This Review is part of a thematic series on **Nuclear Mechanotransduction**, which includes the following articles:

- Nuclear Shape, Mechanics, and Mechanotransduction [2008;102:1307–1318]
- Emerin and the Nuclear Lamina in Muscle and Cardiac Disease [2008;103:16–23]

**Mechanical Control of Tissue Morphogenesis**

_Dennis Discher, Guest Editor_

**Mechanical Control of Tissue Morphogenesis**

Parth Patwari, Richard T. Lee

**Abstract**—Mechanical forces participate in morphogenesis from the level of individual cells to whole organism patterning. This article reviews recent research that has identified specific roles for mechanical forces in important developmental events. One well defined example is that dynein-driven cilia create fluid flow that determines left-right patterning in the early mammalian embryo. Fluid flow is also important for vasculogenesis, and evidence suggests that fluid shear stress rather than fluid transport is primarily required for remodeling the early vasculature. Contraction of the actin cytoskeleton, driven by nonmuscle myosins and regulated by the Rho family GTPases, is a recurring mechanism for controlling morphogenesis throughout development, from gastrulation to cardiogenesis. Finally, novel experimental approaches suggest critical roles for the actin cytoskeleton and the mechanical environment in determining differentiation of mesenchymal stem cells. Insights into the mechanisms linking mechanical forces to cell and tissue differentiation pathways are important for understanding many congenital diseases and for developing regenerative medicine strategies. (Circ Res. 2008;103:234-243.)

**Key Words:** mechanotransduction ■ cytoskeleton ■ shear stress ■ embryonic development ■ stem cells

If you wonder how mechanical forces can regulate the highly orchestrated process of morphogenesis, you have only to look at the tips of your fingers and ask: Are the fingerprints of identical twins also identical?

The people of Stevenage, England still celebrate their most famous identical twins, Albert Ebenezer and Ebenezer Albert Fox, who lived from 1857 to 1926. For their habit of roaming freely on the land while hunting, the Fox twins managed to accumulate 150 convictions for poaching between them. Despite the convictions, their enthusiasm for mocking the local police force never flagged. Inevitably, they complained, the police always managed to convict the wrong twin for the crime. Yet the fingerprints of identical twins are not identical. Some characteristics, such as pattern type and ridge count, are similar between twins. However, the detailed “minutiae”—where skin ridges meet, end, or bifurcate—are different even between identical twins.

The fingertip skin ridges are thought to form as compressive stresses develop in the dermal cell layer. Like the buckling of landmasses under compression, regular ridges form to relieve the stress. Where the skin is flat, uninteresting parallel ridges develop. But primates have raised pads on their fingertips at this stage of the embryo. With the addition of curvature, the ridges are no longer formed in...
Parallel lines; instead, they form along lines of equal stress. Surrounding the highest point of the raised pad, ridges form in concentric circles, like a contour plot of elevation surrounding a hill.

Just as the ridges are forming, the pads are regressing, and the relative timing of the two processes leads to differences in fingerprint pattern. Where the pads remain high longer, ridges form regular concentric circles or “whorls”. Largely regressed pads lead to a simple “arch” pattern. Most commonly, both processes overlap, yielding an intermediate “loop” pattern. The fingerprint ridges are therefore shaped by the combined effects of cell proliferation and mechanical forces.

However, the ridges are not always regularly spaced: there are ridges that divide and ridges that suddenly end. These irregularities are easily changed by small variations in their local environment, so they are not predetermined. Even monozygotic twins will have different positions in the uterus and experience a slightly different environment. In genetically identical twins, then, subtle differences in the mechanical environment of the embryo in utero are sufficient to drive a developing system toward different morphogenic outcomes.

Mechanical forces regulate not only dermal ridge patterns, but also fundamental aspects of morphogenesis. In this review, we present several examples of recent research that have shed light on how mechanical processes can control development (Figure 1).

