Toll-Like Receptor 2 Mediates Apolipoprotein CIII–Induced Monocyte Activation

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Abstract—Apolipoprotein (apo)CIII predicts risk for coronary heart disease. We recently reported that apoCIII directly activates human monocytes. Recent evidence indicates that toll-like receptor (TLR)2 can contribute to atherogenesis through transduction of inflammatory signals. Here, we tested the hypothesis that apoCIII activates human monocyteoid THP-1 cells through TLR2. ApoCIII induced the association of TLR2 with myeloid differentiation factor 88, activated nuclear factor (NF)-κB in THP-1 cells, and increased their adhesion to human umbilical vein endothelial cells (HUVECs). Anti-TLR2 blocking antibody, but not anti-TLR4 blocking antibody or isotype-matched IgG, inhibited these processes (P<0.05). ApoCIII bound with high affinity to human recombinant TLR2 protein and showed a significantly higher (P<0.05) and saturable binding to 293 cells overexpressing human TLR2 than to parental 293 cells with no endogenous TLR2. Overexpression of TLR2 in 293 cells augmented apoCIII-induced NF-κB activation and β1 integrin expression, processes inhibited by anti-apoCIII antibody as well as anti-TLR2 antibody. Exposure of peripheral blood monocytes isolated from C57BL/6 (wild-type) mice to apoCIII activated their NF-κB and increased their adhesiveness to HUVECs. In contrast, apoCIII did not activate monocytes from TLR2-deficient mice. Finally, intravenous administration to C57BL/6 mice of apoCIII-rich very-low-density lipoprotein (VLDL), but not of apoCIII-deficient VLDL, activated monocytes and increased their adhesiveness to HUVECs, processes attenuated by anti-TLR2 or anti-apoCIII antibody. ApoCIII-rich VLDL did not activate monocytes from TLR2-deficient mice. In conclusion, apoCIII activated monocytes at least partly through a TLR2-dependent pathway. The present study identifies a novel mechanism for proinflammatory and proatherogenic effects of apoCIII and a role for TLR2 in atherosclerosis induced by atherogenic lipoproteins. (Circ Res. 2008;103:1402-1409.)

Key Words: apolipoprotein ■ atherosclerosis ■ inflammation ■ monocyte ■ Toll-like receptor

Toll-like receptors (TLRs) contribute importantly to neutrophil-mediated responses during inflammation. Exogenous ligands such as lipopolysaccharide (LPS) and peptidoglycan, as well as endogenous factors produced on stress or cell damage, eg, heat shock proteins, activate these pattern-recognition receptors. The recent identification of mammalian TLRs as principal sensors of the innate immune system provides a mechanistic link among infection, inflammation, and atherosclerosis.1 Moreover, activation of TLRs by exogenous ligands can provoke sterile inflammation linked to atherogenesis susceptibility.2,3 Among TLRs, TLR2 and TLR4 may particularly in the inflammatory response and in atherosclerosis. Recent studies in fat-fed mice have shown that TLR4, TLR2, and myeloid differentiation factor 88 (MyD88) contribute to atherosclerotic plaque accumulation induced by hyperlipidemia.1,4 Michelsen et al found that a loss of TLR4 or its adaptor protein MyD88 reduces disease severity in atherosclerosis-prone apolipoprotein (apo)E-deficient mice.4 Mullick et al1 reported that, in the absence of any known exogenous TLR2 agonist, complete deficiency of TLR2 in LDL receptor–deficient mice reduced atherosclerosis, whereas loss of TLR2 expression in bone marrow–derived cells did not have an impact on disease. These results suggest that unknown endogenous TLR agonists impact atherosclerotic disease. Although the mechanism(s) by which TLRs contribute to atherogenesis remains obscure, we and others previously reported that TLR2 and TLR4 play a role in the enhancement of monocyte–endothelial interaction, a crucial step throughout atherogenesis.5–7

