Lipid-laden macrophages are the predominant cell type in the formative stages of atherosclerosis in most animal models and humans. Lipid deposition appears as large numbers of intracellular droplets that have a foam-like appearance when paraffin-embedded tissue sections are viewed at high magnification. Consequently, the descriptive name of foam cells is applied to these lipid-laden macrophages. Originating from recruited monocytes, it has been hypothesized that foam cells remove lipoproteins that have been retained and modified in the subendothelial space. For this function to be beneficial, lipid-laden macrophages would subsequently egress from the area of the forming lesion. However, the system frequently goes awry. Macrophages recruited to the arterial wall become grossly engorged with lipid, presumably because of an imbalance in lipid metabolism. In this greatly hypertrophied state, macrophages are unable to transit through the endothelium and be transferred back to the blood compartment. Therefore, instead of exiting the artery, these cells are retained and accumulate. In addition to these cells forming the mass of the evolving lesions, there is also the potential for secretion of many bioactive molecules that may perpetuate and modify the atherogenic process.

There have been many approaches to modify the development of foam cells by manipulating intracellular transport, intracellular storage, or efflux of lipids. The transport of extracellular lipid to form intracellular droplets is presumed to occur via endocytosis through lipoprotein receptors that are not downregulated by increased cholesterol content. There are many classes of scavenger receptors that transport modified lipoproteins into macrophages, of which the most intensely studied has been class A scavenger receptor (SR-A) and CD36. However, genetic manipulation of SR-A and CD36 has generated inconsistent findings for effects on atherosclerosis. Once inside the cell, lipoprotein-derived cholesterol ester is cleaved in lysosomes by an acidic cholesterol ester hydrolase. Unesterified cholesterol is transported to the cytosol for reesterification by acyl-coenzyme A:cholesterol acyltransferase (ACAT) to generate lipid droplets that are protein-coated. Once stored in lipid droplets, neutral cholesterol hydrolase can convert the core content back to unesterified cholesterol. Unesterified cholesterol may also partition to the plasma membrane and transfer to extracellular acceptors. Several pathways have been proposed for cholesterol efflux, including the ABC transporters and SR-B1. Therefore, cholesterol homeostasis in macrophages has many levels of regulation involved in their conversion to foam cells (Figure).

In this issue of Circulation Research, Paul et al9 have studied the role of adipose differentiation–related protein (also known as adipophilin, ADRP, or ADFP) on macrophage foam cell formation and atherosclerosis. ADFP, as it is referred to in this article, is a member of the PAT domain family of proteins that are targeted to the cytosol and are involved in cholesterol and triglyceride content and resistance to diet-induced fatty liver. However, they have no difference in body weight, plasma triglyceride and cholesterol concentrations, fat mass, or adipocyte differentiation.

Although mice deficient in ADFP have a mild phenotype, the expression of this protein alters lipid metabolism in cultured macrophages. Overexpression of ADFP increased the storage of triglycerides and cholesterol following incubation with acetylated LDL (AcLDL) in THP-1 cells, whereas depletion of the protein using small interfering RNAs reduced lipid accumulation. This increase occurred without affecting transport of AcLDL particles or regulating many proteins involved in cholesterol efflux. Although these cell culture studies with manipulated ADFP expression portend a protection against atherosclerosis, the field has been led astray in the past by such expectations. Perhaps one of the best examples of this involves studies on ACAT. The development of many pharmacological inhibitors of this enzyme demonstrated that they decreased lipid deposition in macrophages. In contrast, repopulation of irradiated LDL receptor–deficient mice with bone marrow–derived stem cells from ACAT1-deficient mice led to a dramatic increase in macrophage lipid deposition in atherosclerotic lesions. Conversely, macrophage-specific overexpression of neutral cholesterol hydrolase reduced intracellular lipid deposition and atherosclerotic lesion formation. Because the net effect of ACAT inhibition and excess neutral cholesterol hydrolase should both increase unesterified cholesterol, it is a quandary that they produce such divergent results. Despite the several decades of research, it is clear that we have not resolved all the mechanisms of macrophage lipid engorgement and its effects on atherosclerosis.

Therefore, although the cell culture studies on ADFP may be consistent with an effect in reducing atherosclerosis, this needs to be tested experimentally in vivo. In this impressive
ADFP is upregulated in atherosclerotic tissue from apoE study performed by Paul et al, the authors have demonstrated in vitro and in vivo. The effects of macrophage-specific regulation of receptors, enzymes, and transporters on lipoprotein–cholesterol ester trafficking are denoted as: absence decreases atherosclerosis; absence increases atherosclerosis; and overexpression decreases atherosclerosis; and absence has inconsistent effects on atherosclerosis.

ADFP deficiency in macrophages reduces atherosclerosis through a mechanism that is solely based on inhibition of lipid deposition rather than secondary consequences on inflammatory processes.

Although macrophages sequester cholesterol esters in membrane limited droplets associated with PAT domain proteins, macrophages store intracellular lipids in diverse structures within evolving atherosclerotic lesions. In addition to the lipid droplets described in this study, there are also complex lipid structures that are encased by acid phosphatase positive organelles. These structures are likely to arise from the phagocytic engulfment of modified lipoproteins by macrophages coupled with an inability of the cholesterol esters to be hydrolyzed and exit this domain. The presence of excessive lipid storage in acid phosphatase organelles versus lipid droplet changes as a function of time and complexity of atherosclerotic lesions. Presently, it is unclear what the consequences are of excessive storage in these 2 venues. In addition, the relative ability to remove intracellular lipid located in these 2 regions has not been defined. Therefore, the availability of mice that can change the distribution of these lipid stores will provide important insight into the progression of lesions into advanced stages. Furthermore, it would be of interest to determine whether the efficacy of ADFP inhibition on reducing the initiation of atherosclerosis would be matched by inhibiting this protein in established atherosclerotic lesions.

Although there are many factors that modulate atherogenesis, regulation of lipid metabolism is still the mainstay of targets for atherosclerosis therapies. This publication by Paul et al invokes ADFP as a new target to reduce macrophage lipid deposition as an approach to decreasing atherosclerosis.

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References

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