Gastrulation, the first major morphological change in the embryo of the fruit fly *D melanogaster*, is controlled by contraction of nonmuscle myosin. Before gastrulation, the embryo forms a single layer of cells arranged in a cylindrical egg (shown in cross-section, left panel), and myosin is localized to the inner surface. At the start of gastrulation, myosin relocalizes to the outer surface of a few cells, and its contraction pulls the outer portions of these cells together. This compresses the inner portions of the cells, pushing them inwards and creating a bulge (middle panel). Soon the cell layer has folded within itself (right panel). Differentiation of mesenchymal stem cells is controlled by the level of nonmuscle myosin contraction inside the cell and the stiffness of its surroundings. Culturing cells on hard substrates will activate nonmuscle myosin contraction and promote differentiation toward osteoblasts. Culturing cells on soft substrates, or minimizing contact with hard surfaces, promotes differentiation toward adipocytes and chondrocytes.

![Figure 1. Schematic overview of morphological events demonstrating the involvement of mechanical forces.](image)

---

**Figure 1.** Schematic overview of morphological events demonstrating the involvement of mechanical forces. Top row, Early mammalian embryos determine their right from their left by creating fluid flow in one direction. Cilia located in a specialized node at the midline propel flow to the left. The embryo rapidly senses the fluid flow and upregulates the signaling molecule, nodal, on the left side only (shown in blue). Inherited defects in cilia motor proteins result in randomization of left-right patterning. In humans, this leads to Kartegener syndrome in 50% of patients. Second row, During embryonic vasculogenesis, remodeling of immature vessels requires shear stress created by fluid flow. Before the beginning of fluid flow, embryonic mesodermal cells form “blood islands” outside the embryo. After the embryonic heart tube begins to function, fluid begins to flow and rapidly reshapes the blood islands into mature branched arteries and veins. This process specifically requires fluid shear stress. Third row, Gastrulation, the first major morphological change in the embryo of the fruit fly *D melanogaster*, is controlled by contraction of nonmuscle myosin. Before gastrulation, the embryo forms a single layer of cells arranged in a cylindrical egg (shown in cross-section, left panel), and myosin is localized to the inner surface. At the start of gastrulation, myosin relocalizes to the outer surface of a few cells, and its contraction pulls the outer portions of these cells together. This compresses the inner portions of the cells, pushing them inwards and creating a bulge (middle panel). Soon the cell layer has folded within itself (right panel). Bottom row, Differentiation of mesenchymal stem cells is controlled by the level of nonmuscle myosin contraction inside the cell and the stiffness of its surroundings. Culturing cells on hard substrates will activate nonmuscle myosin contraction and promote differentiation toward osteoblasts. Culturing cells on soft substrates, or minimizing contact with hard surfaces, promotes differentiation toward adipocytes and chondrocytes.
Control of Morphogenesis by Fluid Flow
Cilia Determine Left Versus Right in Mammals by Creating Leftward Flow

The classic description of patients with Kartagener’s syndrome has been a puzzling combination: left-right reversal of the major visceral organs (situs inversus) accompanied by mucus-blocked airways in the lungs (bronchiectasis) and chronic sinusitis. Afzelius later observed that male patients with the syndrome were infertile because of defects in sperm flagella motility. Putting this together with the airway defects, he demonstrated that the primary defect was in cilia function. The cilia lacked the dynein motor proteins, leading to lack of airway clearance, frequent respiratory tract infections, and immotile spermatozoa. The relationship between cilia function and left-right patterning, however, remained a mystery for many years. Finally Nonaka et al, investigating a mutation that caused randomization of left-right patterning in mice, noticed at a spot on the midline of the early embryo where the epithelium had cilia, and postulated that cilia-driven “nodal flow” was the cause of left-right patterning.

Genetic studies have confirmed that mutations in ciliary dynein motor proteins are responsible. Mutations in dynein axonemal intermediate chain 1 (DNAI1) and dynein heavy chain have been identified in human cohorts. Notably, loss of cilia function does not always lead to the full Kartagener’s syndrome: only 50% have situs inversus. Left-right patterning still occurs; it is just left to random chance which side will be which.

