature myocardial infarction as well as intermediate phenotypes such as carotid media thickness and coronary calcification. Prospective cohort studies show that decreased leukocyte telomere length independently predicts mortality and cardiovascular events in the general population, patients with stable CAD, and patients with severe congestive heart failure. These studies are complemented by a nested case-control study, carried out as part of a large, randomized trial involving statin treatment. In this study, individuals with shorter leukocyte telomere length had a significantly higher risk for developing coronary heart disease over 5 years. In contrast to the above studies, longer telomeres have been associated with increased left ventricular mass in 2 independent studies, but the relationship between these 2 observations and the consistent association of short telomeres with adverse outcomes requires further study.

The interpretation of the above studies remains complicated, given that most studies are cross-sectional and capture telomere length only at a single time point. As such, leukocyte telomere length may be a consequence of coronary heart disease rather than causative. Furthermore, the extent to which telomere length in circulating leukocytes reflects telomere dynamics in tissues such as the heart remains largely unclear, although a close correlation between leukocyte and aortic wall tissue telomere length has been reported. 

**Cellular and Molecular Mechanisms of Telomere-Driven Aging in the Heart**

The primary mechanism of telomere-driven aging rests on the principle that cellular turnover results in progressive loss of telomere reserves, which ultimately generates short dysfunctional telomeres. These dysfunctional telomeres were shown to activate p53-dependent DNA damage checkpoint mechanisms such as proliferative arrest, apoptosis, and senescence. This telomere-driven process was particularly evident in highly proliferative tissues with high renewal requirements, which aligns with the depletion of stem and progenitor cell reserves in these tissues.

In the heart, cardiac stem and progenitor cells have been reported to undergo telomere attrition during aging despite telomerase activity, similar to what has been described in other stem cells. The documented telomere shortening in aged cardiac stem/progenitor cells could impair cardiomyocyte renewal. However, whereas the decline in cardiac stem/progenitor cell–dependent regeneration might contribute to cardiac aging, the overall renewal capacity, even of young hearts, is low and inadequate to replace ongoing age-associated cardiomyocyte loss.

The impact of dysfunctional telomeres on mature cardiomyocytes is evident on 2 levels. First, telomere attrition has been demonstrated to modestly induce cardiomyocyte apoptosis, and, conversely, cardiac specific telomerase overexpression is associated with decreased apoptosis in vivo and in cultured cardiomyocytes. Second, telomere dysfunction might impair the regenerative capacity of cardiomyocytes or the ability of a subset of cardiomyocytes to regenerate, similar to what has been shown in the liver, where terminally differentiated hepatocytes can reenter cell cycle but are impaired in the setting of telomere dysfunction. Thus, the combined negative impact of dysfunctional telomeres on cardiac stem/progenitor cells and mature cardiomyocytes might explain some aspects of the failing heart in the aged.

More significantly, recent genetic studies in the mouse have established that telomere dysfunction not only induces the cellular phenotypes of proliferative arrest, apoptosis, and senescence but compromises cardiomyocyte function through repression of PGC-1α and PGC-1β. The combined PGC-1α and PGC-1β repression is associated with impaired mitochondrial biogenesis and function in the heart and other tissues. The decrease in mitochondrial mass and impairment in the electron transport chain negatively affects ATP synthesis capacity and leads to a decrease in cardiac ATP levels. Furthermore, metabolic processes such as β-oxidation and fatty acid synthesis are affected. Importantly, the mitochondrial and metabolic changes are reversible as PGC-1α
overexpression in telomere dysfunctional mice rescues the oxidative electron chain defect.6 Interestingly, cardiomyocyte-specific telomerase overexpression prevents the decline in telomere length during postnatal heart maturation and induces hypertrophy.82 The basis for the hypertrophy was not in telomere length during postnatal heart maturation and cyte-specific telomerase overexpression prevents the decline of classic cellular outcomes of apoptosis, senescence, and growth arrest.6,5 Mitochondrial dysfunction per se can lead to growth arrest, senescence, or apoptosis, depending on extent and duration as well as other cellular events. Telomere dysfunction activates other p53-independent pathways to mediate mitochondrial dysfunction and cellular phenotypes. The importance of PGC for stem cell maintenance is also suggested by studies in drosophila. Here, overexpression of the Drosophila PGC-1 homolog (dPGC-1/spargel) and increasing mitochondrial activity in stem and progenitor cells of the digestive tract is sufficient to extend the lifespan.109 That said, further study is needed to firmly establish the relevance of these mitochondrial and metabolic changes in stem cell function, particularly given recent studies demonstrating that HSCs are more dependent on glycolysis than oxidative phosphorylation.110 Whether this is true for cardiac stem cells is unclear.