Plasma levels of apoCIII independently predicts risk for coronary heart disease.8 We recently demonstrated that apoB lipoproteins that contain multiple copies of apoCIII, but not apoCIII-deficient apoB lipoproteins, induce human monocyte adhesion to vascular endothelial cells (ECs)9,10 via a protein
kinase (PKCδ) and nuclear factor (NF)-κB-dependent mechanism. Interestingly, apoCIII alone also had similar effects on vascular cells, suggesting that these effects of apoCIII-containing apolipoproteins are mediated by apoCIII rather than by other apolipoproteins or lipids in very-low-density lipoprotein (VLDL) or LDL and not by apoB/E receptors on monocytes. We thus hypothesized that TLRs mediate apoCIII signaling and contribute to proinflammatory properties for apoCIII. The present study determined whether TLR2 or TLR4 participates in apoCIII-induced activation of monocytes and their adhesion to ECs in vitro and in vivo.

Materials and Methods

Animals and Cells

Seven-week-old male C57BL/6 (wild-type) mice (Oriental Yeast, Tokyo, Japan) or TLR2-deficient mice consumed a standard diet (CLEA Japan, Tokyo, Japan). Food and water were provided ad libitum. Human peripheral blood monocytes were collected under a protocol approved by the Human Research Committee of the Brigham and Women’s Hospital and were cultured as described previously. Human umbilical vein endothelial cells (HUVECs), purchased from Academy Biomedical (Houston, Tex). They were purified from human plasma using HPLC and immunoaffinity column chromatography. Their purity was >99.0%, as determined by SDS-PAGE. Endotoxin levels in apolipoproteins measured using a chromogenic Limulus amebocyte lysate test (Associates of Cape Cod, East Falmouth, Mass) were <0.03 endotoxin unit (EU)/mL. Free fatty acid levels in apolipoproteins determined enzymatically were <20 nmol/L. Antibodies used in the present study were as follows: anti-β1 integrin, anti-MyD88, anti-Rac1, anti-NF-κB p65, fluorescein isothiocyanate (FITC)-conjugated NF-κB p65, anti-CD14, and anti-β-actin (Santa Cruz Biotechnology, Santa Cruz, Calif); anti–PKCα (BD Biosciences, San Jose, Calif); anti-apoCIII (Academy Biomedical); anti–TLR2 and anti–TLR4 (Imgenex, San Diego, Calif); and anti–NF-κB p65 (pS276) (Rockland, Gilbertsville, Pa). Polymyxin B, peptidoglycan (Staphylococcus aureus), and LPS (Escherichia coli O26:B6) were purchased from Sigma.

Static Adhesion Assay

HUVECs seeded on 1% gelatin-coated 96-well culture plates were maintained for 2 days to allow the formation of a confluent monolayer, and stimulated with interleukin 1β (Genzyme, Cambridge, Mass) at 10 U/mL for 4 hours before adhesion assay. After THP-1 cells or freshly isolated mice peripheral blood monocytes were incubated with apoCIII or reagents as indicated, cells were labeled with BCECF-AM (Calbiochem, La Jolla, Calif), were incubated with or without apoCIII (100 μg/mL) for 2 hours. *P<0.01 vs apoCIII (-)/antibodies (-), #P<0.05 vs apoCIII (+)/antibodies (-). IB and IP indicate immunoblotting and immunoprecipitation, respectively. Blots represent 4 to 6 independent experiments using apoCIII from 4 to 6 different donors that yielded similar results (B through E).

Figure 1. TLR2 mediates apoCIII-induced THP-1 cell activation. A through D, THP-1 cells were pretreated with indicated antibodies (μg/mL) for 30 minutes and then incubated with or without apoCIII (100 μg/mL) for 8 hours (A and D) or 2 hours (B and C). mem indicates the membrane fraction of the cell lysates (B). E, THP-1 cells were incubated with or without apoCIII (100 μg/mL) for 2 hours. *P<0.01 vs apoCIII (-)/antibodies (-), #P<0.05 vs apoCIII (+)/antibodies (-). IB and IP indicate immunoblotting and immunoprecipitation, respectively. Blots represent 4 to 6 independent experiments using apoCIII from 4 to 6 different donors that yielded similar results (B through E).