Leftward fluid flow has since been convincingly established as the causal event in left-right patterning through a series of experiments in mouse embryos. Artificially imposing rightward flow during gestation causes reversal of the left-right pattern in wild-type mice. Furthermore, imposing an artificial leftward flow returns left-right patterning to normal in mice with defective nodal cilia. But how do rotating cilia cause flow across the surface in one direction? The cilia are not perpendicular to the surface—they are sufficiently tilted in the posterior direction so that they brush against the posterior surface and fail to generate much force during this part of their rotation. The net result is that fluid is driven in only one direction, although at a low velocity.

Soon after flow begins, the TGF-family signaling molecules, nodal, is expressed more strongly on the left side of the embryo, leading to a cascade that rapidly induces nodal and other factors throughout the left side of the embryo. However, the mechanism the cells use to sense flow and differentially induce nodal remains unresolved. One possibility is that the directional flow redistributes morphogens released from the node, such as retinoic acid or sonic hedgehog, to the adjacent cells on the left. However, there is another intriguing hypothesis. The node has a separate set of cilia that are nonmotile, and there is evidence these cilia are specifically designed to sense the shear stress created by fluid flow. The cilia are coupled to a calcium channel in sensing fluid flow during morphogenesis of the kidneys.

Fluid Shear Stress Is Critical for Remodeling Vessels During Vasculogenesis

One of the most critical effects of fluid flow on morphogenesis is its control of vasculogenesis, which refers to the development of new blood vessels directly from precursor cells. While the importance of fluid flow has long been postulated, only recently has it emerged that fluid shear stress is specifically required for remodeling in vasculogenesis.

In contrast, the process of angiogenesis in adults, in which new blood vessels sprout off from existing vasculature, was classically thought to be independent of fluid flow. Endothelial cells can form tubular structures in the absence of fluid flowing through them. Angiogenesis seems largely controlled by signaling integrated through the hypoxia-induced factors, and is stimulated by low local oxygen tension as sensed by the prolyl hydroxylases.

The importance of fluid flow for proper vascular function in adults is also well recognized. Mature endothelial cells respond rapidly to fluid flow by multiple mechanisms, and disturbed fluid flow promotes endothelial dysfunction. Furthermore, fluid flow is required for formation of the lymphatic vessels. The relatively low velocity of fluid flow in the extracellular space allows flow-induced gradients of growth factors such as VEGF to form, and these gradients promote the alignment of lymphatic vessels in the direction of the flow.

A major site of vasculogenesis during mouse development is outside the embryo itself, where cells migrate to form the “blood islands” of the yolk sac primary vascular plexus. Soon after the heart begins to produce a fetal circulation (8.5 to 9.5 dpc), these islands must be remodeled from largely undirected passages into a vascular tree with branching arteries and veins (Figure 1, second row). Initially, it was not clear whether precursor cells formed an artery or a vein based on their environment or whether this was genetically predetermined. Eichmann and colleagues answered this question by demonstrating that stopping flow to the yolk sac prevents remodeling even though the blood islands continue to grow.

Fluid flow is important for both transport and mechanical processes. One might expect that stopping fluid flow prevents vascular remodeling by preventing transport of nutrients, oxygen, or other morphogenic factors. Surprisingly, Lucitti et al have recently shown that fluid shear stress is the critical requirement. By physically sequestering the erythroblasts, the hematocrit is greatly reduced, and the preexisting yolk sac vessels do not mature into an organized vascular tree (Figure 2D through 2F). However, artificially increasing the fluid viscosity by adding starch rescues blood vessel formation (Figure 2G through 2I), demonstrating that it is shear stress and not transport that is required for vascular remodeling during vasculogenesis.

Recent in vitro studies have also suggested an important role for fluid flow in differentiation of mesenchymal precursor cells into endothelial cells. In a mesenchymal progenitor cell line (C3H/10T1/2), cells plated on a collagen-based gel and exposed to fluid flow could spontaneously form into
tubular structures.\textsuperscript{43} As the sharp distinction between embryonic vasculogenesis and adult angiogenesis becomes more blurred, it has also become evident that angiogenesis involves more than just sensing oxygen levels. Fluid flow may also play a role in some circumstances. However, there are other mechanical inputs that are known to be important: angiogenesis is directed in part through contraction of the actin cytoskeleton driven by nonmuscle myosins.\textsuperscript{44}