Although the molecular pathways that sense telomere dysfunction and activate cellular checkpoint responses as well as PGC repression have yet to be established, p53 has been shown to be the central effector of telomere dysfunction (Figure 3).5 The importance of p53 activation for the development of degenerative disorders was demonstrated by the rescue of most degenerative phenotypes in telomerase knock-out mice also null for p53.5 This widespread rescue by p53 deficiency was mainly attributed to the rescue of proliferative arrest, apoptosis, and senescence. In more recent studies, however, the role of p53 has been expanded by the discovery that activated p53 can directly repress PGC-1α and PGC-1β gene expression on the transcriptional level in a manner dependent on the binding of p53 to PGC-1α and PGC-1β promoters. Accordingly, p53 deficiency leads to increased PGC expression and partially rescues mtDNA copy number in heart and liver tissues in telomere dysfunctional mice. On a functional level, p53 deficiency improves low-dosage, doxorubicin-induced cardiomyopathy in telomere dysfunctional mice.6 The prominent role of p53 in driving cardiac failure/regeneration is further supported by the increase in p53 in aging hearts and in heart failure.5,111,112 Genetic evidence for an essential role of p53 in the development of cardiac failure derives further support from cardiac-specific
deletion of Mdm4, a negative regulator of p53. These mice dilitate cardiomyopathy through increased apoptosis and senescence.113 Furthermore, chronic p53 elevation inhibits hypoxia-inducible factor-1 (Hif-1)-dependent induction of angiogenic factors and induces the transition from hypertrophy to heart failure in a mouse model of cardiac hypertrophy.112 Similarly, hyperactivation of p53 in the setting of cardiac specific deletion of Sirt1 (which inhibits p53 through deacetylation) predisposes cardiomyocytes to cell death, and, conversely, modest Sirt1 overexpression in cardiomyocytes attenuates age-dependent cardiomyopathy.107,114,115 Given that Sirt1 is an important activator of PGC, cardiac specific increased PGC expression could be a major contributor to the attenuated cardiomyopathy phenotype in this model.

Conclusion and Perspective

The study of human telomere maintenance disorders and genetic model systems has advanced our understanding of the role of telomeres in the aging process. These studies have pointed to the importance of telomere integrity in maintaining tissue homeostasis and regulating lifespan. Whereas the role of telomere dysfunction in driving tissue atrophy in high turnover tissues is well established, there is also growing recognition that tissues with less proliferative profiles are also affected. Increasing evidence indicates that critical short telomeres play an important role in cardiac aging and disease. Some of the conflicting results relate to the differences in assessing telomere status and the reliance on measurements of telomere length in peripheral leukocytes. For future human studies, it would be desirable to standardize protocols for analyzing the degree of telomere dysfunction (ie, TIFs) rather than telomere length in cardiomyocytes, where possible. It will be also important to identify new markers that might allow more straightforward and technically less challenging assessment of telomere dysfunction. How telomere dysfunction develops in cells with a low degree of cell turnover such as cardiomyocytes is largely unclear and remains an important are of investigation. The relevance of telomeres for CVD might find further clarification from much-needed prospective studies in patients with telomere maintenance syndromes. Systematic analysis of cardiovascular function in these patients will establish the prevalence of cardiac compromise, which might be underreported due to the absence of symptoms.

On a mechanistic level, recent reports linking telomere dysfunction to metabolic and mitochondrial compromise provide a novel mechanism as to how dysfunctional telomeres can compromise cardiac function. This telomere-p53-PGC-mitochondrial axis aligns with many changes seen in aged hearts: impaired OXPHOS, decreased ATP generation, and increased ROS levels. Other metabolic changes, including compromised FAO and a shift to glucose utilization, can be expected to be altered with telomere dysfunction, based on the downregulation of the PGC-PPARγ axis in mice with critical short telomeres. It will be important to establish whether similar mitochondrial/metabolic decline occurs in hearts and other tissues of patients with telomerase mutations and short telomeres.

Besides p53, other genes and pathways appear to be involved in compromising mitochondrial function in the setting of telomere dysfunction, as p53 deficiency only partially rescues PGC expression and mitochondrial dysfunction. The identification of these other pathways will expand our understanding of how critical short telomeres compromise cellular function and precipitate aging. The telomere-p53-PGC link might be useful in both identifying aging biomarkers as well as illuminating targeted therapeutic avenues in the prevention and reversal of age-associated cardiovascular decline. In keeping with this, therapeutic strategies to stabilize DNA integrity, attenuate DDR signaling, or enhance mitochondrial function through pharmacological and nonpharmacological means such as continuous exercise—which is demonstrated to reverse the premature aging phenotype of the mutator mouse—are possible avenues to pursue.116 This telomere-mitochondrial link further adds to the notion that compromised energy maintenance pathways are central to heart aging and failure. Accordingly, metabolic pathways can be considered a promising target to delay and treat heart failure.

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None.

References

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