Reagents

Human apoC1, -CII, -CIII, and -E were purchased from Academy Biomedical (Houston, Tex). They were purified from human plasma using HPLC and immunoaffinity column chromatography. Their purity was >99.0%, as determined by SDS-PAGE. Endotoxin levels in apolipoproteins measured using a chromogenic Limulus amebocyte lysate test (Associates of Cape Cod, East Falmouth, Mass) were <0.03 endotoxin unit (EU)/mL. Free fatty acid levels in apolipoproteins determined enzymatically were <20 nmol/L. Antibodies used in the present study were as follows: anti-β1 integrin, anti-MyD88, anti-Rac1, anti-NF-κB p65, fluorescein isothiocyanate (FITC)-conjugated NF-κB p65, anti-CD14, and anti-β-actin (Santa Cruz Biotechnolog, Santa Cruz, Calif); anti–PKCα (BD Biosciences, San Jose, Calif); anti-apoCIII (Academy Biomedical); anti–TLR2 and anti–TLR4 (Imgenex, San Diego, Calif); and anti–NF-κB p65 (pS276) (Rockland, Gilbertsville, Pa). Polymyxin B, peptidoglycan (Staphylococcus aureus), and LPS (Escherichia coli O26:B6) were purchased from Sigma.
lipoproteins and reagents was examined by staining with 0.25% trypan blue solution.

Immunoblotting and Immunoprecipitation
Total cell lysates and the membrane fraction of the indicated cells (1×10^6) were prepared as described previously. An equal amount of protein (10 μg) from each fraction was subjected to 12% SDS-PAGE and transferred to poly(vinylidene difluoride) membrane. Immunoreactive proteins in the membrane were detected using indicated antibodies with an enhanced chemiluminescence (ECL) plus (Amersham Biosciences, Piscataway, NJ). Activation of PKC was examined by detecting the membrane-bound protein that translocated from cytosol fraction.

For immunoprecipitation, a cell lysate from THP-1 cells was incubated with anti-TLR2 antibody. Then, 50 μL of anti-IgG affinity gel (MP Biomedicals, Solon, Ohio) was added for an additional 60 minutes, after which the immunocomplexes were collected and resuspended in SDS-PAGE sample buffer for immunoprecipitation as described previously.

Protein-Binding Studies
Ninety-six-well tissue culture plates were coated with or without recombinant TLR2/Fc chimera protein, TLR4/Fc chimera protein (R&D Systems, Minneapolis, Minn) at 2 μg per well. ApoCIII proteins were labeled with FITC using EZ-Label FITC protein-labeling kit according to the instructions of the manufacturer (Pierce, Rockford, Ill). After 96-well tissue culture plates were blocked with the albumin (Sigma), FITC-labeled apoCIII (100 μg/mL) was added to 96-well plates and incubated for 10 minutes at 4°C. Some experiments included nonlabeled apoCIII or other potential competitors. After extensive washing, the FITC associated with 24-well tissue culture plates was measured using a chromogenic Limulus amebocyte lysate test (Associates of Cape Cod) were <0.03 EU/mL.

Lipoprotein Preparation
Blood was drawn in tubes containing EDTA from newly diagnosed hypertriglyceridemic patients at 12-hours fasting. The study was approved by the Institutional Review Board of the Tokyo Medical and Dental University. The subjects had no other serious diseases and had not taken cardiovascular medications, lipid-lowering medications, pharmacological doses of antioxidants, or estrogen for more than 14 days. ApoCIII-rich VLDL (VLDL CIII) and apoCIII-deficient VLDL (VLDL CIII') were isolated from plasma as described previously. Endotoxin levels in the lipoprotein fractions measured using a chromogenic Limulus amebocyte lysate test were <0.03 EU/mL.