Regulation of Morphogenesis by Cellular Contractility

Nonmuscle Myosin Causes Shape Change in Gastrulation

Control of actin cytoskeleton contractility is a fundamental mechanism for molding shape changes in development. A well-studied example of this effect is gastrulation in the fruit fly, \textit{D melanogaster}. Gastrulation is the first major shape change of the developing embryo, and cell-directed mechanical forces have a critical role.\textsuperscript{11} Before gastrulation, the middle of the \textit{D melanogaster} embryo appears roughly as a cylindrical tube composed of a single layer of cells. Gastrulation begins when a furrow appears across the length of the tube, and the furrow deepens until it has created a new layer of cells by folding within itself (Figure 1, third row).

The appearance of the furrow, it had long been hypothesized, would be neatly explained as a purely mechanical event: just like the formation of the dermal ridges, increasing compression in the cell layer could cause it to buckle in the direction with the least resistance.\textsuperscript{45} Here, instead of buckling upwards to form a ridge, the cells buckle inwards to form a furrow. This raises two questions: how does the cell layer generate compression, and how does it control the direction in which the layer buckles?

Early biologists proposed that the compression was created as cells divided and grew while being restrained from expanding in size.\textsuperscript{46} However, compressive forces created by cell proliferation alone cannot determine whether the tissue folds inwards or outwards. To ensure the layer folds inwards, the inner side of the cell layer must be less stiff than the outer side, so there must be a mechanism to regulate stiffness. Eventually, it was realized that the primary regulator of mechanical forces in the cell layer is not cell proliferation, but contraction of the actin cytoskeleton by the type II nonmuscle myosins.

The role of muscle-specific type II myosin motors in generating muscle contractions is well known. However, all cells contain related type II myosins, referred to as the nonmuscle myosins, that bind to actin and control contraction of the cytoskeleton. Initially found to be required for cell division, the roles of nonmuscle myosins continue to expand into many aspects of cell biology.\textsuperscript{47}

In Drosophila, the requirement of the nonmuscle myosins for early events in embryogenesis was first demonstrated by

\begin{figure}
\centering
\includegraphics[width=\textwidth]{image.png}
\caption{Vessel remodeling during vasculogenesis requires fluid shear stress. In the mouse, vasculogenesis begins largely outside the embryo in the yolk sac. The yolk sac vessels mature from largely undirected passages into branched arteries and veins soon after the beginning of the fetal heart function (A through C). The normal mature vessels can be seen both by injection of fluorescent-labeled dextran (B, "TR-Dextran") and by imaging erythroblasts genetically labeled with green fluorescent protein (C, "Tg(\epsilon-globin-GFP)"). Physically sequestering the erythroblasts outside the embryo completely prevents remodeling of yolk sac vessels (D through F, low hematocrit). However, increasing the viscosity of the fluid by adding starch rescues vasculogenesis, even though the erythroblasts remain sequestered (H through I, "Low Hematocrit+Hetastarch"). The results demonstrate that heart function initiates remodeling by fluid shear stress, not by transport of oxygen or other factors in the blood. Reproduced from Lucitti et al\textsuperscript{42} with permission of the Company of Biologists.}
\end{figure}
characterization of flies with deletion of the nonmuscle myosin II heavy chain gene. Flies have only one isoform of nonmuscle myosin II heavy chain, and the deletion is lethal because of failure of an event late in gastrulation known as dorsal closure. Gene deletion, however, does not capture the importance of myosin for the earliest events in morphogenesis, because the embryos carry over enough maternal myosin to make it through most of gastrulation without synthesizing new myosin.

Visualizing the location of nonmuscle myosin by immunostaining strongly but indirectly suggests the role of myosin in regulating tension and compression during gastrulation. Before gastrulation begins, myosin is located in a sharp ring around the inner circumference of the cells. Just as the initial furrow appears, myosin disappears from the inner surface and relocates to the outer surface precisely where the furrow is forming. The presumed mechanical effect is that myosin is causing the outer (apical) surface to contract, creating compression of the cells simultaneously with the loss of tension around the inner surface. This causes the cell layer to buckle inwards and deepens the furrow on the outside, leading to invagination.