In Vivo Stimulation
For in vivo stimulation, the mice were injected intravenously with VLDL preparations (500 μg apoB per mouse) from femoral veins. After 2 hours, monocytes were isolated as described above for immunoblotting or immunofluorescence microscopy. For static adhesion assay, they were isolated 6 hours after VLDL injection.

Immunofluorescence Microscopy
For NF-κB p65 staining, isolated mouse monocytes were fixed with 4% formaldehyde for 30 minutes and then treated with 0.05% Triton X-100 for 5 minutes. The cells were incubated with FITC-conjugated NF-κB p65 antibody (1:200) for 45 minutes. The cells were rinsed and mounted onto slides, then analyzed and imaged using a fluorescent microscope (Olympus) with a 400-fold magnification.

For cell-binding studies, 293 cells were cultured in 6-well plates and then preincubated for 30 minutes at 4°C. The cultures were then incubated with the indicated amounts of FITC-labeled apoCIII alone (100 μg/mL) and in the presence of nonlabeled apoCIII or other potential competitors for 30 minutes at 4°C before extensive washing. Cells were dissolved in 0.1 N NaOH before the measurement of cell-associated FITC using CytoFluor II.
Statistical Analysis
Results are given as the means±SD. Data were analyzed using unpaired t test or 2-way ANOVA, with a value of P<0.05 considered significant.

Results
Involvement of TLR2 in ApoCIII-Induced THP-1 Cell Activation
ApoCIII induces the adhesion of THP-1 cells to vascular endothelial cells through activation of PKCα and NF-κB.11 We first examined whether these processes involve TLR2 or TLR4 using blocking antibodies to these receptors. Anti-TLR2 antibody treatment of THP-1 cells inhibited apoCIII-induced THP-1 cell adhesion in a concentration-dependent manner. Neither anti-TLR4 nor an irrelevant isotype-matched IgG inhibited apoCIII-induced THP-1 cell adhesion (Figure 1A). Anti-TLR2 antibody also attenuated apoCIII-induced PKCα activation and NF-κB activation, as determined by PKCα membrane translocation and NF-κB p65 phosphorylation, respectively (Figure 1B and 1C), whereas anti-TLR4 antibody or isotype-matched IgG had minimal effects on these processes (data not shown). We also reported that apoCIII increases NF-κB activation and NF-κB phosphorylation, respectively (Figure 1D). Recent studies showed that CD14 physically interacts with TLR2 and facilitates ligand binding. Miller et al reported that minimally modified LDL binds to macrophage CD14.17 We thus examined the involvement of CD14 in apoCIII-induced monocyte activation. Anti-CD14 binding blocking antibody had minimal effect on these processes (data not shown). We also performed additional experiments using known ligands for TLRs to validate the efficacy and specificity of these antibodies. Anti-TLR2 antibody inhibited the association of apoCIII with TLR2 and its adhesion to HUVECs.9 Anti-TLR2 antibody but not anti-TLR4 antibody attenuated these processes (Figure 2A through 2C).

Association of ApoCIII With TLR2
We then examined the binding of FITC-labeled apoCIII with TLR2 protein−coated plastic plates. ApoCIII bound to TLR2 protein with significantly higher affinity than albumin or TLR4 protein (Figure 3A). Pretreatment of apoCIII with anti-apoCIII blocking antibody abolished apoCIII binding (Figure 3B). Excess nonlabeled apoCIII, but not apoCI, apoCII, or apoE (data not shown), competed for binding of FITC-labeled apoCIII with TLR2 protein (Figure 3C). Anti-TLR2 antibody inhibited the association of apoCIII with