Further evidence for control of gastrulation by nonmuscle myosins is that the gastrulation requires signaling through the Rho family of GTPases, which control myosin contractile activity. Nonmuscle myosin II, unlike skeletal and cardiac myosin II, is regulated by phosphorylation of one of the two myosin light chains. One of the major kinases that phosphorylates myosin light chain and leads to increased myosin activity is Rho-associated coiled-coil protein kinase (Rock). Rock is regulated by association with the small GTPase RhoA, and Rho GTases are regulated by guanine nucleotide exchange factors (RhoGEFs). In Drosophila, all 3 levels of control are required for gastrulation: gene deletion of the homologs for Rock, RhoA, and RhoGEF2 each prevent dorsal closure. Contraction of the apical side of epithelial cells by relocation of RhoGEFs, Rho family GTases, and nonmuscle myosin is a fundamental theme for morphogenesis that recurs frequently in subsequent epithelial cell shape changes. Depending on the pattern of activation and the mechanical properties of the surroundings, apical contraction can lead to shape change, buckling, folding, or pit formation.

Dorsal Closure: Visualizing the Role of Myosin Contraction

Toward the end of gastrulation in Drosophila, two sheets of epithelial cells on the outer surface of the embryo move toward each other and meet at the dorsal midline, where they neatly fuse together. This event, known as dorsal closure, is a major test of the organism’s ability to produce and regulate the mechanical forces produced by the cells. As noted above, flies with null mutations in nonmuscle myosin heavy chain are unable to complete dorsal closure. More generally, of gene deletions that are lethal to embryos, failure to complete dorsal closure is a frequent cause of death for a surprisingly wide range of genes. Furthermore, the events of dorsal closure may provide insights into the regenerative processes involved in wound healing.

Dorsal closure has thus become an important model for understanding the role of myosin contraction, its relative importance, and its regulation. Studying the precise role of myosin contraction in vivo is challenging because there are few ways to directly visualize the forces produced by myosin in any particular cell. However, unlike the initial steps of gastrulation, dorsal closure involves only epithelial sheets on the outer surface, so the events are relatively easy to visualize. Recently, innovative genetic and engineering approaches have been applied to directly observing and measuring the mechanical forces involved in dorsal closure.

The importance of the nonmuscle myosin II heavy chain homolog (zipper) is visually clear in transgenic flies expressing nonmuscle myosin-GFP (Figure 3). The epithelial cell layer first meets at either side, forming an eye-shaped gap. Myosin-GFP is localized to the leading edge of the cells, suggesting that it acts as a drawstring that pulls the edges of the cells together as it contracts. However, to directly visualize the tension generated by nonmuscle myosin contraction, Franke et al took advantage of a fortuitous observation in a myosin-null embryo that was also transgenic for overexpression of myosin-GFP. Although the transgenic myosin-GFP is driven by the same promoter in all cells, a few cells do not express any visible myosin-GFP and therefore remain null for myosin. The motions of these cells during dorsal closure were then followed by real-time fluorescence imaging. As the “eye” closes up, gaps are seen where nonexpressing cells are located on the leading edge of the drawstring. As closure progresses, the gaps become larger—visual evidence that myosin contraction creates tension that pulls on the adjacent cells in a ring around the opening.

Thus, nonmuscle myosin drives contraction of the purse-string at the leading edge, and this contractile force is required for closure. But myosin plays other roles in cells, and there are other sources for the forces driving closure. A clear understanding of the process requires quantifying the relative roles of all these forces. Remarkably, Hutson et al accomplished this feat: they measured forces by making incisions in the epithelium with a laser and observing how far and how quickly the cells snap back. These empirical data were then used to create a model for the relative contributions of various forces. Indeed, contraction of the leading edge myosin provided most of the forces for dorsal closure. However, the most astonishing aspect of the process is the extent of mechanical redundancy. Even without the contraction of myosin at the corners of the eye, adhesion forces from the underlying cell layer are enough to guide closure of the opening. This result echoes the conclusions of Franke et al that expression of nonmuscle myosin in either the leading edge cells, or the cells in the underlying layer, is sufficient for dorsal closure.

Cardiogenesis in Vertebrates

Although nonmuscle myosin is also required for cardiogenesis, identifying the specific roles of mechanical forces in a mechanically functional tissue is more challenging than the previous examples. Nevertheless, understanding these roles has clinical implications because abnormal development of...
mechanical forces may be a major contributor to congenital heart defects. The interplay of developmental form and mechanical function during cardiogenesis is remarkable. As the heart tube begins to function, the heart experiences both mechanical deformations and fluid flow. At the same time, developmental processes are coordinating major changes in form. Beginning as a linear tube in which the ventricles are upstream of the future atria, the tube loops around, fuses, and reconnects itself—all while continuing to function as a pump.58,59

Targeted gene deletion in mice shows that nonmuscle myosin is required for cardiogenesis. Unlike Drosophila, vertebrates express three different genes for nonmuscle myosin II heavy chain (NMHC2 A, B, and C).47 Most cells express both the A and B isoforms. In cardiomyocytes, however, expression is largely restricted to the B isoform. Mice with deletion of Nmhc2b can gastrulate but die as embryos due to failure of cardiogenesis.60 The mice have multiple cardiac malformations including ventricular septal defects, aortic root malformation, and an inability to form organized cardiac muscle. Thus, even though cardiomyocytes have normal amounts of the myosin that produces contraction of cardiac muscle, nonmuscle myosin is still required for normal patterning and morphogenesis of the heart.

The development of the heart endocardial cushions is particularly important for investigators of congenital heart defects. The endocardial cushions are the forerunners of the heart valves as well as parts of the septae. Endocardial epithelial cells transdifferentiate into mesenchymal cells and migrate into bulges in the cardiac jelly to form the cushions.51,62 Yet the forces involved in cushion development have resisted simple characterization. One major question has been whether the forces of fluid flow are required for inducing the endocardial cushions, or whether they develop through an independent program. Evidence supporting both views has recently been observed.58,63–65

Hove and colleagues mechanically blocked flow through the early zebrafish heart tube and demonstrated that without fluid flow, the cushions failed to develop.63 Although blocking flow may change local fluid pressures as well, blocking flow either into the heart tube or out of the heart tube produced similar defects, suggesting that the lack of fluid shear stress was the critical factor. However, blocking flow also prevented cardiac looping, an earlier event in heart development, so it is possible that the lack of endocardial cushions was secondary to generally defective cardiogenesis.

On the other hand, Bartman and colleagues used a pharmacological inhibitor of myosin ATPase activity and observed that the endocardial cushion development failed to develop.63 Although blocking flow may change local fluid pressures as well, blocking flow either into the heart tube or out of the heart tube produced similar defects, suggesting that the lack of fluid shear stress was the critical factor. However, blocking flow also prevented cardiac looping, an earlier event in heart development, so it is possible that the lack of endocardial cushions was secondary to generally defective cardiogenesis.

On live fluorescent imaging of the flies during dorsal closure, these nonexpressing cells are seen as gaps in the drawstring surrounding the opening between the epithelial cell sheets (A, arrows). As closure progresses, the space between GFP–myosin expressing cells clearly lengthens (A, arrow labeled with an asterisk; C, arrows labeled 1 and 2), visually demonstrating that contraction of myosin in the drawstring pulls on the neighboring cells to direct dorsal closure. Reproduced with permission from Elsevier, Franke et al.56
possible that the remaining contractile function was still sufficient to produce fluid shear stress on the endocardium. Although the specific roles of forces such as shear stress continue to be debated, it seems clear that many steps in cardiogenesis can be viewed as an integrated feedback loop: alterations in shape change heart function, which changes the forces experienced by the cells, stimulating them to modify heart function both by secreting matrix and by continuing to change the heart shape. Such biomechanical feedback loops are common throughout the development of the load-bearing and force-generating tissues.

In Vitro Cell Differentiation Is Controlled by Nonmuscle Myosins in Multiple Lineages

A final example of developmental processes controlled by myosin contractility is the in vitro differentiation of stem and precursor cells. Despite the nonphysiological environment, a major lesson of in vitro cell culture is that the mechanical properties of the materials surrounding the precursor cells are tightly linked to Rho family GTPase signaling and nonmuscle myosin contraction within the cell. Remarkably, manipulating any one of these three variables can be sufficient to change the differentiation pathway of a precursor cell. These insights are likely critical for guiding development of better tissue engineering and stem cell therapeutics.