Figure 3. Binding of apoCIII with TLR2. A, TLR2 protein or TLR4 protein were fixed on 96-well tissue culture plates (2 μg/well) and blocked, and then FITC-labeled apoCIII (100 μg/mL) was placed on it for 10 minutes. *P<0.01 vs (−) or TLR4. B, TLR2 protein was fixed on 96-well tissue culture plates (2 μg/well) and blocked, and then FITC-labeled apoCIII (100 μg/mL) was placed on it for 10 minutes. In experiments using antibodies, FITC-labeled apoCIII was pretreated with indicated antibodies (50 μg/mL) for 30 minutes. *P<0.05 vs apoCIII alone. C, TLR2 protein was fixed on 96-well tissue culture plates (2 μg/well) and blocked, and then FITC-labeled apoCIII (100 μg/mL) was placed on it for 10 minutes in the presence of an excess amount of nonlabeled apoCIII (self) (3- or 30-fold), indicated antibodies (50 μg/mL), or peptidoglycan (PGN) (10 μg/mL). *P<0.05 vs (−).
TLR2 (Figure 3C). Interestingly, peptidoglycan failed to compete with apoCIII binding to TLR2 protein (Figure 3C).

Association of ApoCIII With Human TLR2-Transfected 293 Cells

We further examined the association of apoCIII with TLR2 in situ using human TLR2-transfected 293 cells (hTLR2-293 cells). ApoCIII showed significantly higher affinity with hTLR2-293 cells compared with parental 293 cells that did not express TLR2 (Figure 4A and 4B). ApoCIII showed saturable binding with hTLR2-293 cells over 100 μg/mL (Figure 4B). In line with the results shown in Figure 3, anti-TLR2 antibody and nonlabeled apoCIII inhibited the association of apoCIII with hTLR2-293 cells (Figure 4A and 4C). Peptidoglycan did not compete with apoCIII binding, suggesting the apoCIII binding to a region of TLR2 distinct from the peptidoglycan-binding site (Figure 4C). We then tested whether overexpression of TLR2 functionally enhances the response to apoCIII treatment. Baseline levels of NF-κB activation and β1 integrin expression did not change between parental 293 cells and hTLR2-293 cells. Although apoCIII induced NF-κB activation and β1 integrin expression in 293 cells that did not express TLRs or CD14, apoCIII effects were further enhanced in hTLR2-293 cells (Figure 4D and 4E). Anti-TLR2 antibody but not anti-TLR4 antibody inhibited apoCIII-induced NF-κB activation in hTLR2-293 cells (supplemental Figure II, A). The NF-κB inhibitor SN50 but not a control scrambled peptide inhibited apoCIII-induced β1 integrin expression in hTLR2-293 cells (supplemental Figure II, B), without affecting apoCIII-binding to hTLR2-293 cells (data not shown). These results indicate that TLR2 not only increases the binding of apoCIII but also functions as the modulator of apoCIII-induced proinflammatory signal transduction, whereas some apoCIII has the ability to activate 293 cells without TLR4 or CD14.

Involvement of TLR2 in ApoCIII-Induced Mouse Peripheral Blood Monocyte Activation

We further used monocytes isolated from C57BL/6 (wild-type) mice or TLR2-deficient mice to examine the mechanism of proinflammatory action of apoCIII ex vivo. Our validation studies demonstrated that LPS activated NF-κB in TLR2-deficient mouse monocytes, as well as wild-type mouse monocytes. However, peptidoglycan failed to activate NF-κB in TLR2-deficient mouse monocytes (supplemental Figure III). Although apoCIII significantly promoted PKCα activation and NF-κB activation in wild-type mouse monocytes, their activation was not significant in TLR2-deficient...
VLDL CIII after intravenous administration of VLDL preparations. Monocytes were isolated from C57BL/6 (wild-type) mice or TLR2-deficient mouse monocytes but not TLR2-deficient mouse monocytes (Figure 6C). VLDL CIII wild-type mouse monocytes but not TLR2-deficient mouse monocytes (Figure 6B). TLR2-deficient mouse monocytes also showed less incremental adhesiveness to HUVECs induced by VLDL CIII compared with wild-type mouse monocytes (Figure 6E). Taken together, these results suggest that the TLR2 signaling pathway plays a role in the effect of apoCIII or VLDL CIII in the activation of monocytes in vivo.