It has been known since the 1970s that differentiation of precursor cells into chondrocytes and adipocytes in vitro is promoted by high cell density. For example, embryonic mesodermal cells isolated from chick limb buds will spontaneously undergo chondrogenesis when cultured at a very high density. Similarly, subsets of 3T3 fibroblastic cells will spontaneously accumulate lipid when kept in high-density confluent monolayers. Interestingly, disrupting the actin cytoskeleton by treatment with cytochalasin also promotes differentiation of precursor cells into chondrocytes and adipocytes. What mechanism links both cytoskeletal contractility and high cell density to cell differentiation? One might assume that the answer is simply related to cell–cell contact: by disrupting actin or culturing at high cell density, cells are prevented from spreading, instead forming rounder cells with more cell–cell contacts.

Recent experiments suggest that cell–cell contact is not the answer. Using micropatterning techniques to precisely manipulate the area available for cells to attach to a substrate, in the absence of any cell–cell contact, McBeath et al showed that isolated human mesenchymal stem cells can change their differentiation pathway based solely on whether they are allowed to attach to a small fibronectin area (32×32 μm) or a large one (100×100 μm square). Cultured in the same mixture of adipogenic and osteogenic factors, stem cells grown on small fibronectin islands remain round and differentiate into adipocytes, whereas stem cells grown on the large fibronectin islands spread out over the entire island surface.
form stress fibers, and differentiate into osteoblasts. At least for several mesenchymal cell lineages, myosin contraction of the actin cytoskeleton, the area of interaction with hard substrates, and cell shape seem to play primary roles in differentiation (Figure 4).

These experiments leave two major possibilities for the mechanism controlling differentiation in vitro. One possibility is that differentiation could depend on recapitulating the normal three-dimensional round shape of the cell. Indeed primary chondrocytes stop producing cartilage matrix when cultured in vitro but maintain their phenotype when kept round either by treating with cytochalasin or by resuspending them in a soft gel. Cell shape could affect spatial patterns of signaling through integrin or cadherin complexes on the cell surface. A second major possibility is that there could be a largely mechanical effect on differentiation: high cell density and round shape both minimize contact with the hard tissue culture surfaces, reducing the area available for stimulating formation of focal adhesions, stress fibers, and myosin contraction.

Further supporting the role of myosin contraction is that signaling through the Rho family GTPases also controls differentiation in a consistent manner. Overexpression of RhoA or Rock1, which stimulate myosin contraction, promotes osteogenesis but inhibits adipogenesis and chondrogenesis. Conversely, overexpression of Rac, which tends to oppose RhoA action, promotes differentiation into chondrocytes and adipocytes.

Why might cells use formation of intracellular mechanical stress to control differentiation? Is this just an artifact of in vitro culture or is there a reason for myosin contractility to control differentiation in vivo? It is not related to the need to resist mechanical stresses as a differentiated cell: chondrocytes and osteoblasts both become part of load-bearing structures, yet myosin contraction drives precursor cell differentiation in opposite directions.

The normal developmental program for making endochondral bone suggests one reason for the opposed effects of myosin contraction on differentiation into chondrocytes and osteoblasts. Chondrocytes first form directly from coalescing embryonic mesodermal cells in the absence of significant matrix or mechanical loading. The chondrocytes then synthesize a matrix, hypertrophy, and die by apoptosis. It is to the already hardened matrix that osteoblasts migrate and mature. This suggests the hypothesis that precursor cells might use the stiffness of their surroundings as a critical cue for determining differentiation in vivo.

Supporting this hypothesis, several investigators have reported that cellular phenotypes do indeed change in response to the stiffness of their physiological surroundings. For example, fibroblasts form stress fibers when in contact with stiff surroundings, regardless of whether they are grown on two-dimensional tissue culture surfaces or seeded in three-dimensional collagen gels. This in vitro behavior may recapitulate the physiological response of fibroblasts in healing skin wounds: as the wound begins to heal, tension is generated, which promotes differentiation of fibroblasts into myofibroblasts and generates more contraction in turn.