**Discussion**

We recently reported that VLDL CIII activated \( \beta_1 \) integrin through PKCa in THP-1 cells and increased their adhesion to ECs under static or flow conditions. VLDL CIII also induced vascular EC activation and increased adhesion molecule expression. Interestingly, in both studies, apoCIII alone as well as VLDL CIII activated these cells, suggesting that distinct signaling pathways that do not involve apoB/E receptors mediate the direct proinflammatory and proatherogenic effects of apoCIII. The present study implicates TLR2 in apoCIII-induced monocyte activation. Blocking antibody to TLR2 or genetic inactivation of TLR2 reduced the response of human peripheral monocytes exposed to apoCIII or apoCIII-Rich VLDL. Overexpression of TLR2 in 293 cells not only increased the binding of apoCIII to the cells but also augmented apoCIII-induced NF-\( \kappa \)B activation and \( \beta_1 \) integrin expression. Thus, cells that express TLR2 or conditions that increase expression of TLR2 may exhibit enhanced response to apoCIII, although this study does not exclude a TLR2-independent process, because apoCIII activated NF-\( \kappa \)B in 293 cells that did not express TLRs. Our results indicate direct interaction of apoCIII with TLR2. Several studies have described endogenous ligands for TLRs. Recent reports suggest that endogenous unidentified ligands for TLR2 contribute to atherogenesis. The present results raise the possibility that apoCIII in apoB lipoproteins can serve as an endogenous TLR2 ligand.

MyD88 plays a crucial role in the signaling pathway downstream of the TLRs. However, recent studies reported a MyD88-independent pathway. Several studies showed the involvement of a Rac1-dependent pathway in TLR2-mediated NF-\( \kappa \)B activation of THP-1 and in 293 cells. Harokopakis et al reported that TLR2 mediated monocyte adhesion and transmigration via Rac1- and phosphatidylinositol 3-kinase–mediated signaling in response to Porphyromonas gingivalis fimbriae. In the present study, apoCIII stimulation recruited MyD88 to associate with TLR2 protein, whereas the association with Rac1 was not prominent, and apoCIII did not activate phosphatidylinositol 3-kinase (data not shown). These results suggest that apoCIII exerts proadhesive effects through a distinct signaling pathway that involves TLR2 and MyD88.

In the previous study, pertussis toxin (PTX), a specific G\( \alpha \) protein inhibitor inhibited apoCIII-induced PKCa and NF-\( \kappa \)B activation. Heterotrimeric G proteins couple with various

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**Figure 5.** TLR2 mediates apoCIII-induced mice monocyte activation. A through C, Monocytes isolated from C57BL/6J mice or TLR2-deficient mice were incubated with or without apoCIII (100 \( \mu \)g/ml) for 2 hours (A and B) or 8 hours (C). *P<0.05 vs apoCIII (-)/C57BL/6. Blots represent 5 independent experiments using apoCIII from 5 different donors that yielded similar results (A and B). IB indicates immunoblotting; mem, the membrane fraction of the cell lysates.
types of membrane receptors, and their Gα subunit mediates signal transduction, including PKC activities.21–23 We previously showed that apoCIII-rich remnant lipoproteins activated PKCα in rat smooth muscle cells and that PTX inhibited PKCα activation,15 suggesting that specific components of VLDL or VLDL remnants interact with PTX-sensitive G protein or its membrane receptors. Recent studies showed that G proteins mediate TLR signaling in several cell types,24–26 supporting our previous and present studies. Because the crosstalk between G protein coupled receptor and TLR signaling pathways exists,26 addressing the detailed role of PTX-sensitive G protein in apoCIII-induced responses requires further investigations.

ApoCIII mediates the activation of mouse monocytes triggered by VLDL CIII− and their adhesion to HUVECs, as suggested by experiments using anti-apoCIII antibody and reconstituted VLDL CIII−.9 The present study supports a pivotal role for TLR2 in these processes. Notably, apoCIII on VLDL particles correlates with a high lipid content and additional apolipoprotein content.27 Thus, these other (lipid) components that stimulate TLR2 may augment the effects of apoCIII-rich VLDL.

In conclusion, this study demonstrated that the TLR2 signaling pathway participates in the proinflammatory action of apoCIII, alone or in association with VLDL, inducing NF-κB activation and β1 integrin expression in monocytes and their adhesion to ECs. This pathway may contribute to the diverse inflammatory responses to apoCIII and the link between apoCIII levels and adverse clinical outcomes and may further support the involvement of TLR2 in atherogenesis induced by dyslipidemia. Our observations shed new light on the molecular pathways that link dyslipidemia, inflammation, atherosclerosis, and cardiovascular events.

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Disclosures
None.

References


Toll-Like Receptor 2 Mediates Apolipoprotein CIII–Induced Monocyte Activation: Retracted

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The corresponding author, Dr Akio Kawakami, admitted to the editors to improperly handling the collection and presentation of data in this article such that the authors can no longer verify the authenticity and accuracy of the data presented. These errors include, but may not be limited to, the blots in Figure 2A, Figure 4D, and Online Figure III originating from unrelated experiments of the corresponding author, and the incorrect reporting of “n” in Figures 5 and 6, which are less than indicated. As such, data in those figures are not verifiable.

All co-authors involved in this study other than the corresponding author, Dr Kawakami, had no knowledge of any scientific impropriety related to the collection, analysis, or presentation of data in this article. Dr Kawakami takes full responsibility for this.
Online Figure I

(A) THP-1 cells were pretreated with indicated antibodies (50 µg/mL), and then incubated with peptidoglycan (PGN) (10 µg/ml) or LPS (200 ng/ml) for 2 hours. (B) THP-1 cells were incubated with apoCIII (100 µg/mL) for 2 hours in the presence or absence of polymyxin B (PMB) (0.5 µg/ml). (C, D) THP-1 cells were incubated with apoCIII(100 µg/mL) for 2 hours (C) or 8 hours (D). In some experiments, apoCIII was pretreated with indicated antibodies (50 µg/mL) for 30 minutes. *p<0.05 vs. apoCIII(-), #p<0.05 vs. apoCIII alone. Blots represent 3 independent experiments using apoCIII from 3 different donors that provided similar results (B, C).
Online Figure II

(A) Cultured hTLR2-293 cells were incubated with FITC-labeled apoCIII (100 µg/mL) for 2 hours. In some experiments, the cells were pretreated with indicated antibodies (50 µg/mL) for 30 minutes. *p<0.01 vs. apoCIII(-)/antibodies(-), #p<0.05 vs. apoCIII(+)/antibodies(-). (B) Cultured hTLR2-293 cells were incubated with FITC-labeled apoCIII (100 µg/mL) for 8 hours. In some experiments, the cells were pretreated with NF-κB inhibitor SN50 or control scrambled peptide (CP) (20 µmol/mL) for 30 minutes. *p<0.01 vs. apoCIII(-)/inhibitors(-), #p<0.05 vs. apoCIII(+)/inhibitors(-). Blots represent 4 independent experiments using apoCIII from 4 different donors that provided similar results.
Monocytes isolated from indicated mice were incubated with peptidoglycan (PGN) (10 µg/ml) or LPS (200 ng/ml) for 2 hours. *p<0.01 vs. PGN(-) or LPS(-)/C57BL/6, #p<0.01 vs. LPS(-)/TLR2 deficient. Blots represent 4 independent experiments that provided similar results.