Engler et al have recently tested this hypothesis more broadly by investigating whether stem cell differentiation can be regulated by the stiffness of the surroundings in vitro. Human mesenchymal stem cells were cultured as a monolayer on collagen-I–coated gels with varying stiffnesses. Under identical culture conditions except for the stiffness of the gel, stem cells were induced to express early markers of differentiation into neurons (for the softest gel, with an elastic modulus of 0.1 to 1 kPa), myocytes (8 to 17 kPa), or osteoblasts (25 to 40 kPa). The myosin contractile state of the cells adapted to the stiffness of the gel: on the hardest surface, the cells spread out the most, and developed focal adhesions. Importantly, Engler et al provide evidence that the stiffness promoting stem cell differentiation into each lineage corresponded to the conditions normally present during differentiation of that lineage. For example, the gel stiffness that maximized expression of osteoblast markers was similar to the stiffness of matrix laid down by hypertrophic chondrocytes.

Further investigation of how the mechanical environment changes during muscle development may explain aspects of the less straightforward results regarding differentiation of mesenchymal stem cells into myotubes. Differentiation of myoblasts into myotubes in vitro is prevented by cytochalasin and requires integrin-mediated phosphorylation of focal adhesion kinase, RhoA signaling, and myosin contraction. On the other hand, RhoA or Rock1 can also inhibit myogenesis by preventing cell fusion and myotube formation. Myogenesis probably requires a balance of signals regulating contractility that changes during the multi-step differentiation process. Similarly, the differentiation of stem cells into cardiomyocytes requires a balance of Rho and Rac signaling that varies with time.

In vivo, primary myoblasts are induced directly from mesodermal cells in the absence of significant mechanical stresses. In contrast, after the myoblasts migrate to the site of muscle formation and fuse into myotubes, the developing muscles attach to tendons and are subjected to significant loads. Therefore, secondary myogenesis, as well as adult myogenesis from satellite cells, will experience an entirely different mechanical environment, and may require different programs for regulating contractile state during maturation.

Although the hypothesis that cells use myosin contraction to adjust their developmental pathway as a function of extracellular substrate stiffness is attractive as a unifying concept, its full extent is not yet clear. It will be interesting to test this hypothesis in a wider range of differentiation pathways and situations. Also, basic questions about the pathway from sensing matrix stiffness to myosin contraction and Rho signaling remain unanswered. Extracellular matrix stiffness may be sensed by signaling through integrin or cadherin-catenin events that activate Rho signaling and then myosin contraction. On the other hand, intracellular myosin contractile forces must be balanced by forces from the extracellular matrix. Therefore matrix stiffness must limit myosin contraction directly as well, because soft surroundings physically do not support much contraction. Most likely, as with most mechanotransduction signaling path-
ways, multiple pathways participate, and feedback mecha-
nisms play important roles.

Sources of Funding
R.T. Lee is supported in part by NIH grant R01-HL081404; P. Patwari is supported in part by NIH grant K25-HL081523 and a Lerner Cardiovascular Award.

Disclosures
None.

References
4. Stevenson J. Man’s murder charge dismissed because DNA couldn’t tell him from identical twin; A genetic loophole and the suspects’ silence made it impossible to determine who was involved in the July 2000 beating death. The Herald-Sun (Durham, NC). Feb 1, 2001.C1.


Butcher JT, McQuinn TC, Sedmera D, Turner D, Markwald RR. Transitions in early embryonic atrioventricular valvular function correspond with changes in cushion biomechanics that are predictable by tissue composition. Circ Res. 2007;100:1503–1511.


Patwari and Lee Mechanical Control of Morphogenesis 243
Mechanical Control of Tissue Morphogenesis
Parth Patwari and Richard T. Lee

Circ Res. 2008;103:234-243
doi: 10.1161/CIRCRESAHA.108.175331
Circulation Research is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2008 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7330. Online ISSN: 1524-4571

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://circres.ahajournals.org/content/103/3/234

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Circulation Research can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Circulation Research is online at:
http://circres.ahajournals.org/subscriptions/